

# Magnetic Nanomaterials.

Some Biomedical Applications: Magnetofection, Magnetic Hyperthermia, and Ferrogels for Drug Delivery

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*Physics Building, 1905*



Using magnetization of magnetic phase (NPs)  $M_p$

$$M_p = \gamma^3 M$$

hence

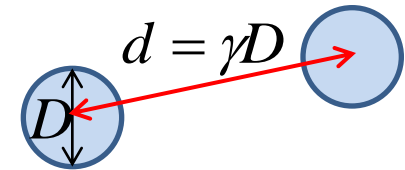
$$H_i \approx \frac{\kappa\beta\lambda_i^s}{\gamma^3} M_p = -N_{Def} M_p$$

Determined by shape and  
NP dilution

$$N_{Def} = \frac{N_D}{\gamma^3}$$

Determined by shape

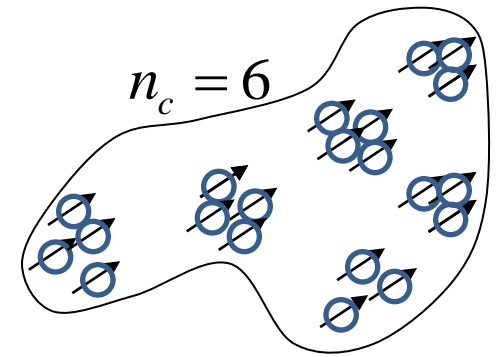
$$\text{if } \gamma > 3.16 \Rightarrow N_{Def} < 0.1N_D$$



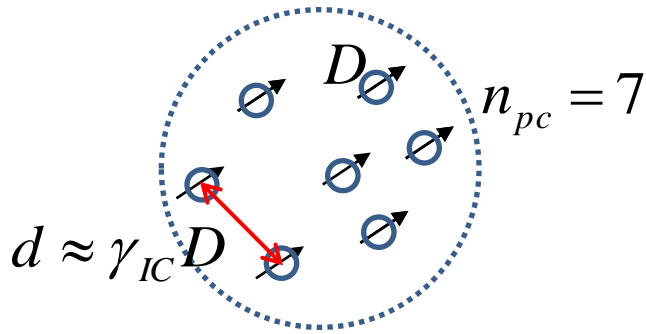
$$\text{or } N_{Def} = \left(\frac{D}{d}\right)^3 N_D$$

# Case of interest

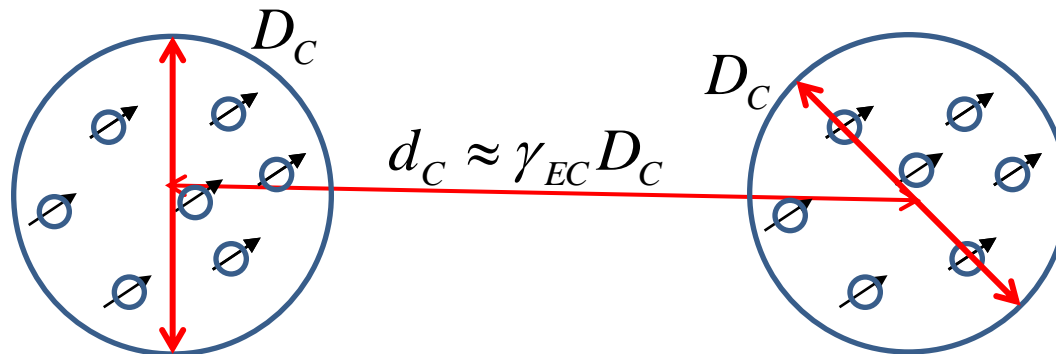
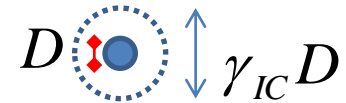
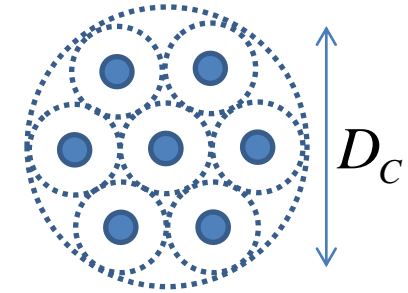
NPs are aggregated in clusters, and clusters interact among them.  $N_D$  is the demagnetizing factor which corresponds both to sample shape and to cluster average shape.



NP number  $n_p = n_c n_{pc}$



$$D_C^3 = n_{pc} \gamma_{IC}^3 D^3$$



Leads to

$$N_{Def} \approx \frac{N_D^{cluster}}{\gamma_{IC}^3} + \frac{N_D^{sample} - N_D^{cluster}}{\gamma_{IC}^3 \gamma_{EC}^3}$$

This is the sought result, it gives the dependency of the effective demagnetizing factor in terms of cluster and sample ones, and the characteristic distances (dilution factors) of the problem.

# Analysis with Langevin model.

## Comparison with Allia's proposition

$$\vec{H} = \vec{H}^{ap} + \vec{H}^{dip}$$

$$M_p = M_{pS} L\left(\frac{\mu_0 \vec{\mu} \cdot \vec{H}}{kT}\right) = M_{pS} L\left(\frac{\mu_0 \mu (H^{ap} + H^{dip})}{kT}\right)$$

$$\vec{H}^{dip} = -N_{Def} \vec{M}_p$$

Assumptions  $\vec{M}_p \uparrow\uparrow \vec{H} \uparrow\downarrow \vec{H}^{dip}$

$$M_p(H, T) = M_{pS} L\left(\frac{\mu_0 \mu (H^{ap} - N_{Def} M_p)}{kT}\right) = \frac{\mu}{V_p} L\left(\frac{\mu_0 \mu (H^{ap} - N_{Def} M_p)}{kT}\right)$$

When Langevin function argument is  $\ll 1$

$$M_p = \frac{\mu_0 \mu^2 (H^{ap} - N_{Def} M_p)}{3kTV_p}$$

$$M_p = \frac{H^{ap}}{\frac{3kTV_p}{\mu_0 \mu^2} + N_{Def}}$$

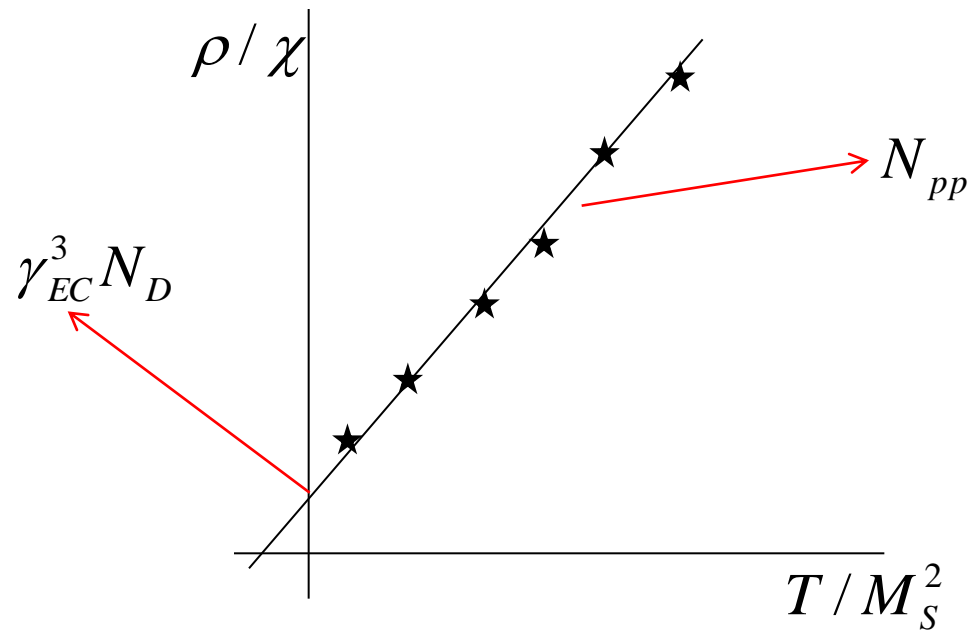
$$\frac{1}{\chi_p} = \frac{3kT}{\mu_0 M_{pS}^2 V_p} + N_{Def}$$

$$\frac{1}{\chi_p} = \frac{3kTN_p}{\mu_0 M_{pS}^2} + N_{Def}$$

$$N_p = 1/V_p$$

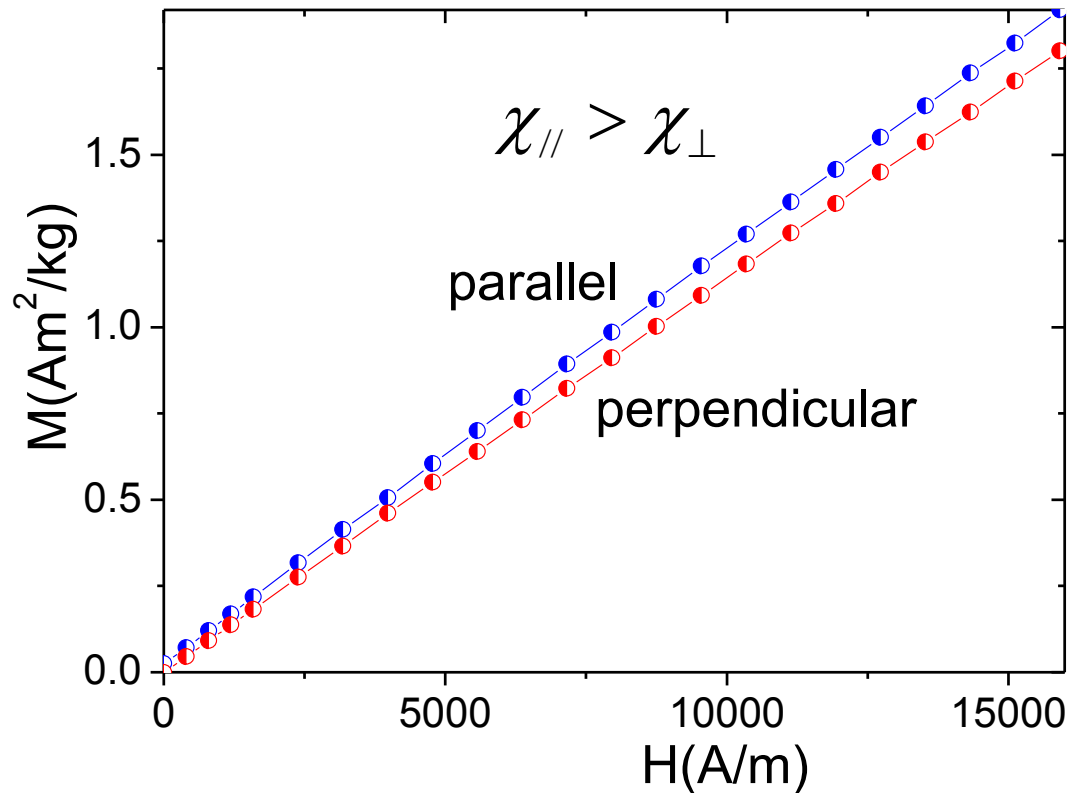
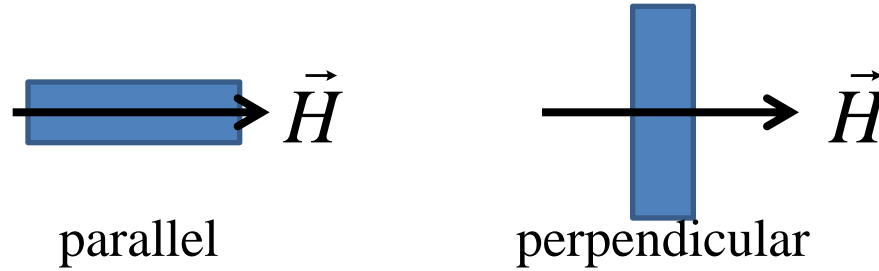
 **Measured** susceptibility of magnetic phase.

$$\frac{\rho}{\chi} = \frac{3kN_{pp}}{\mu_0} \left( \frac{T}{M_S^2} \right) + \gamma_{EC}^3 N_D$$



# Shape effects

Ferrogel PVA/maghemite 15.7% mass concentration





$$\chi_{//} \approx 1.13 \times 10^3 \text{ m}^3 / \text{kg}$$

$$\chi_{\perp} \approx 1.08 \times 10^3 \text{ m}^3 / \text{kg}$$

$$N_{Def \perp} - N_{Def //} \approx \frac{10^3}{4\pi\rho} \left( \frac{1}{\chi_{\perp}} - \frac{1}{\chi_{//}} \right)$$

Assuming a uniform distribution of NPs

$$\gamma^3 \approx \frac{N_{D\perp} - N_{D//}}{N_{Def \perp} - N_{Def //}}$$

De la forma de la muestra

## Demagnetizing factors for rectangular ferromagnetic prisms

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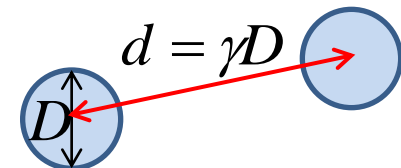
15 MARCH 1998

$$N_{//} = 0.06$$

$$N_{\perp} = 0.80$$



$$\gamma \approx 2.1$$

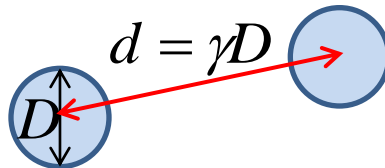


Estimation of  $\gamma$  from FG density and mass concentration of Fe oxide  $x$ , assuming a uniform distribution of NPs

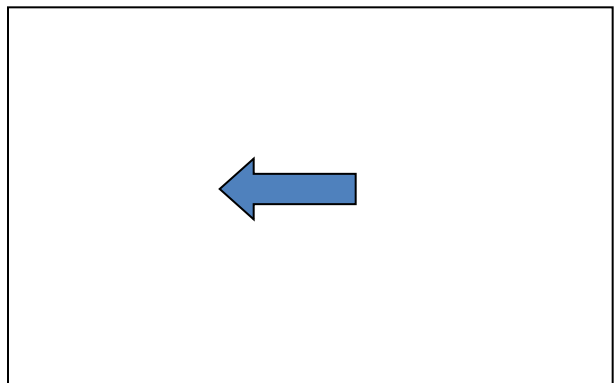
$$x = \frac{m_{oFe}}{m_{FG}} \qquad \rho_{FG} = \frac{m_{FG}}{V_{FG}}$$

For FG 10\_1

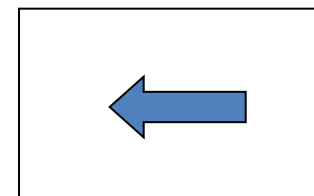
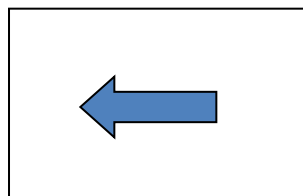
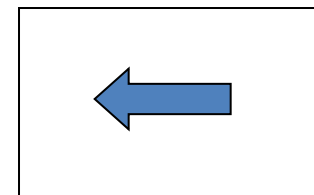
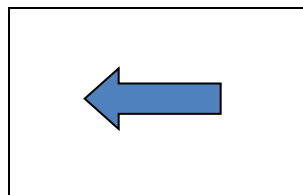
$$\text{using } \begin{cases} \rho \approx 5.18 \text{ g / cm}^3 \\ x = 0.157 \\ \rho_{FG} \approx 1.1 \text{ g / cm}^3 \end{cases} \Rightarrow \gamma \approx 2.5$$



$$\vec{\nabla} \cdot \vec{M} = 0$$

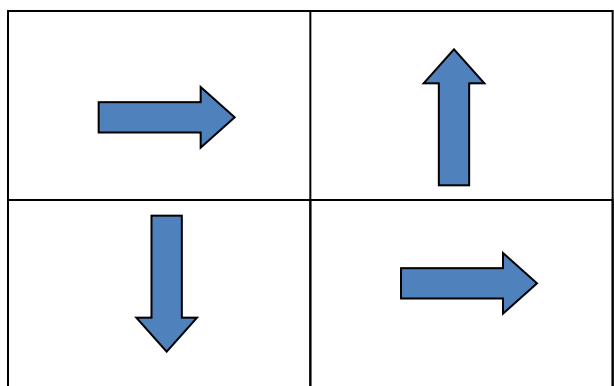


$$\vec{\nabla} \cdot \vec{M} \neq 0$$

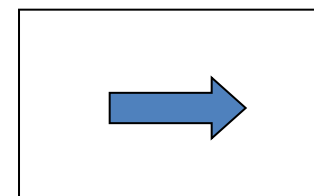
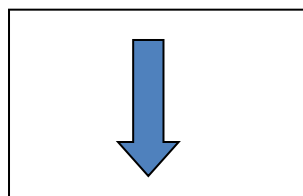
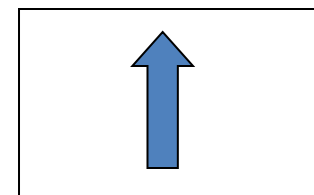
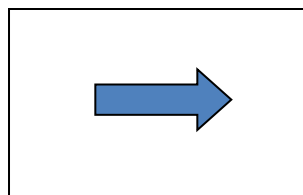


$$\vec{H} = -N_D \vec{M}$$

$$\vec{\nabla} \cdot \vec{M} \neq 0$$



$$\vec{\nabla} \cdot \vec{M} \neq 0$$



## **Class 2a**

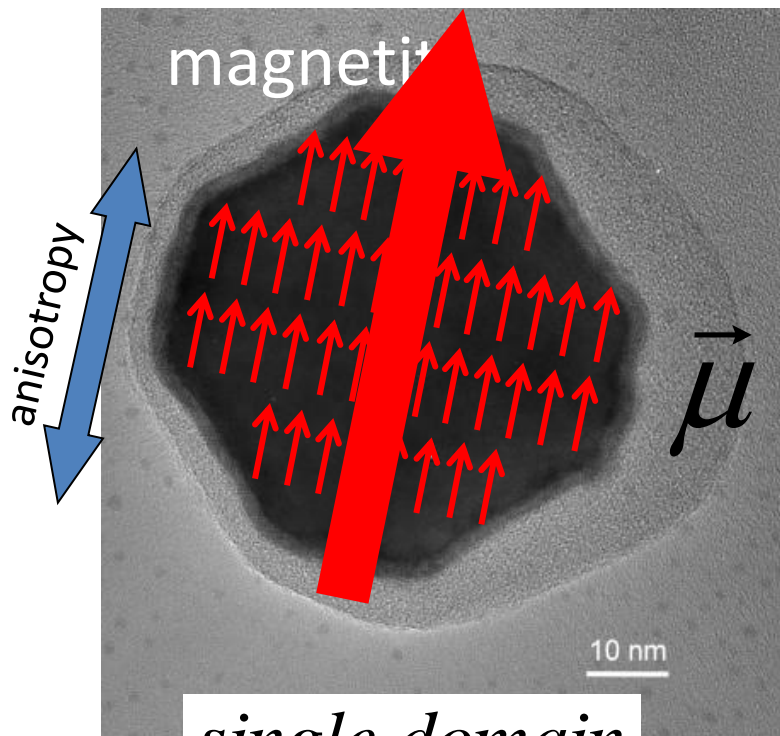
Understanding SAR and searching for high performance nanomaterials  
zinc-doped magnetite nanoparticles and ferrofluids for hyperthermia  
applications

Citric Acid Coated Magnetite Nanoparticles for Magnetic Hyperthermia  
In vitro experiments

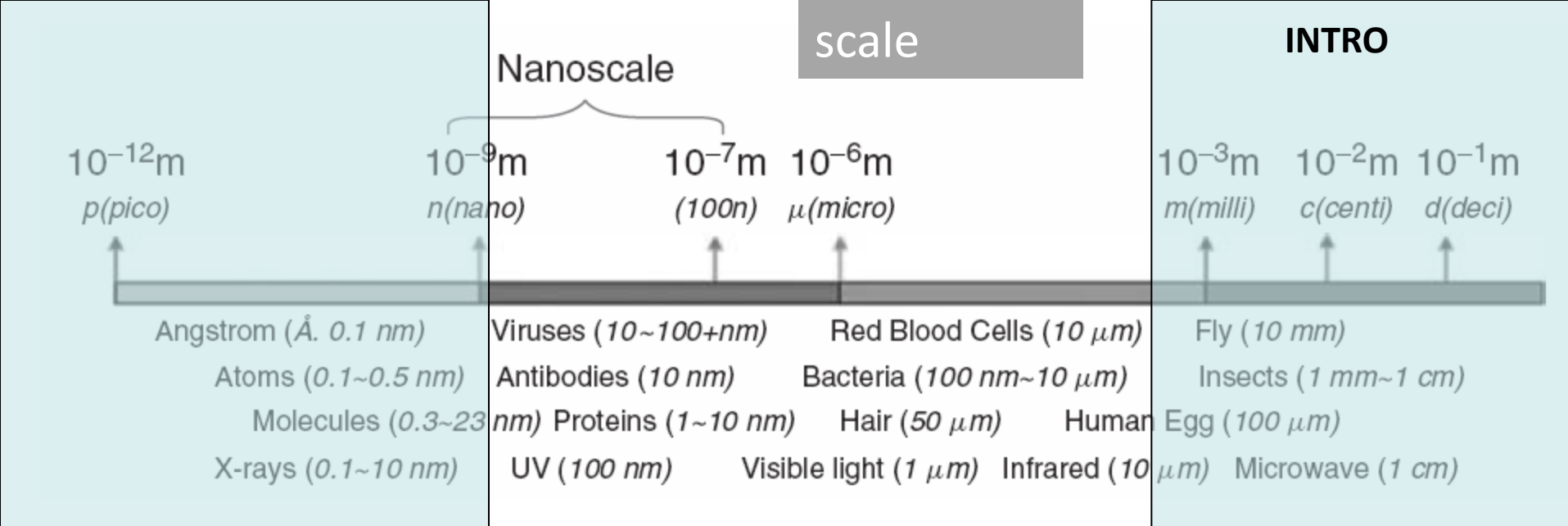
# Magnetic nanomaterials



*small magnet*



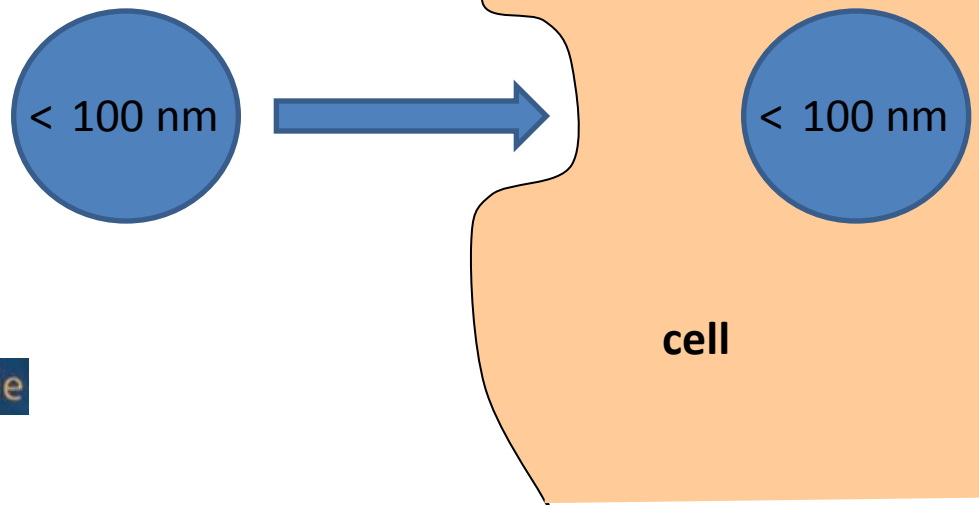
*single domain*



# NANOMEDICINE

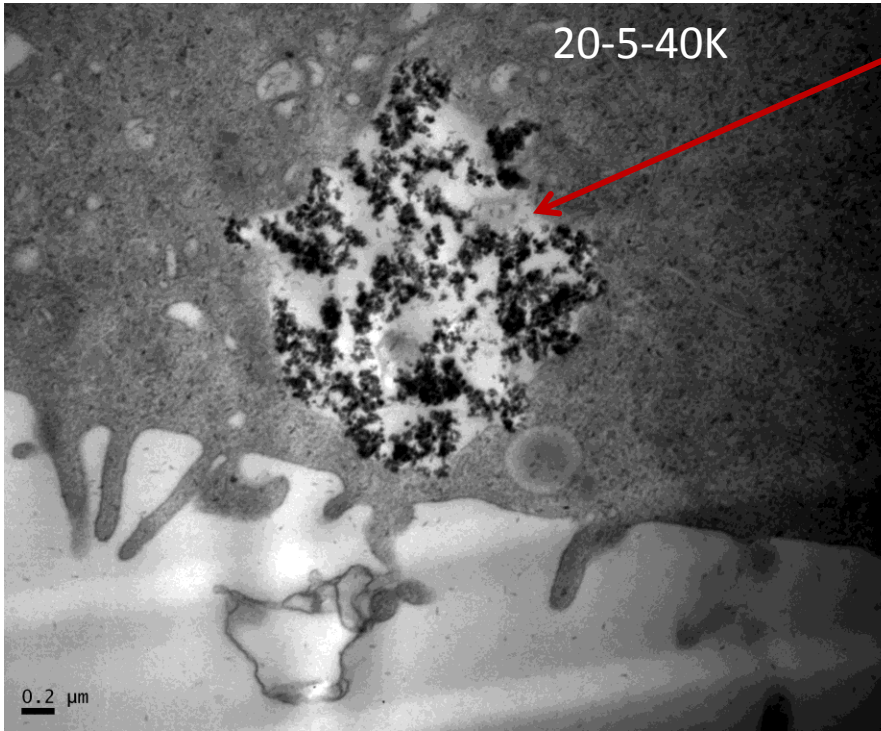
DESIGN AND APPLICATIONS OF  
MAGNETIC NANOMATERIALS,  
NANOSENSORS AND NANOSYSTEMS

Vijay K. Varadan, Linfeng Chen, Jining Xie



# endocytosis

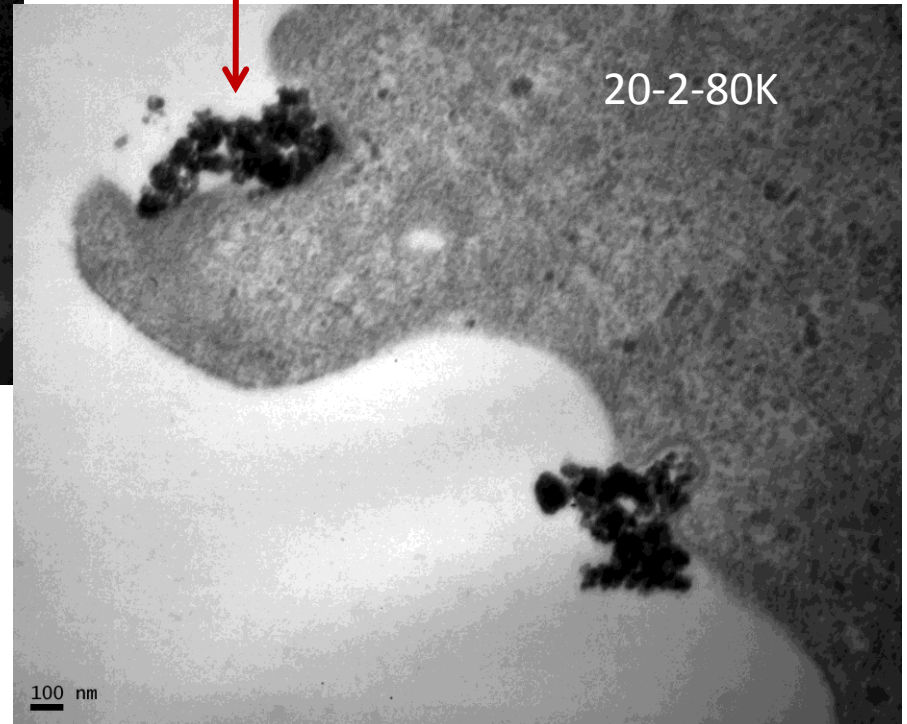
20-5-40K



Cell endosome  
with MNP

endocytosis

20-2-80K



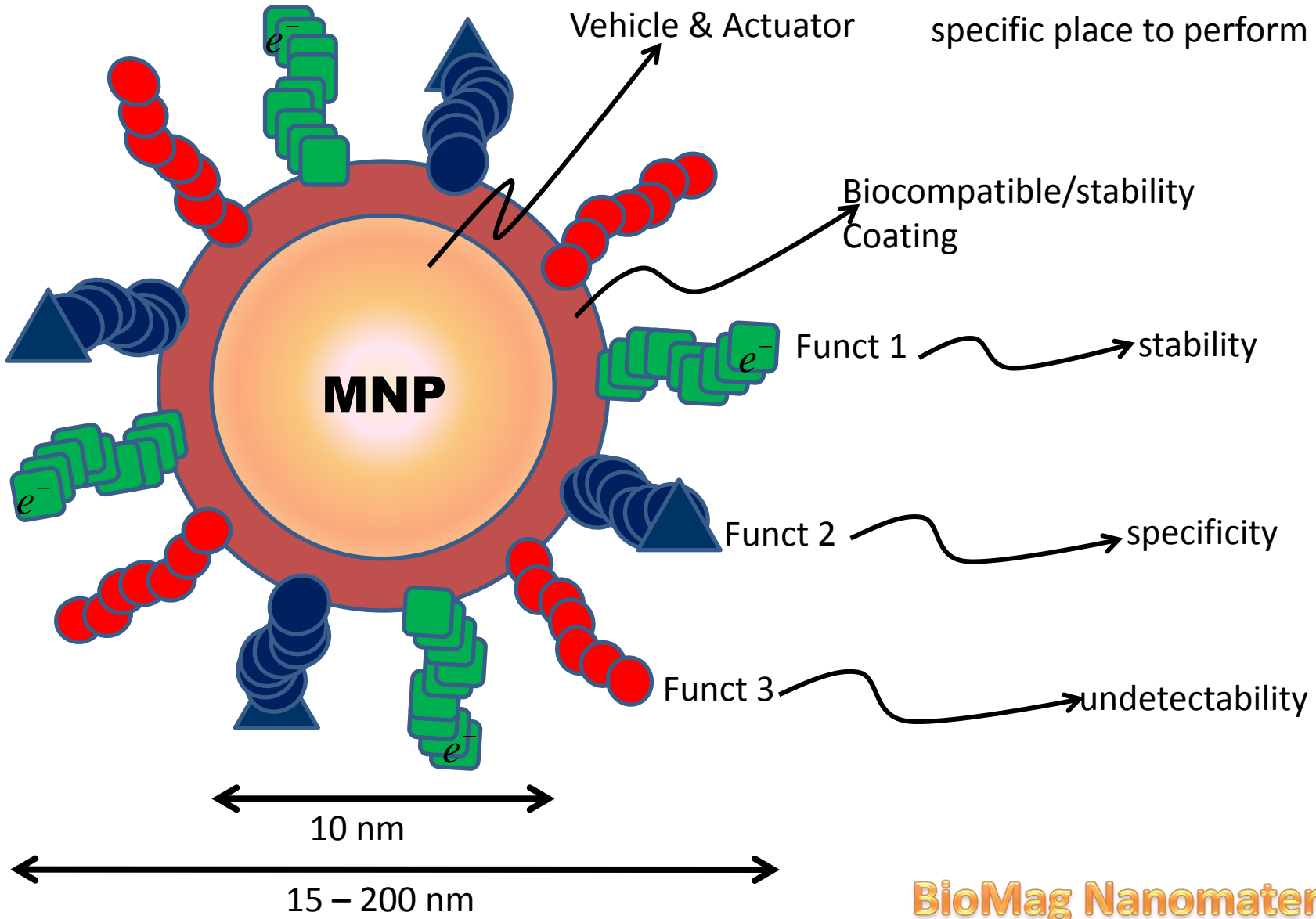
A549  
 $\text{Fe}_3\text{O}_4$   
10 nm  
20  $\mu\text{g}/\text{ml}$

# **Magnetic nanomaterials**



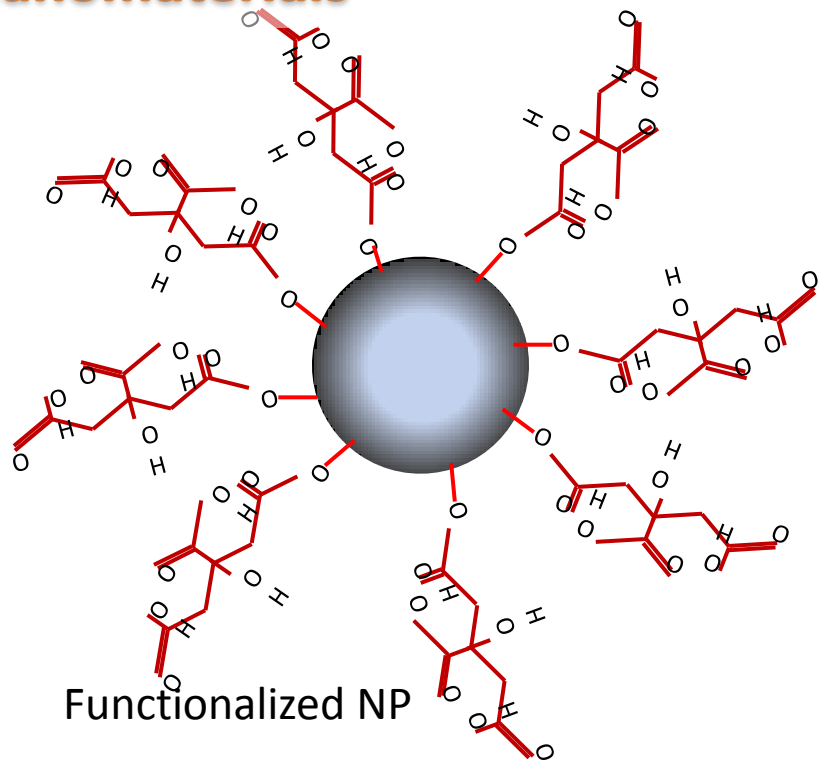
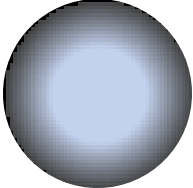
# Magnetic nanomaterials

MNP can be manipulated with static and dynamic magnetic fields and delivered to a specific place to perform a task

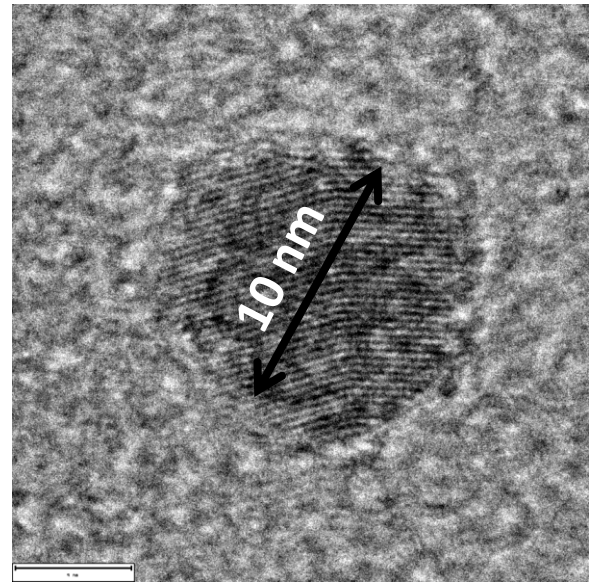


# Magnetic nanomaterials

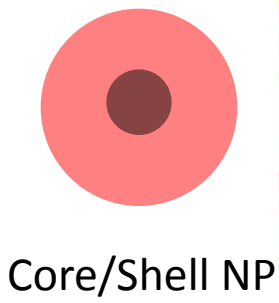
naked NP



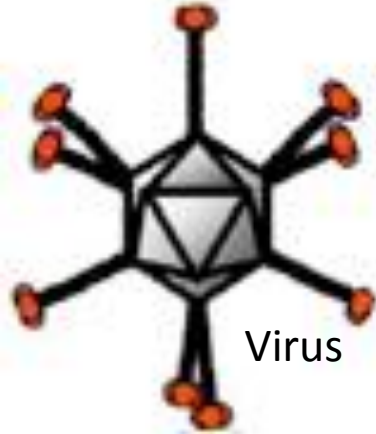
Functionalized NP



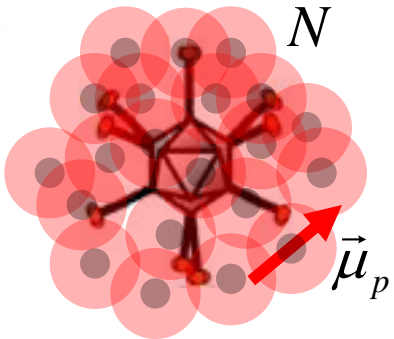
Citric Acid coated Magnetite



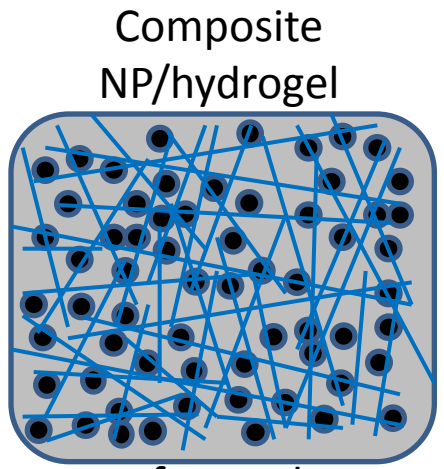
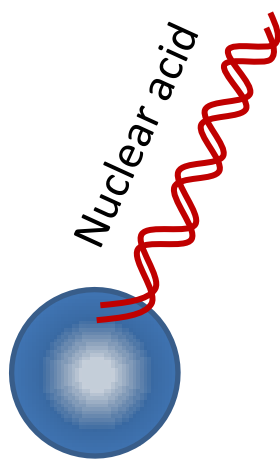
Core/Shell NP



Virus



Complex NP/Virus



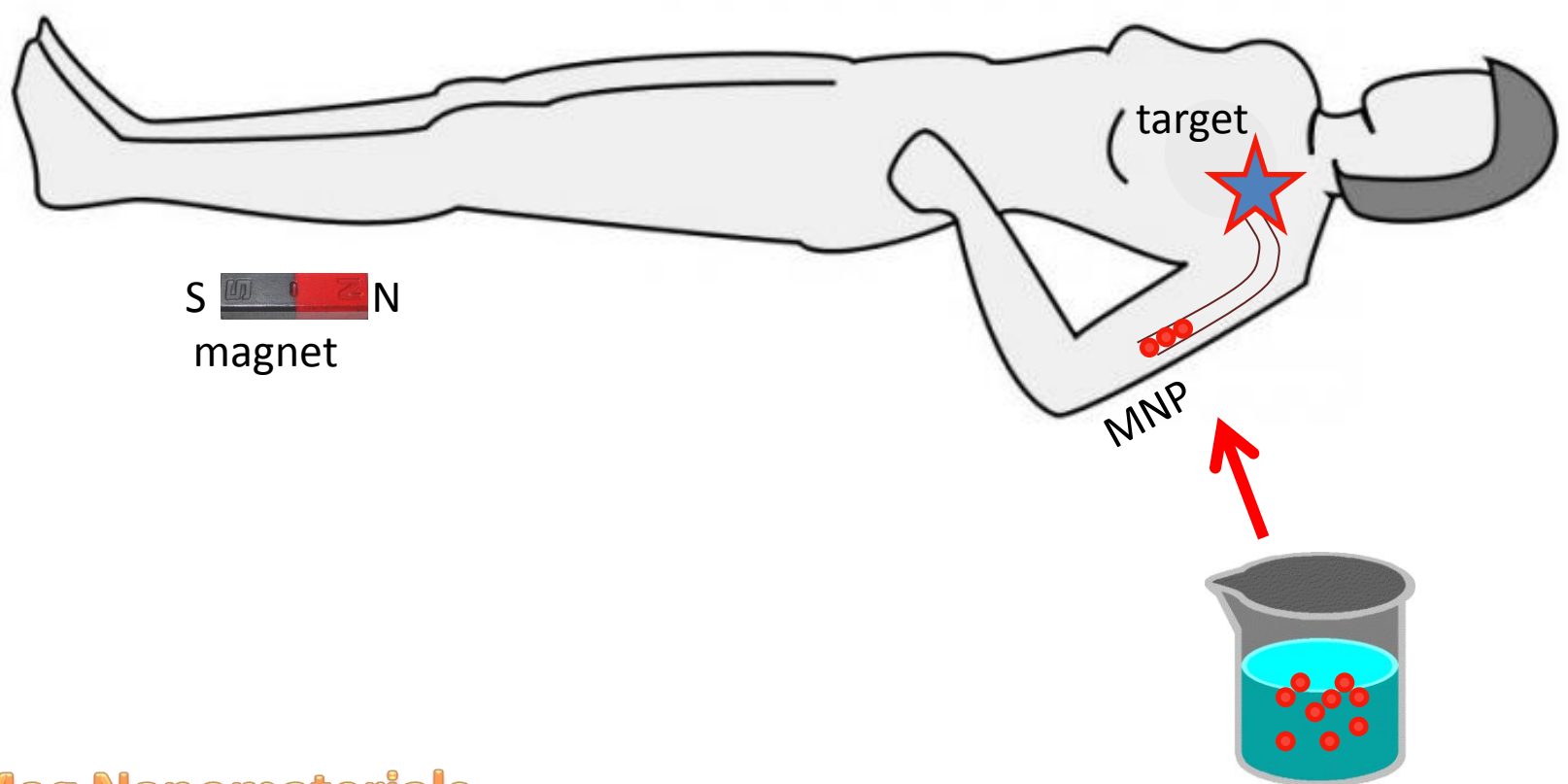
Composite NP/hydrogel

ferrogel

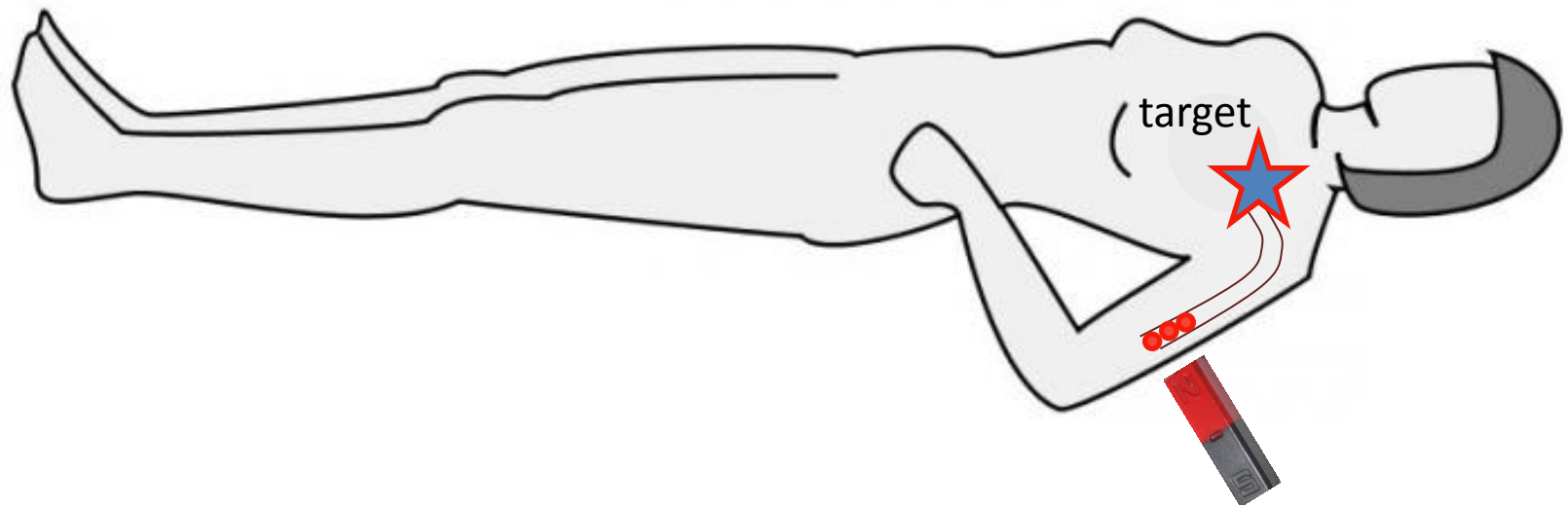
# BioMag Nanomaterials

# **Some uses of Magnetic nanomaterials**

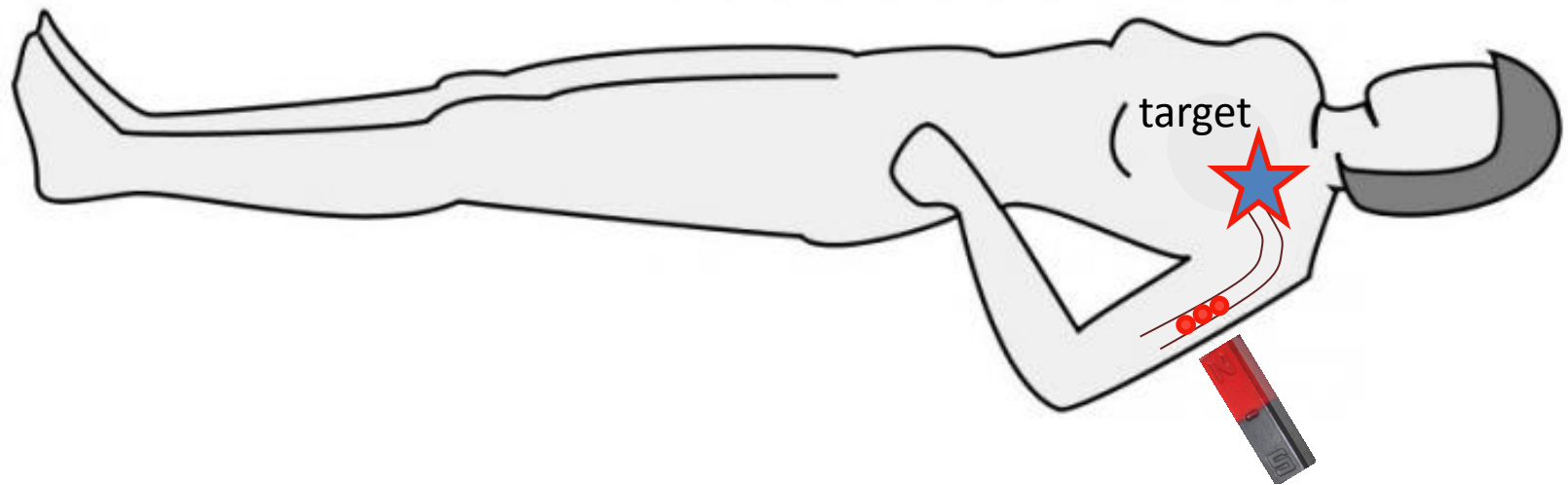
# Magnetic separation, magnetic delivery



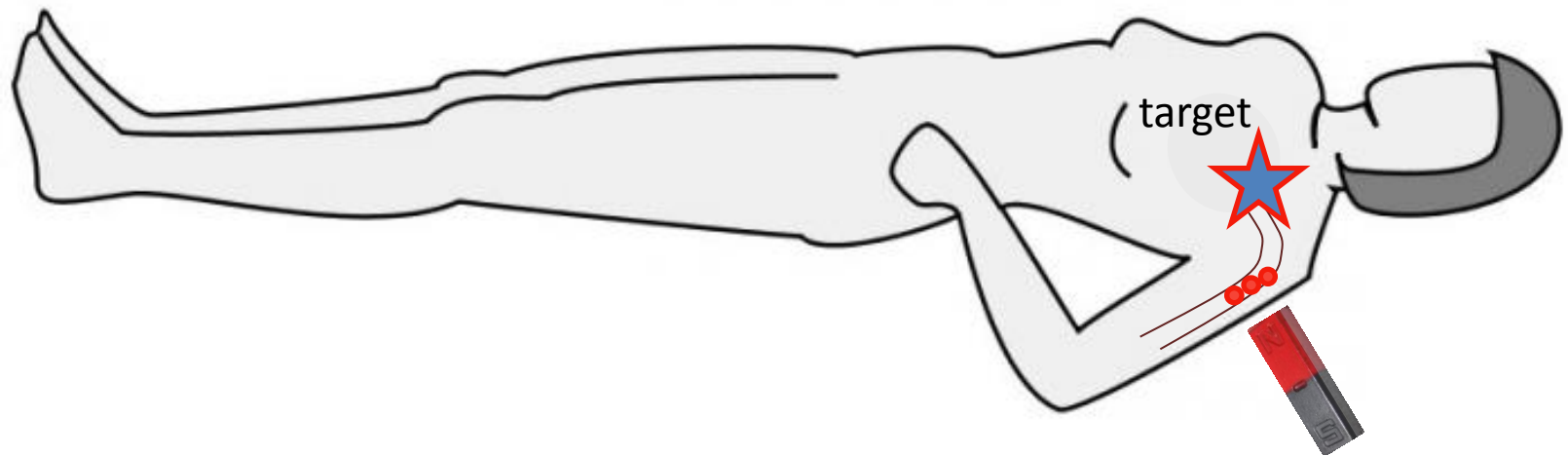
# Magnetic separation, magnetic delivery



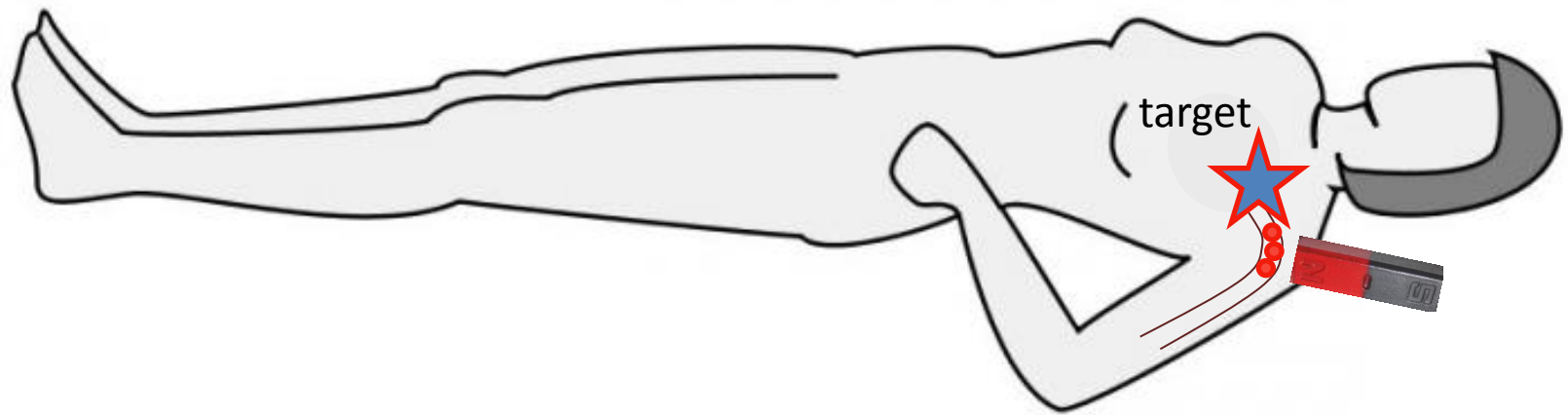
# Magnetic separation, magnetic delivery



# Magnetic separation, magnetic delivery

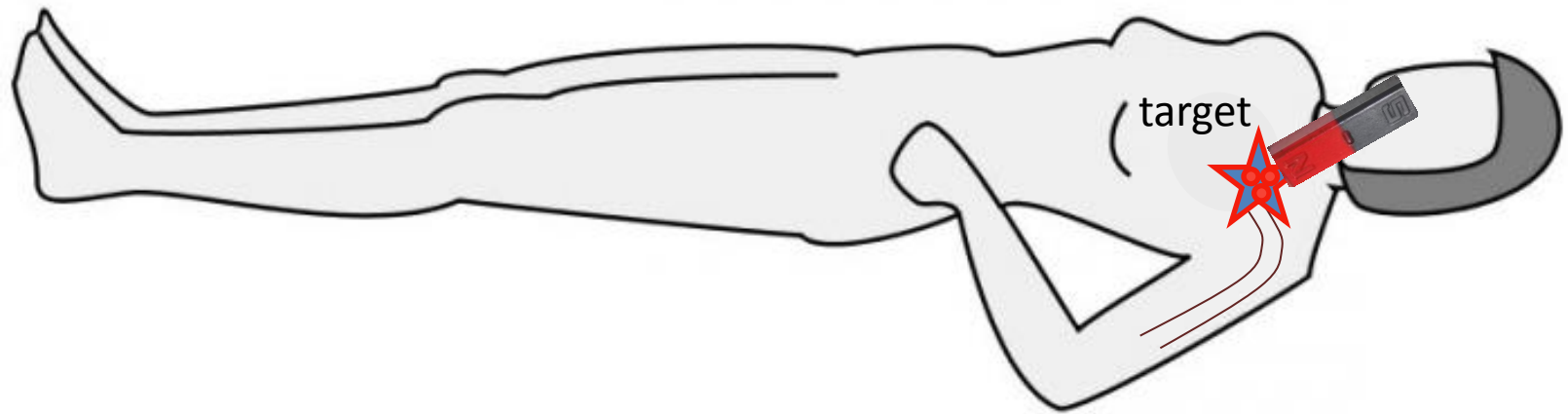


# Magnetic separation, magnetic delivery

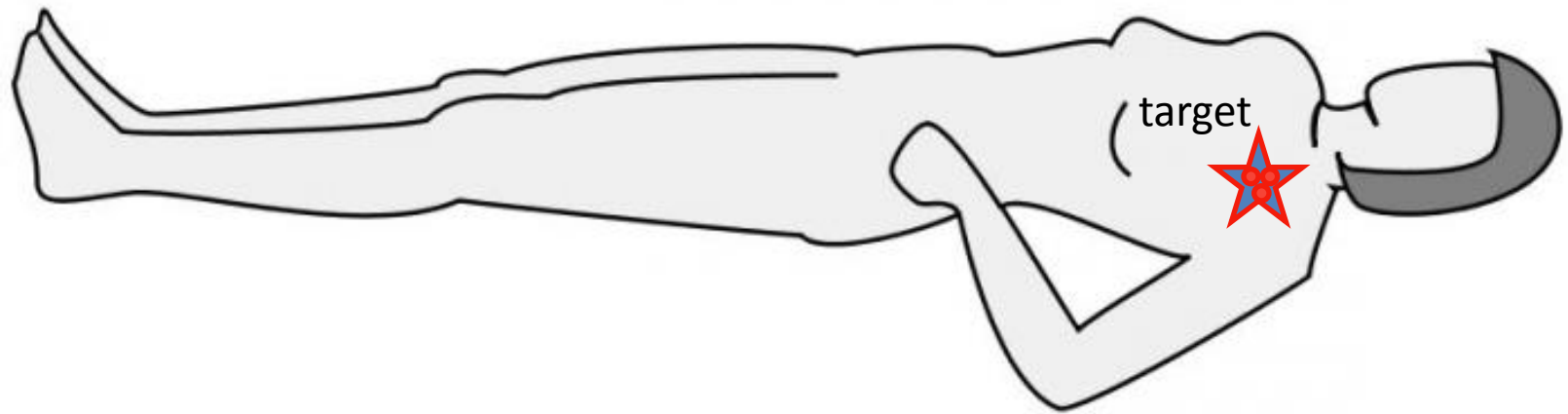




# Magnetic separation, magnetic delivery

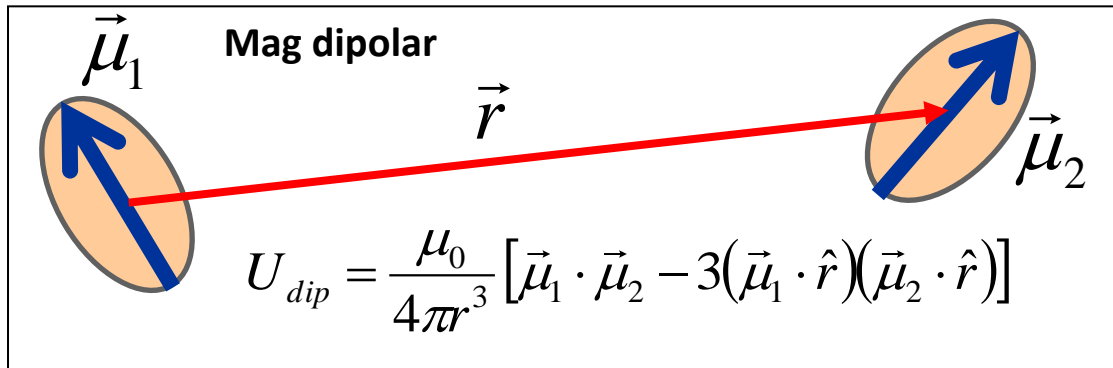
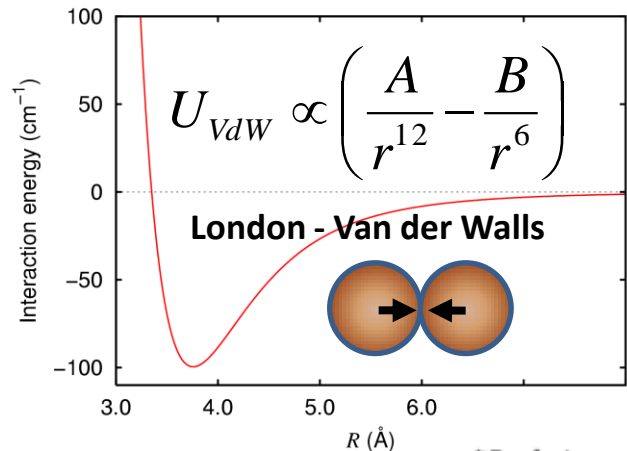


# Magnetic separation, magnetic delivery



# FORCES

# Interactions among MNP

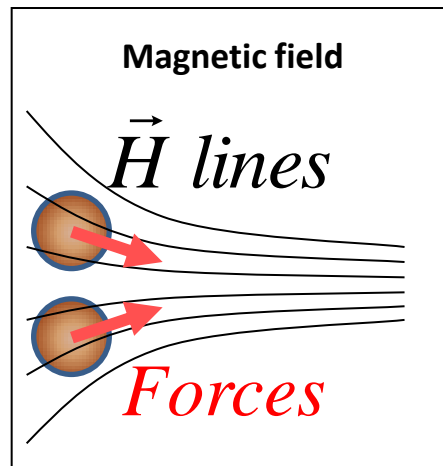
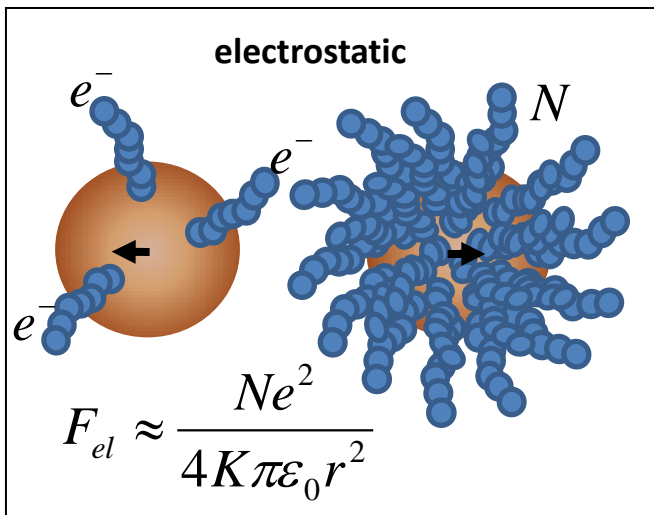


Hamaker

$D$   $d$

$E_1(x) = \frac{1}{12} \left\{ \frac{1}{x^2 + 2x} + \frac{1}{x^2 + 2x + 1} + 2 \ln \frac{x^2 + 2x}{x^2 + 2x + 1} \right\}_{x=d/2D}$

The diagram shows two blue spheres of diameter  $D$  separated by a distance  $d$ . The Hamaker equation is given below.



$F_g = \frac{(\rho - \rho_s)\pi d_p^3 g}{6}$

gravitational

The diagram shows a brown sphere with a downward-pointing arrow representing the gravitational force.

viscous

$\vec{F}_\eta = 6\pi\eta R_v \vec{v}$

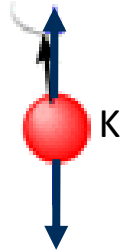
The diagram shows a brown sphere with a black arrow labeled  $\vec{v}$  pointing to the right, representing the velocity. A larger black arrow labeled  $\vec{F}_\eta$  also points to the right, representing the viscous force.

interaction	value	comment
Dipolar magnetic	5e-3 nN	10 nm NP ~ in contact
Van der Waals - London	4e-1nN	10 nm NP ~ in contact
Gravitational	2e-11 nN	10 nm NP ~ in contact
Electrostatic	2e-3 to 5e3 nN	10 nm NP ~ in contact
viscous	6e-4 to 3e-2 nN	1cm/s < $B_v$ < 100 cm/s
Magnetic field	2e-9 to 2e-7 nN	1 T <sup>2</sup> /m < $G_B$ < 100 T <sup>2</sup> /m**

\*\*1e5 T<sup>2</sup>/m would be required to compensate 1 cm/s viscous flow

# Single Domain Magnetic Nanoparticle Relaxation Mechanisms

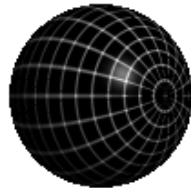
*fixed particle*  
*Néel :  
moment oscillation*



$$\tau_N = \tau_0 \exp\left(\frac{KV}{kT}\right)$$

*anisotropy constant*  
*particle volume*

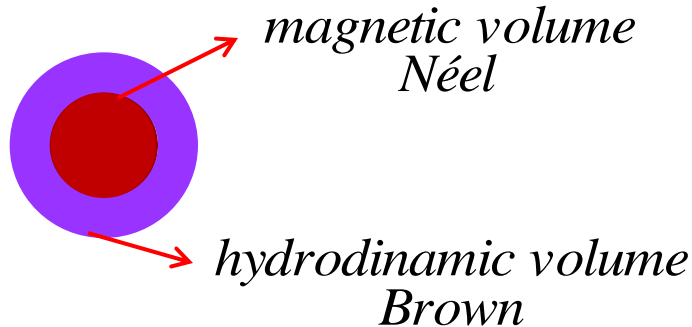
*viscous media*  
*Brown :  
particle rotation*



$$\tau_B = \frac{3\eta V}{kT}$$

*viscosity*  
*particle volume*

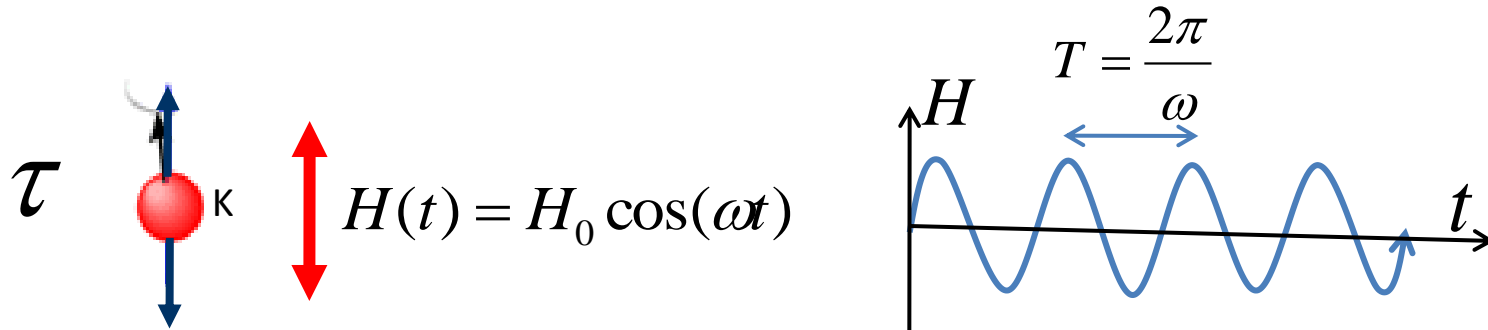
*volume?*



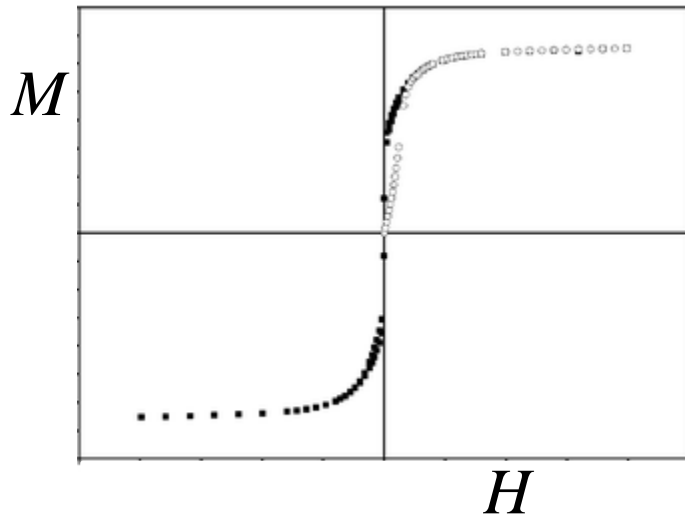
$$\frac{1}{\tau} = \frac{1}{\tau_B} + \frac{1}{\tau_N}$$

## SAR

MNP specific energy absorption rate from RF field



Linear response theory



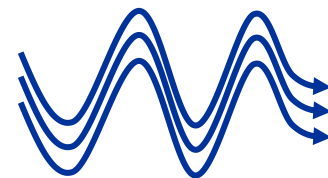
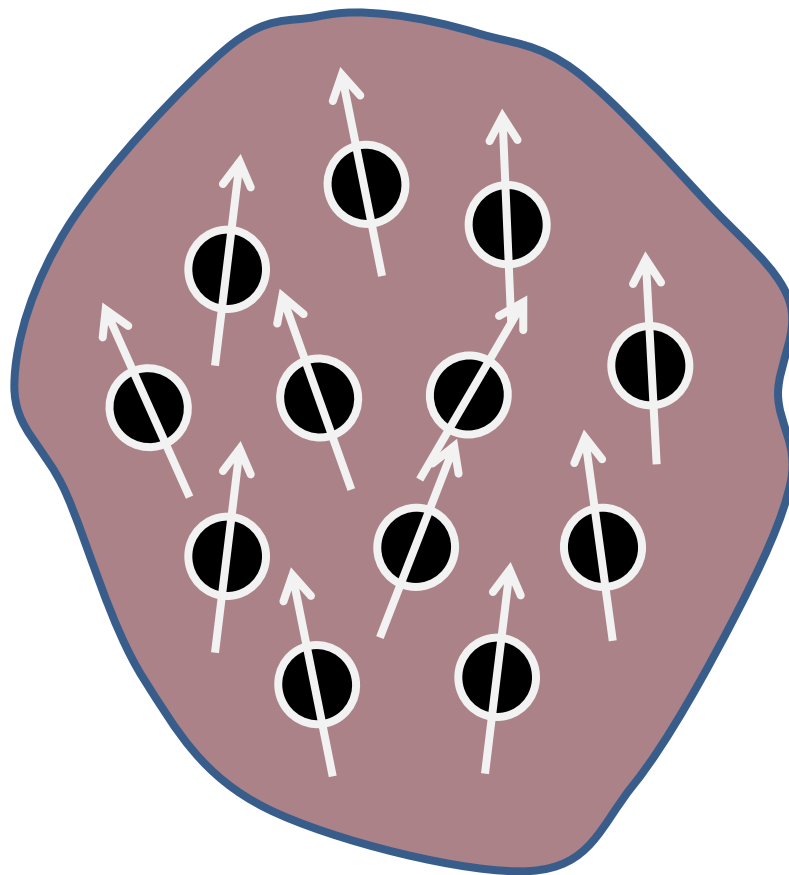
$$SAR = f \varepsilon_{cycle} = -f \mu_0 \oint M dH = \mu_0 \pi \chi''_{MNP} f H_0^2$$

$$SAR = \frac{\pi \mu_0 H_0^2 f}{\rho} \frac{\mu_0 M_S^2 V \rho^2}{3kT} \frac{\omega \tau}{1 + (\omega \tau)^2}$$

$$\tau = 1 / \omega$$

energy  
absorption from  
a RF field  
(SAR)

$$\tau \approx 1/\omega$$



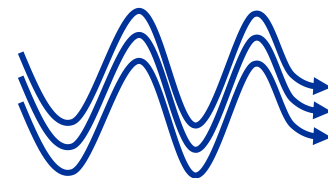
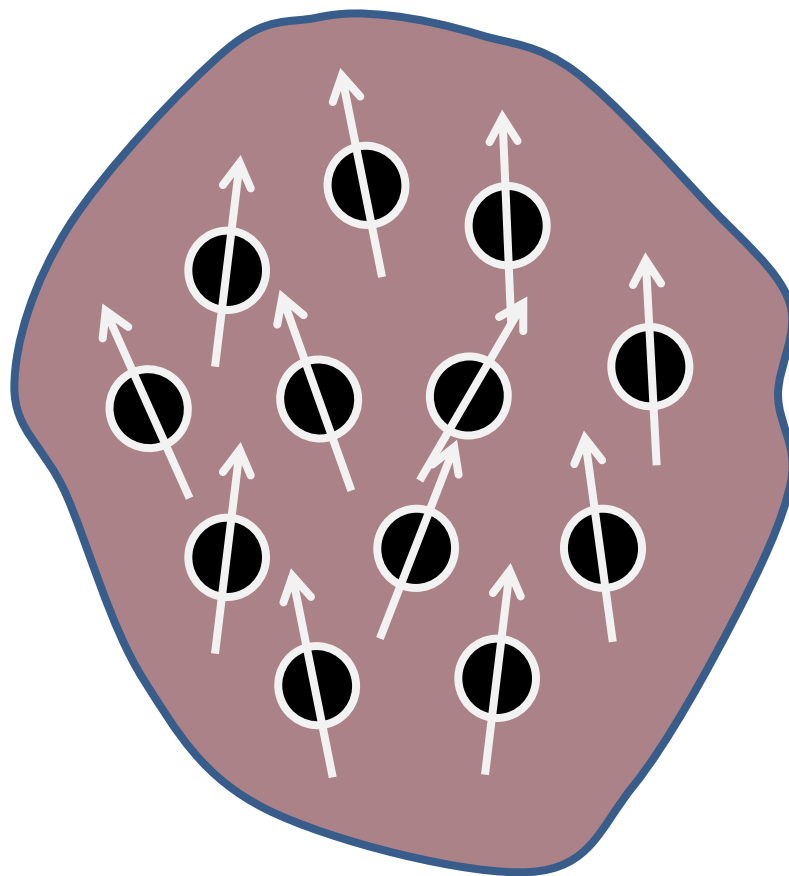
**T**





energy  
absorption from  
a RF field  
(SAR)

$$\tau \approx 1/\omega$$



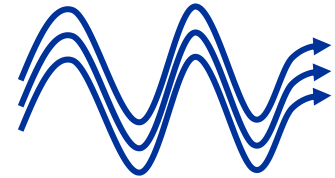
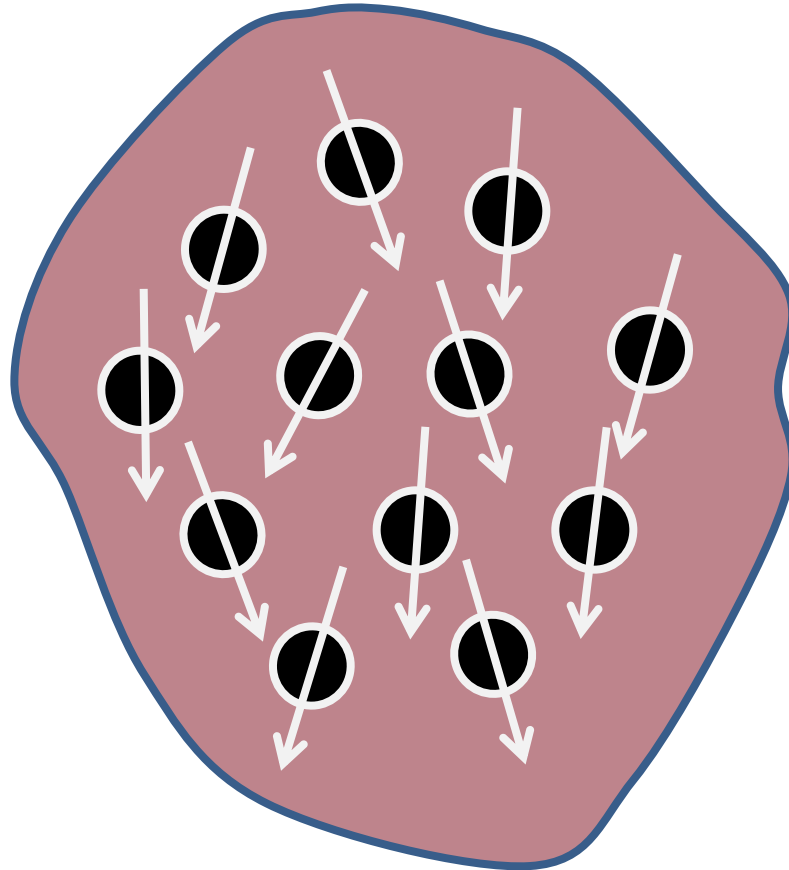
**T**



energy  
absorption from  
a RF field  
(SAR)

$$\tau \approx 1/\omega$$

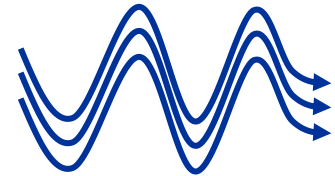
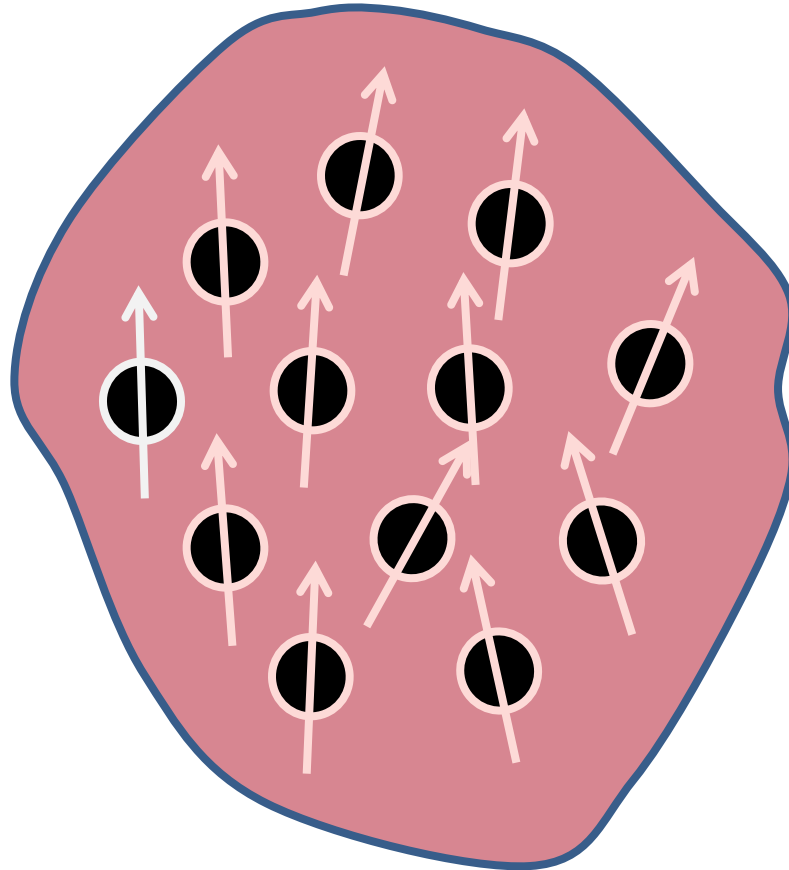
**T**



energy  
absorption from  
a RF field  
(SAR)

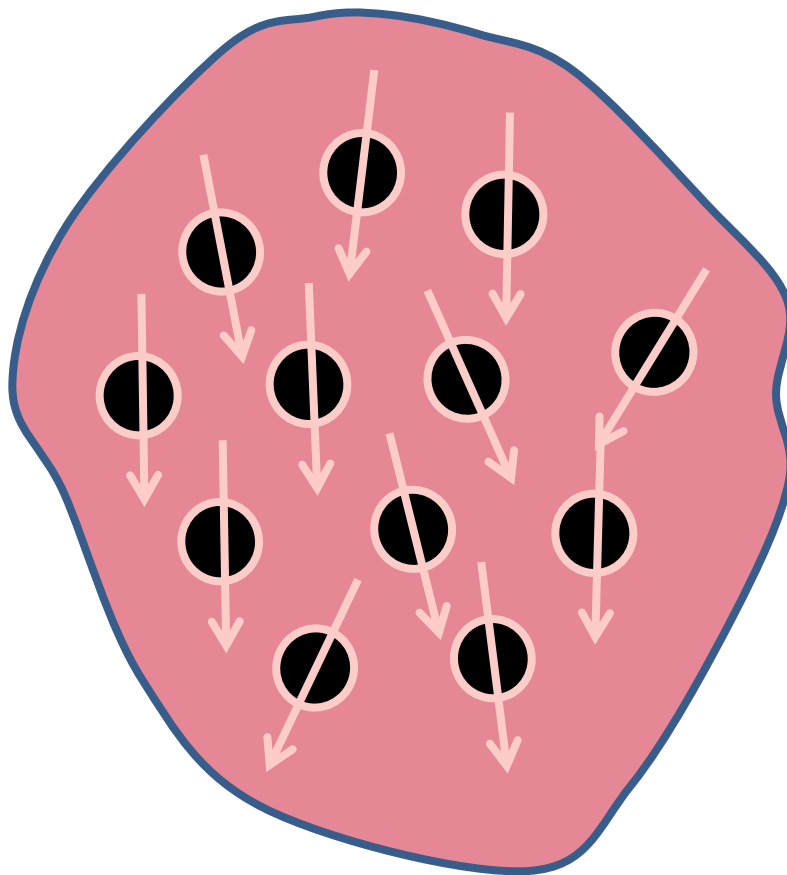
$$\tau \approx 1/\omega$$

**T**

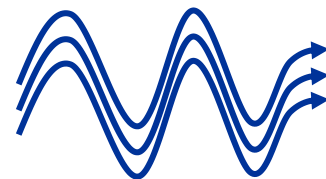


energy  
absorption from  
a RF field  
(SAR)

**T**

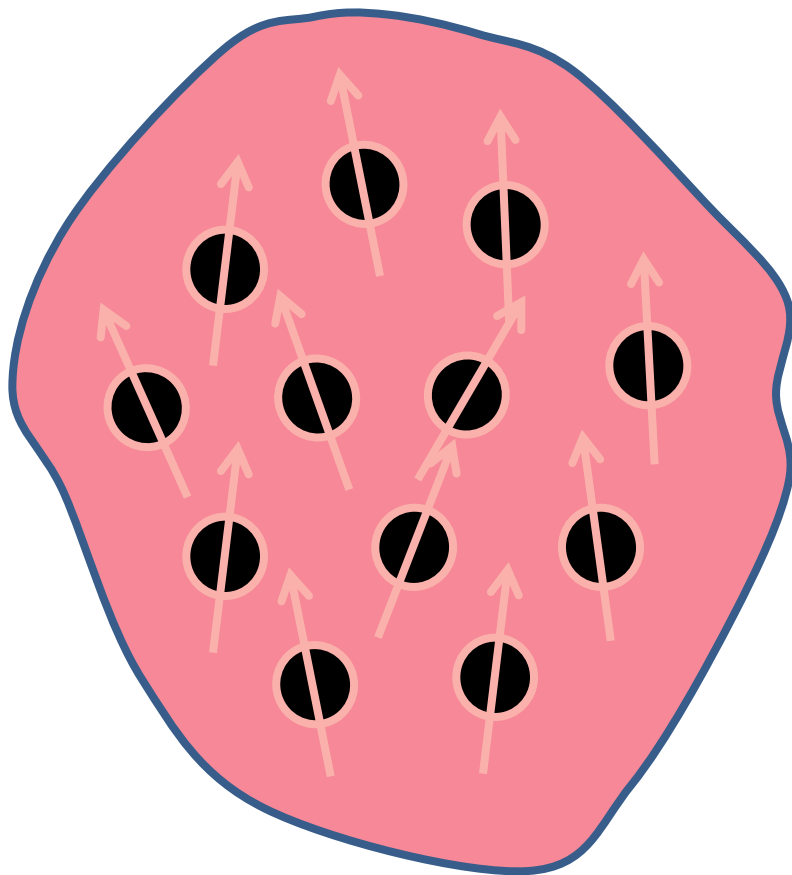


$$\tau \approx 1/\omega$$

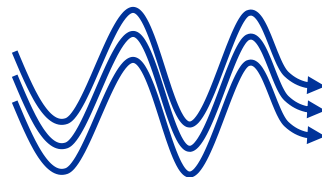


energy  
absorption from  
a RF field  
(SAR)

**T**

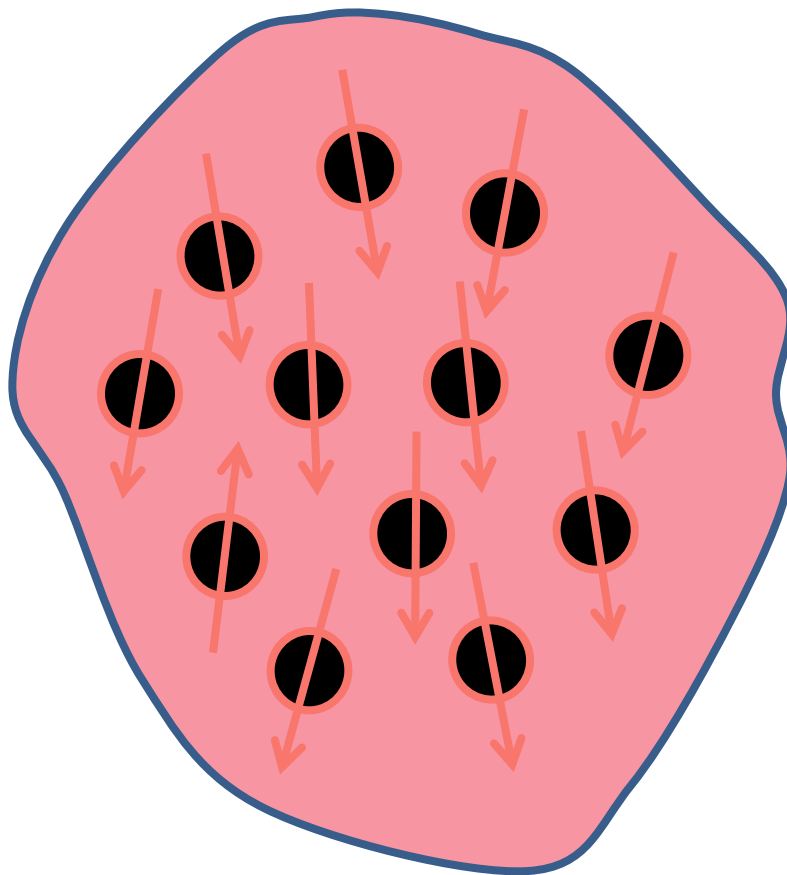
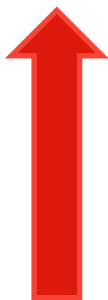


$$\tau \approx 1/\omega$$

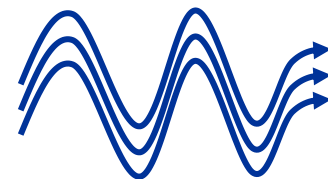


energy  
absorption from  
a RF field  
(SAR)

**T**

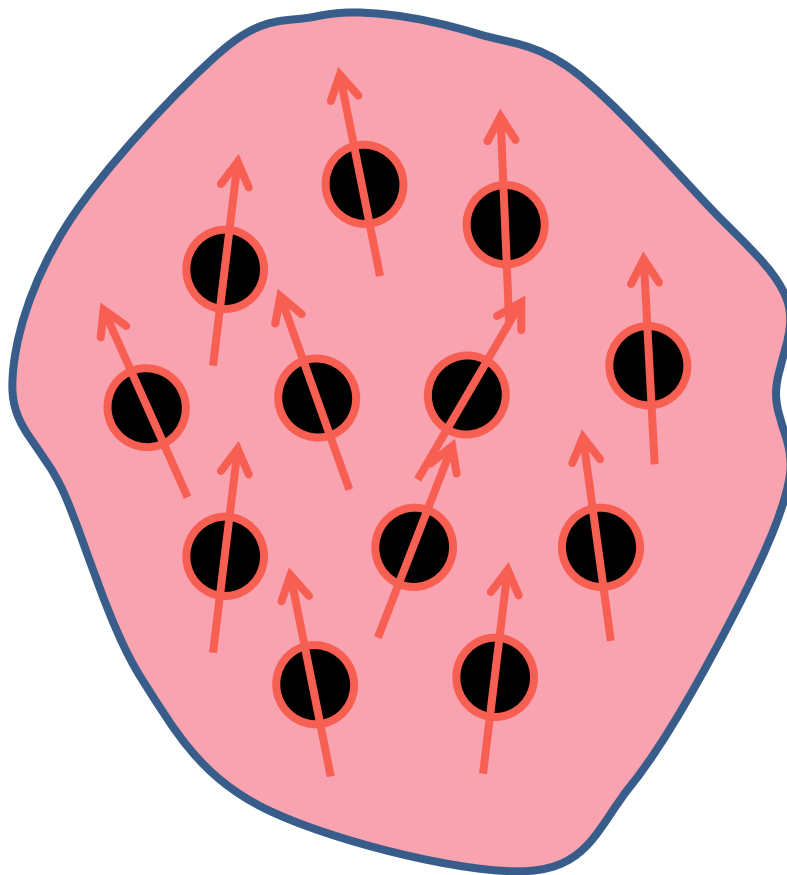


$$\tau \approx 1/\omega$$

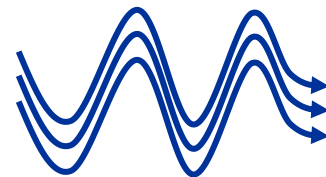


energy  
absorption from  
a RF field  
(SAR)

**T**

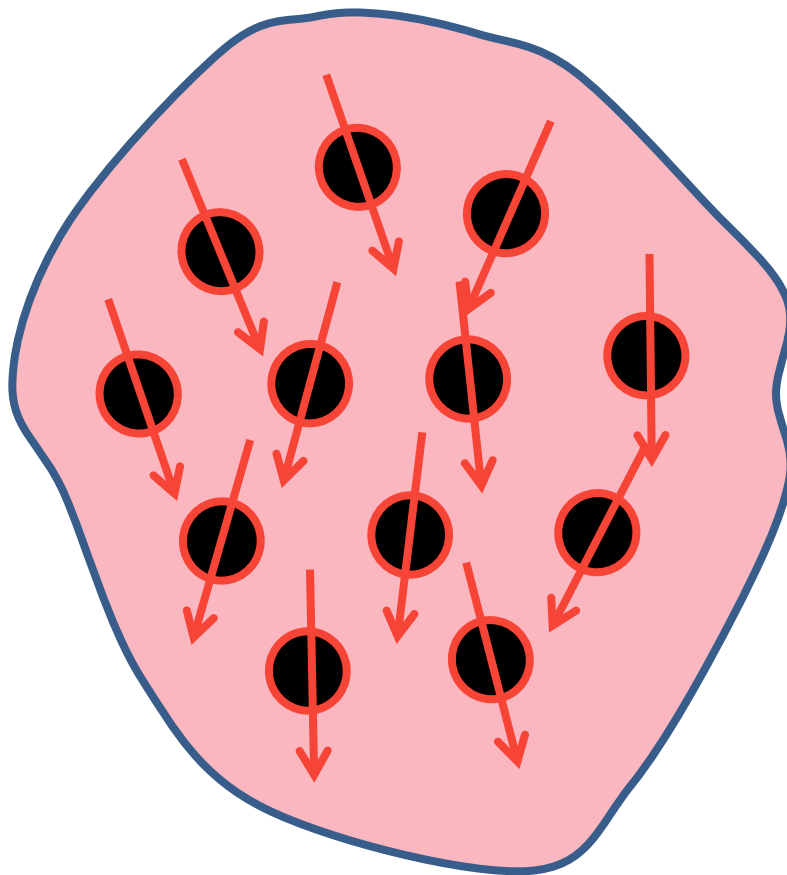


$$\tau \approx 1/\omega$$

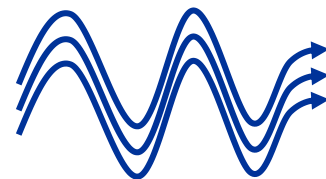


energy  
absorption from  
a RF field  
(SAR)

**T**



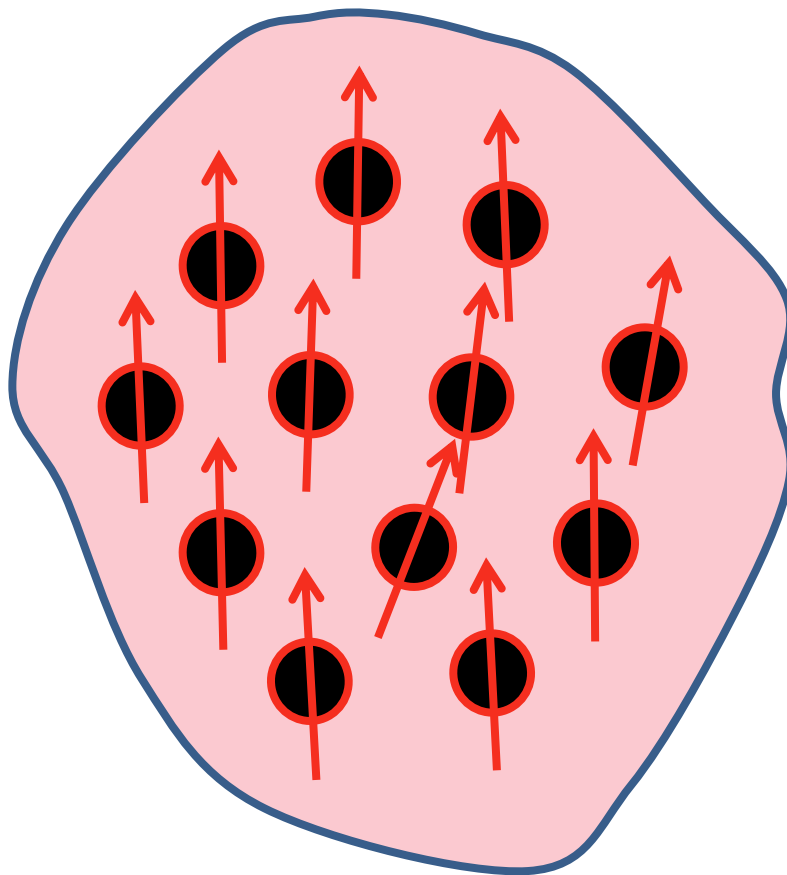
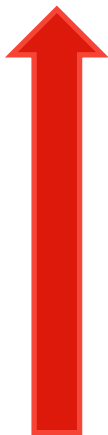
$$\tau \approx 1/\omega$$



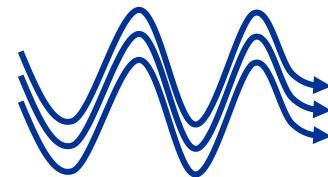


energy  
absorption from  
a RF field  
(SAR)

**T**

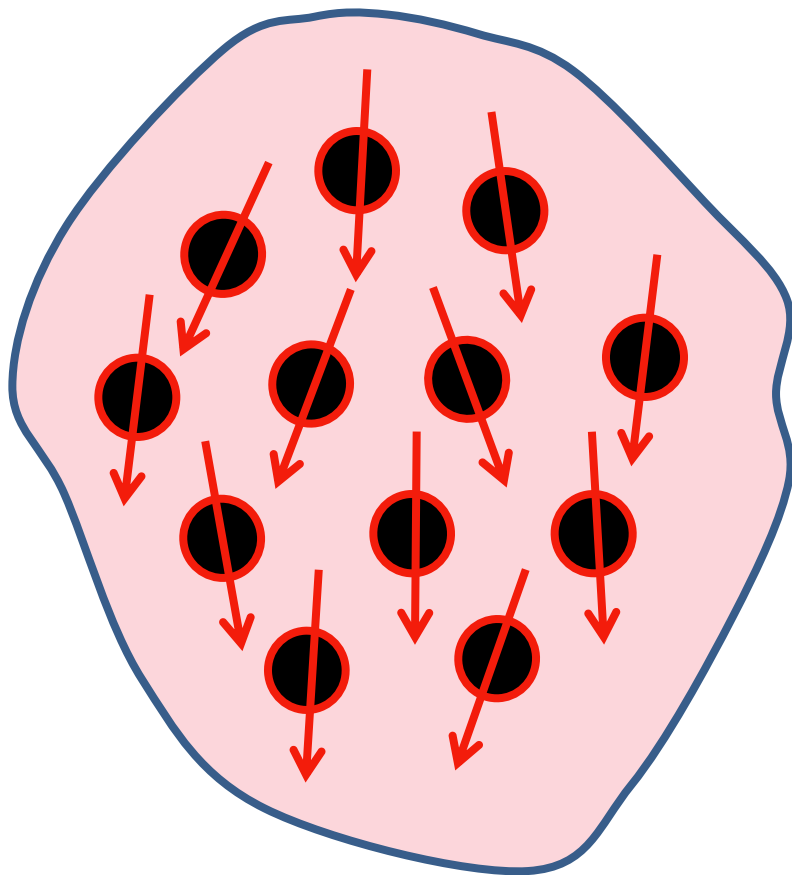
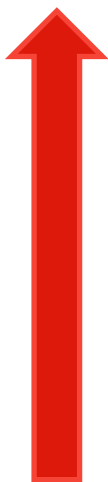


$$\tau \approx 1/\omega$$

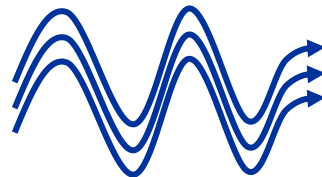


energy  
absorption from  
a RF field  
(SAR)

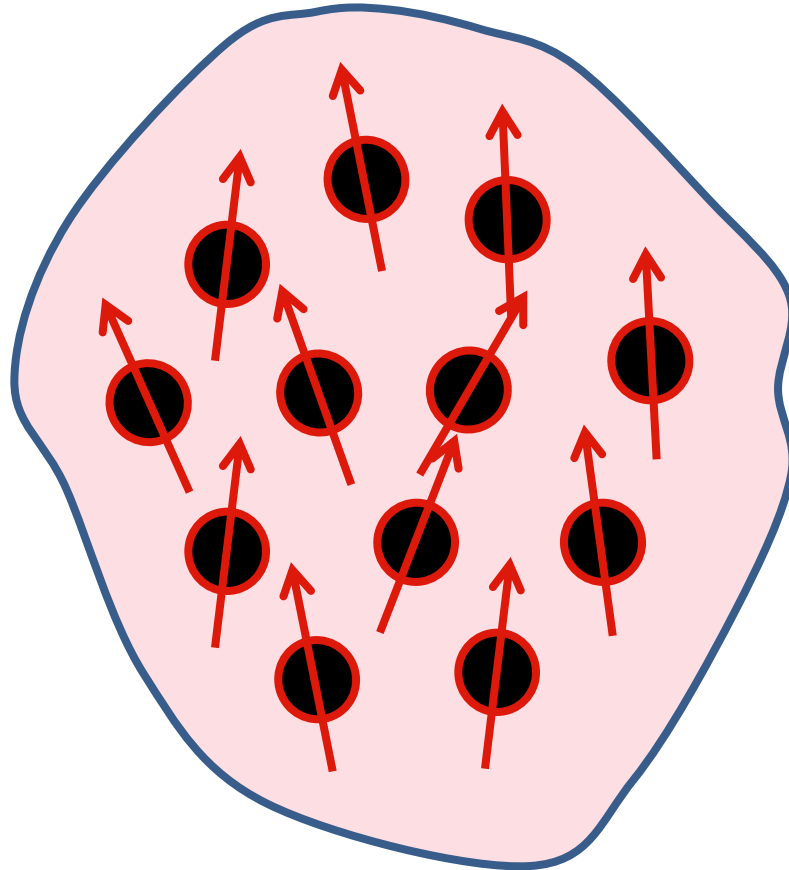
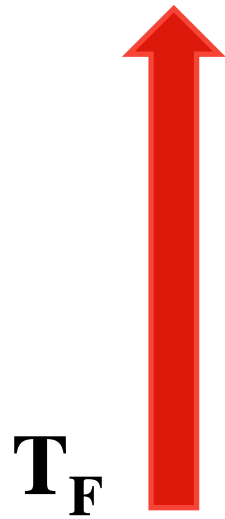
**T**



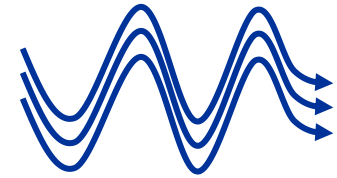
$$\tau \approx 1/\omega$$



energy  
absorption from  
a RF field  
(SAR)



$$\tau \approx 1/\omega$$



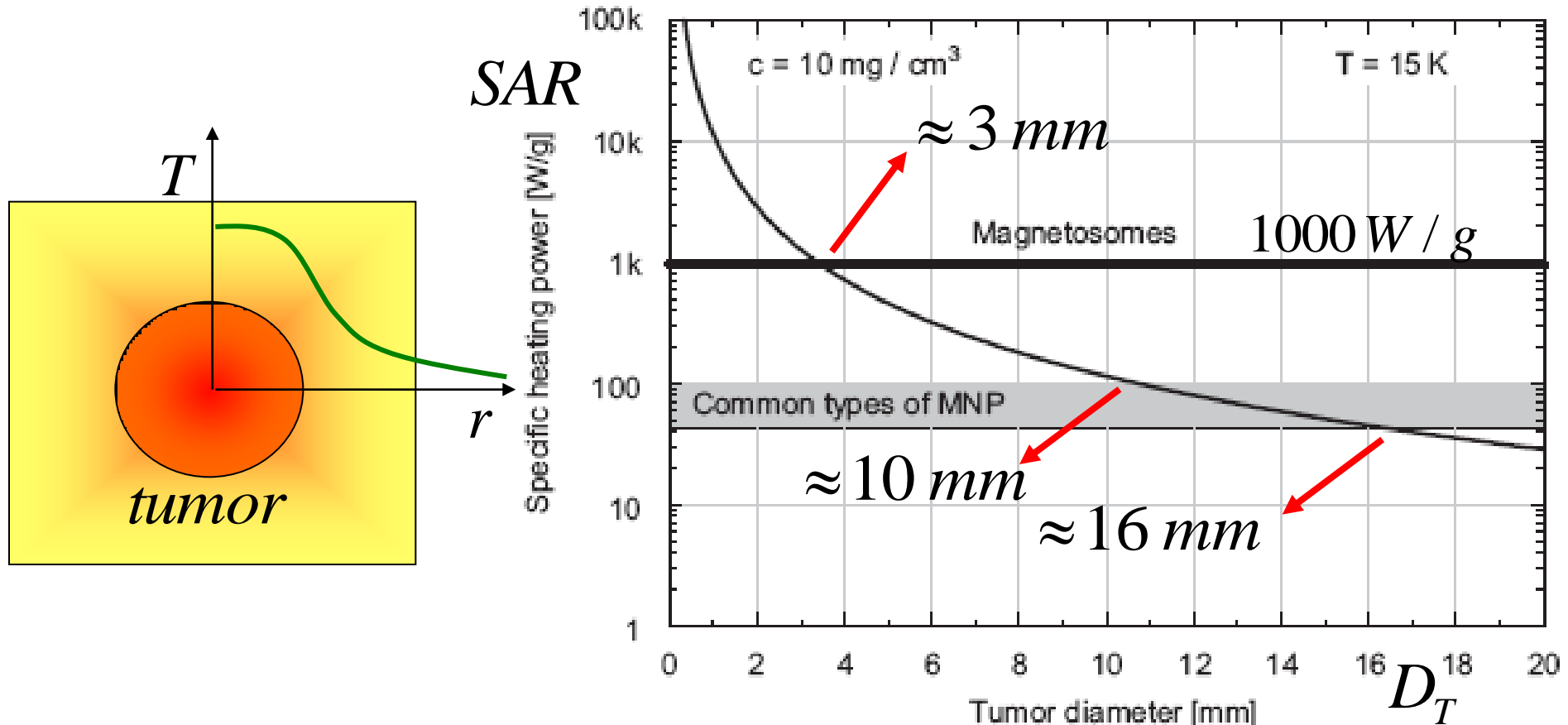
$$SAR \approx f(M_S, V_{MNP}, H_0, \omega, T) \frac{\omega\tau}{1 + (\omega\tau)^2}; \quad \text{need for } 1\text{ kW/g}$$

# Smallest treatable tumor

Magnetic particle hyperthermia—biophysical limitations of a visionary tumour therapy

Rudolf Hergt\*, Silvio Dutz

Journal of Magnetism and Magnetic Materials 311 (2007) 187–192



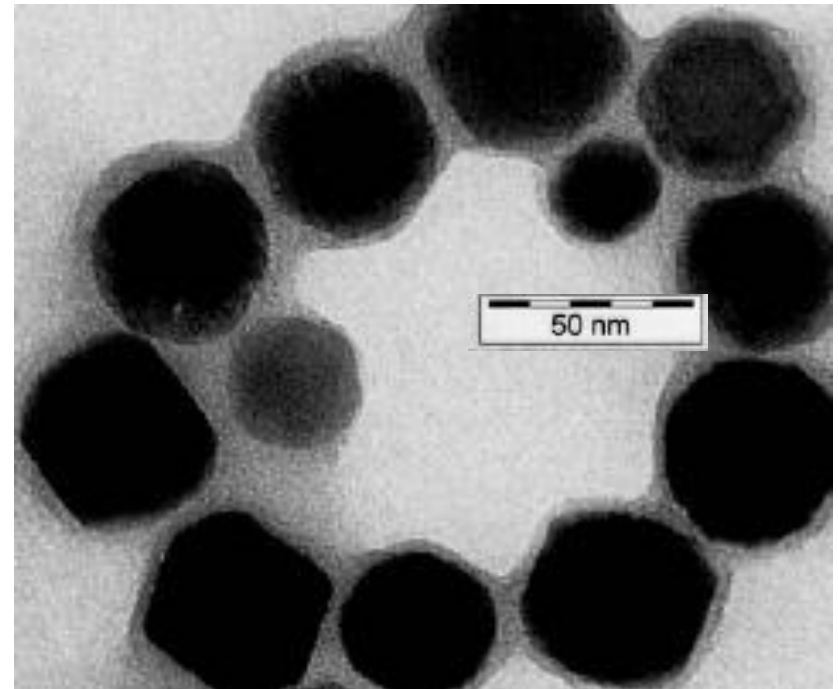
# NP and FF for Magnetic Hyperthermia

Searching for high SAR nanomaterials



Magnetotactic bacteria

magnetosomes



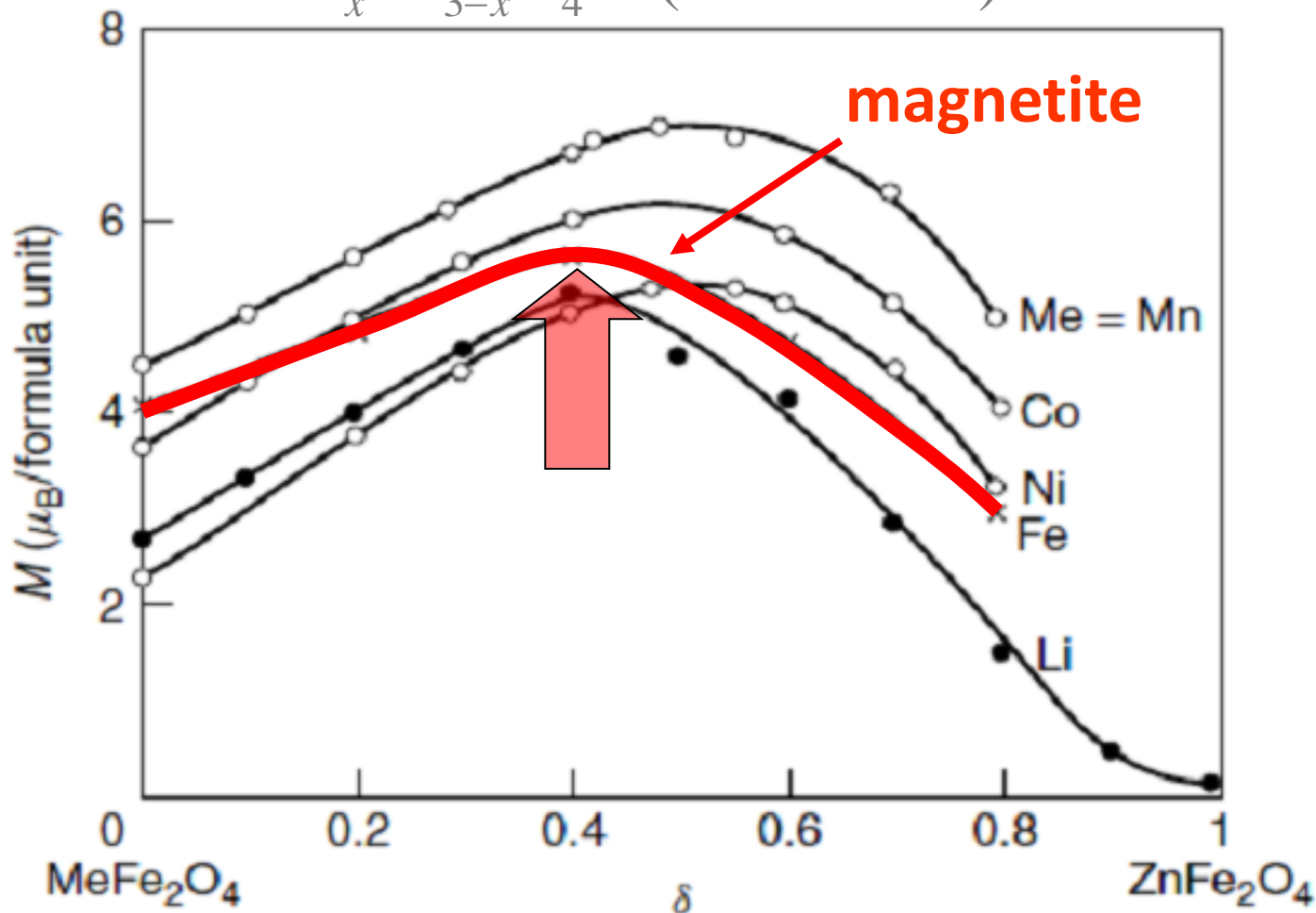
Journal of Magnetism and Magnetic Materials 293 (2005) 80–86  
Hergt et al. 2005

$$SAR > 1 \text{ kW} / \text{g}$$

# Zn doped ferrites



Fe moment  
enhancement

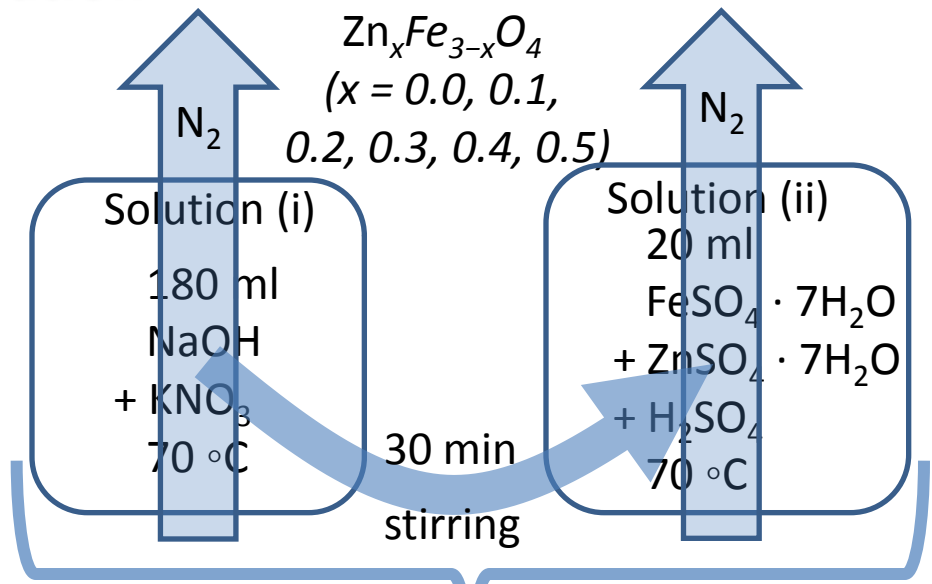


# zinc-doped magnetite nanoparticles and ferrofluids for hyperthermia applications

*P Mendoza Zélis, G A Pasquevich, S J Stewart, M B Fernández van Raap, J Apesteguy, I J Bruvera, C Laborde, B Pianciola, S Jacobo and F H Sánchez*

*Structural and magnetic study of zinc-doped magnetite nanoparticles and ferrofluids for hyperthermia applications, **J. Phys. D: Appl. Phys.** 46, (2013), 125006 doi:10.1088/0022-3727/46/12/125006.*

# NP preparation



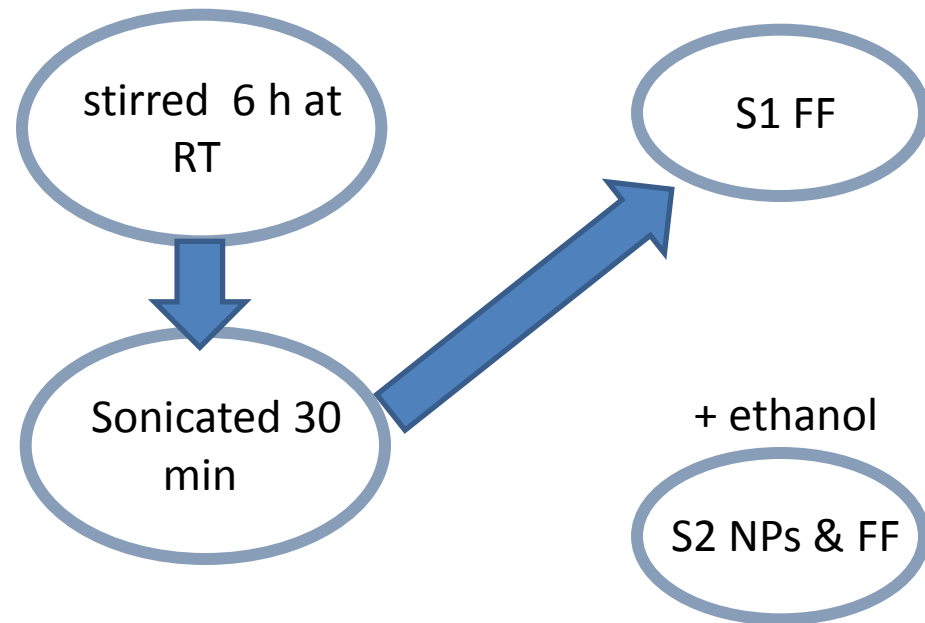
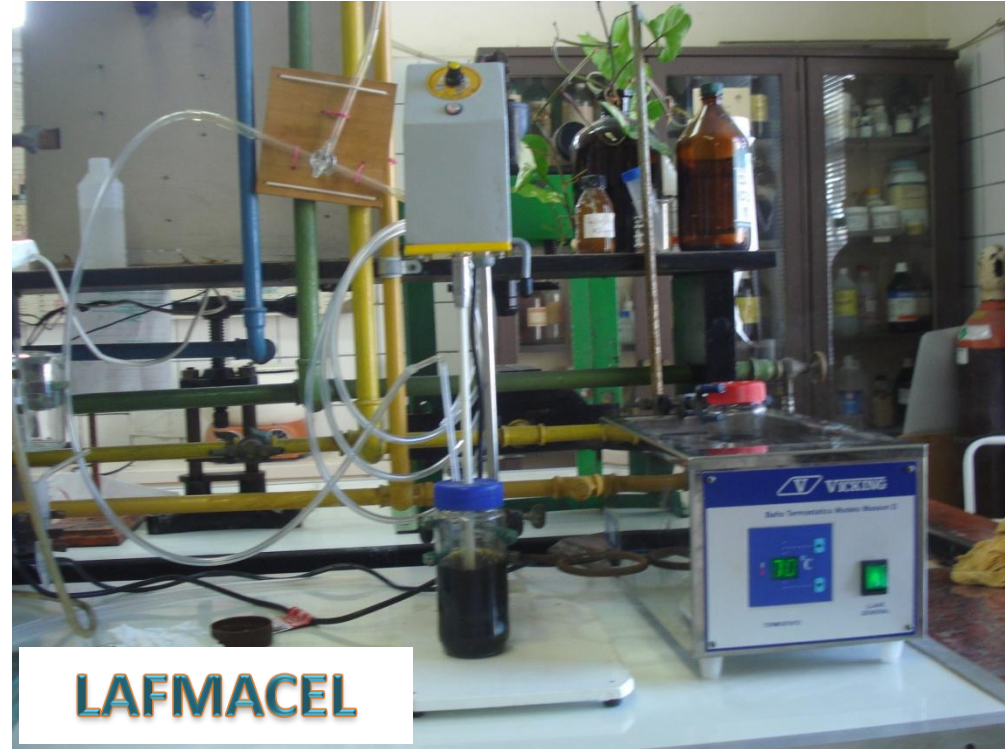
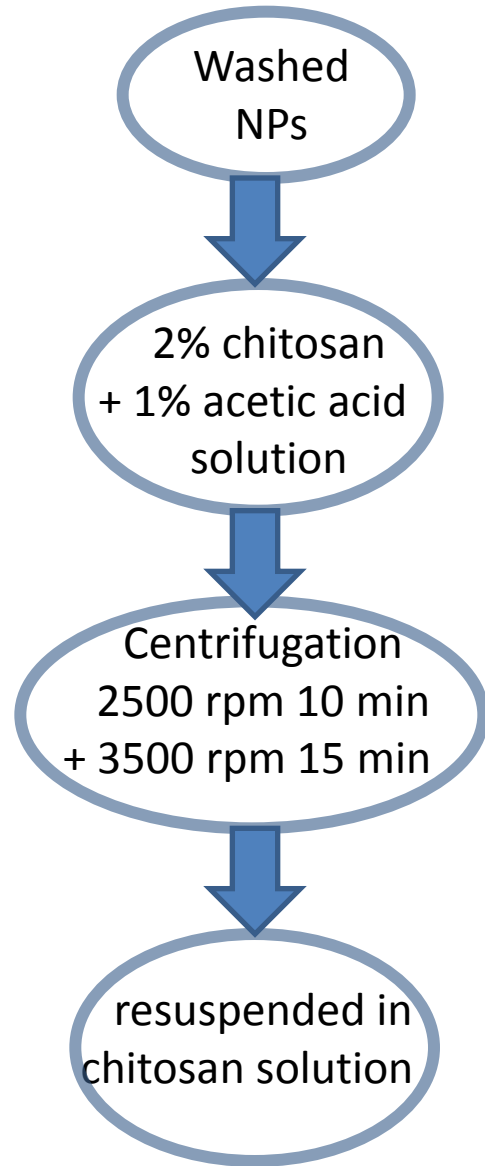
- kept at 70 °C for 2 h
- cooled down to RT
- magnetic decantation
- washed several times
- centrifuged
- Dried 40 °C Vacuum 24 h
- S1 NPs



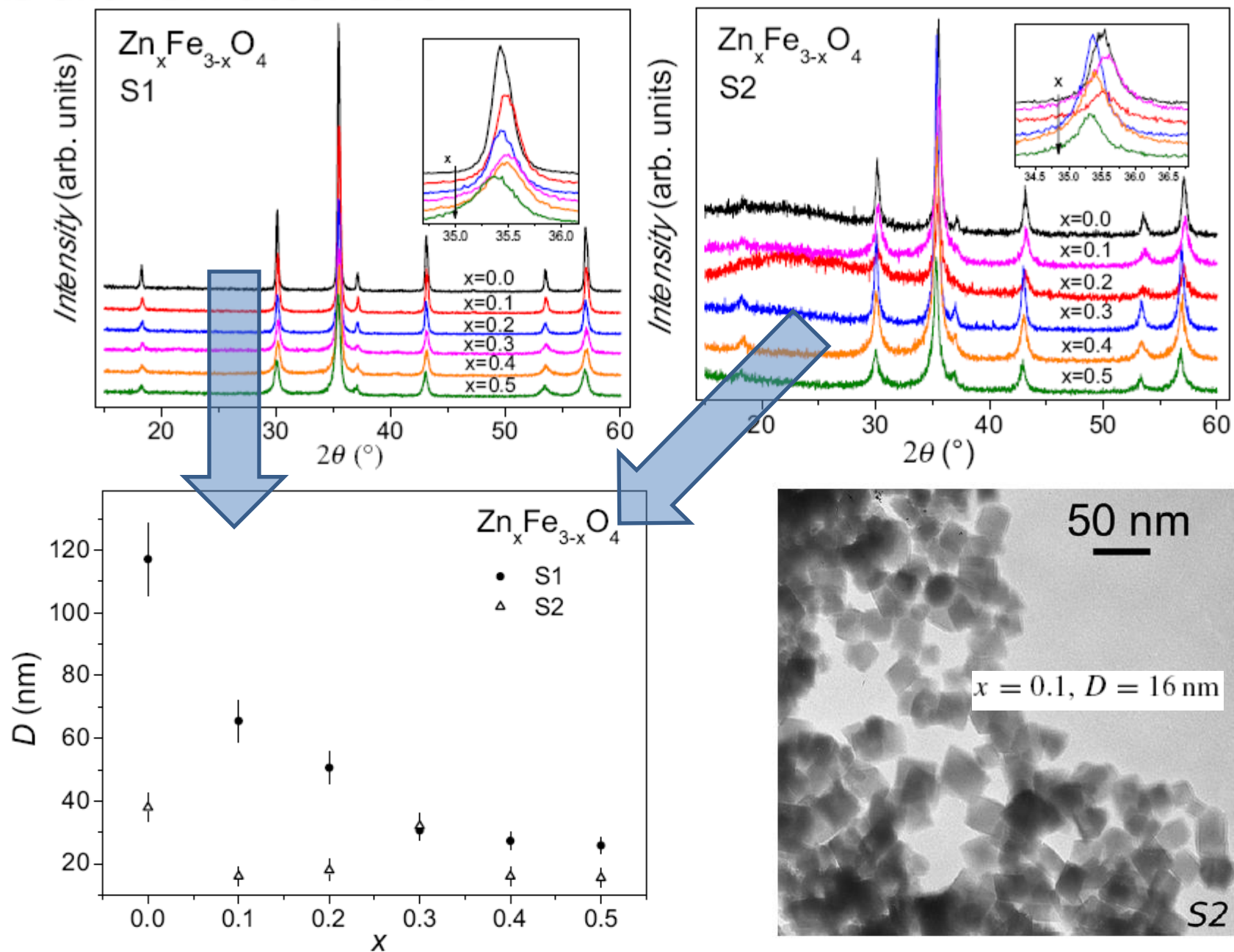
LAFMACEL



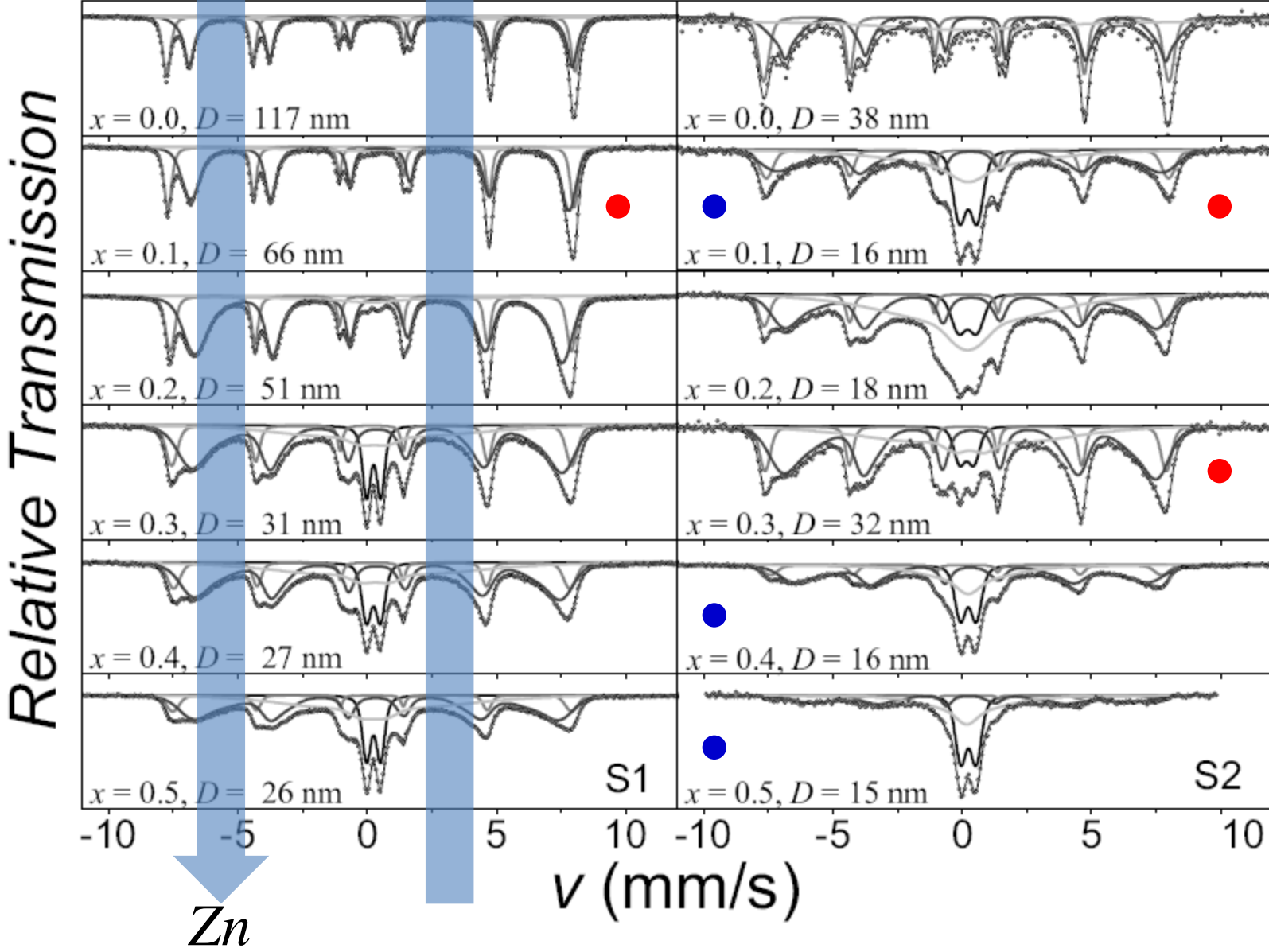
# FF preparation and purification



# XRD and TEM observation



# Mössbauer effect results



# Mössbauer effect results

subspectra

*slow relaxing*

*fast relaxing*

*A B*

$x = 0.3, D = 31 \text{ nm}$

-10

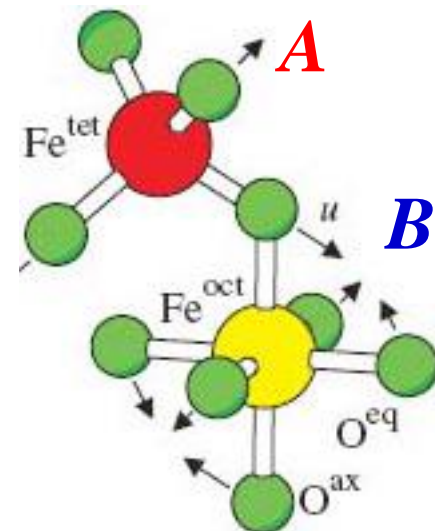
-5

0

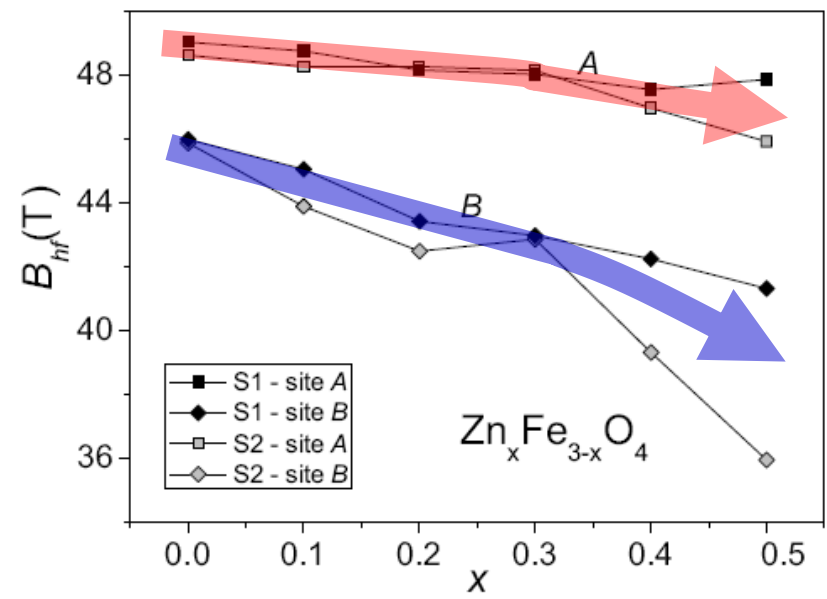
5

10

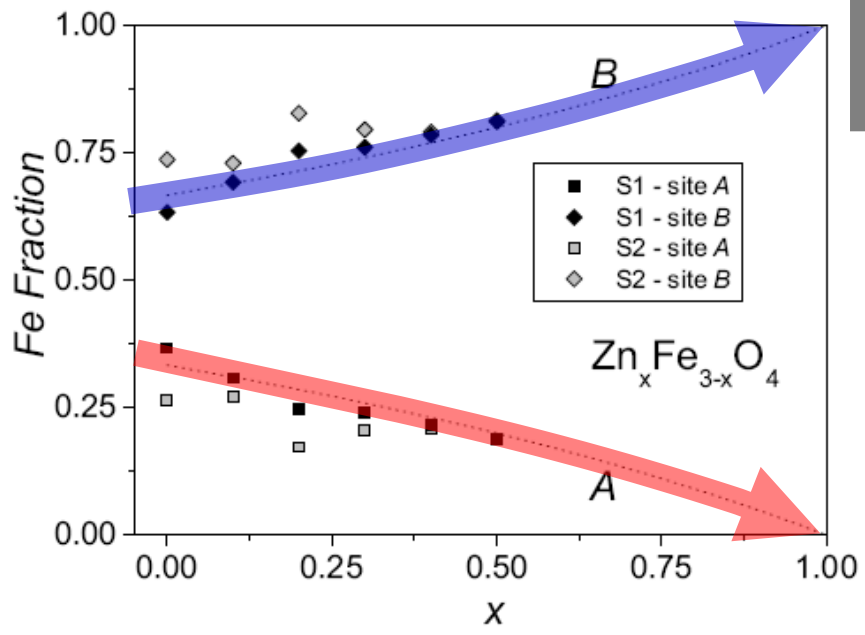
$v \text{ (mm/s)}$



# Mössbauer effect results

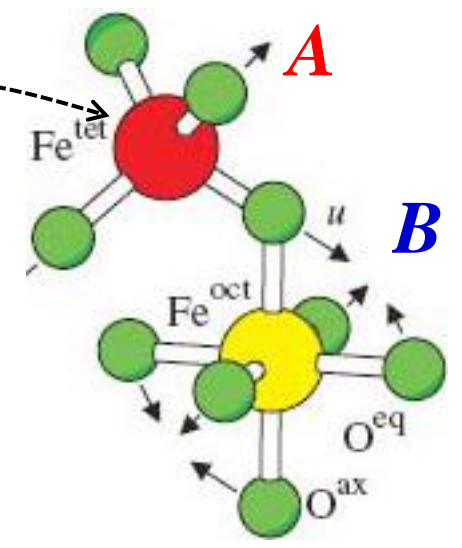


*Hyperfine Field  $B$*



**Zn**

*Fe site fraction*



# Mössbauer effect results

*in magnetite:*

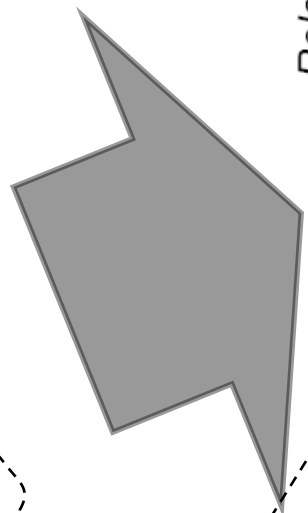
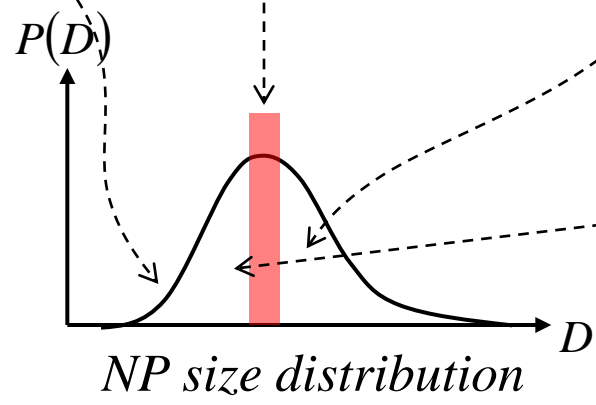
$$\tau_{Larmor} \approx 30 \text{ ns}$$

*Mössbauer spectra*

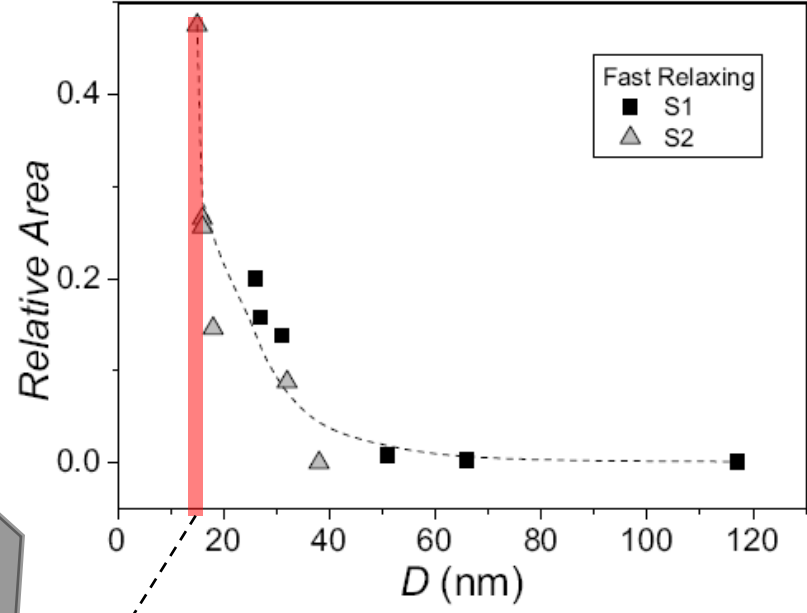
$\tau < \tau_{Larmor} \Rightarrow$  *fast relaxing*

$\tau > \tau_{Larmor} \Rightarrow$  *non relaxing*

$\tau \approx \tau_{Larmor} \Rightarrow$  *slow relaxing*



*Mössbauer blocking size at RT ~15 nm*

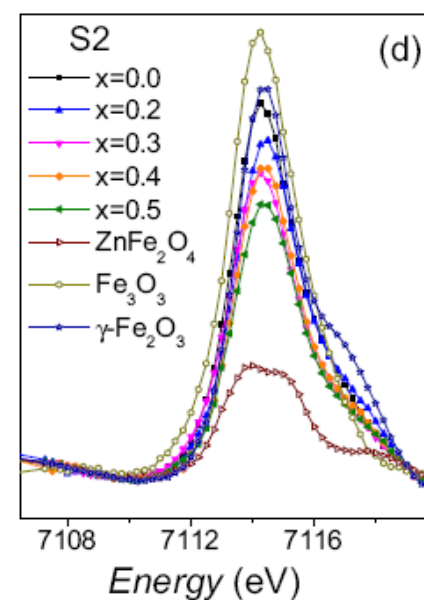
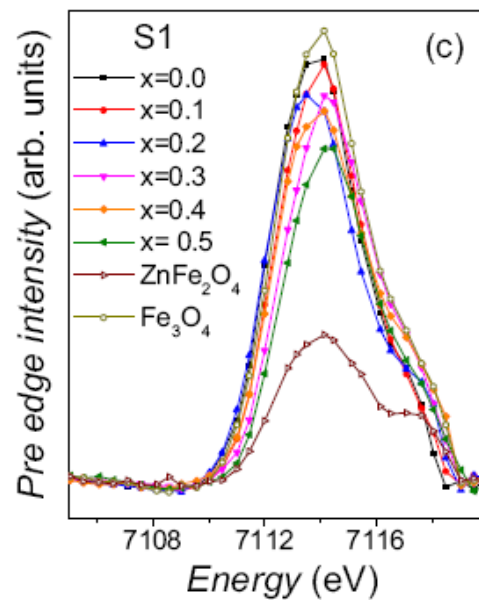
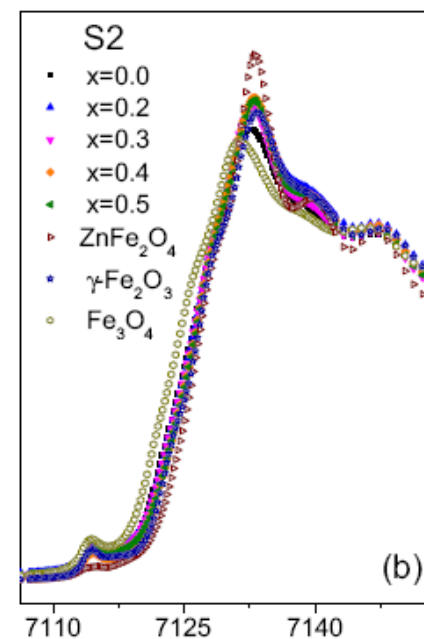
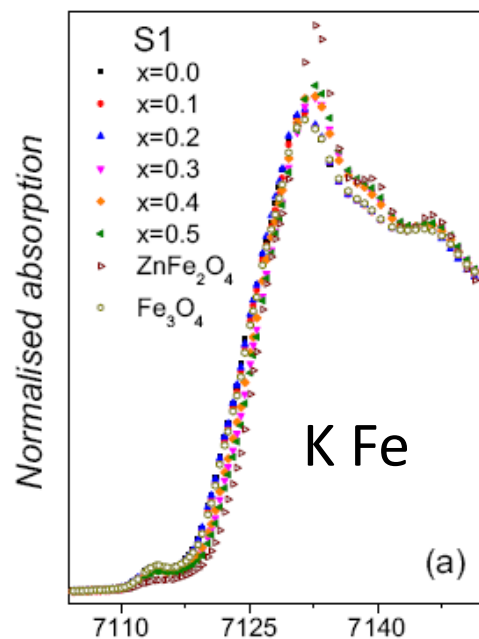
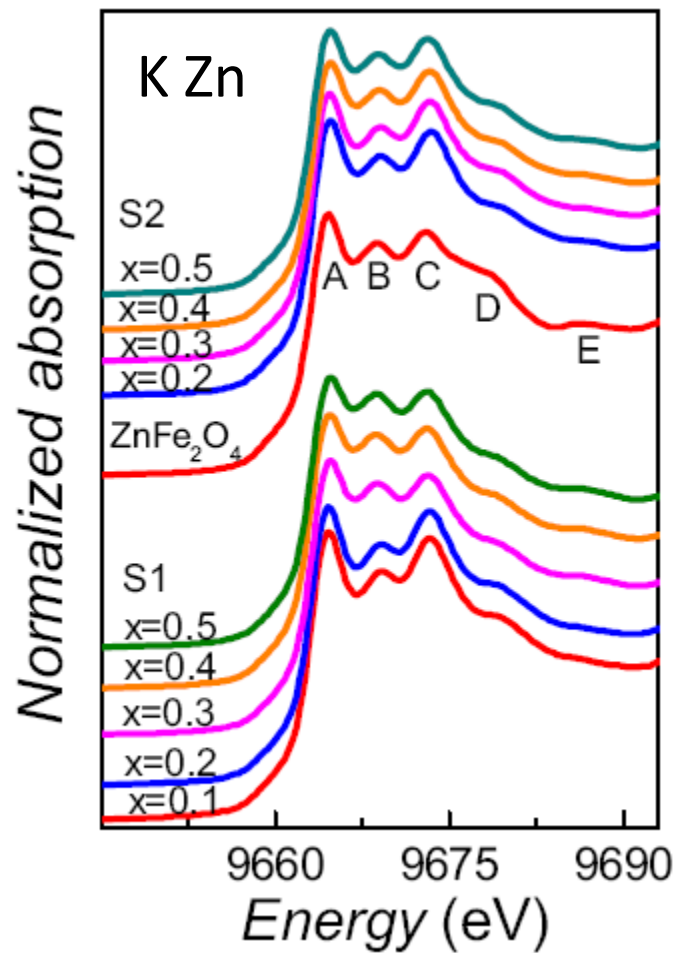


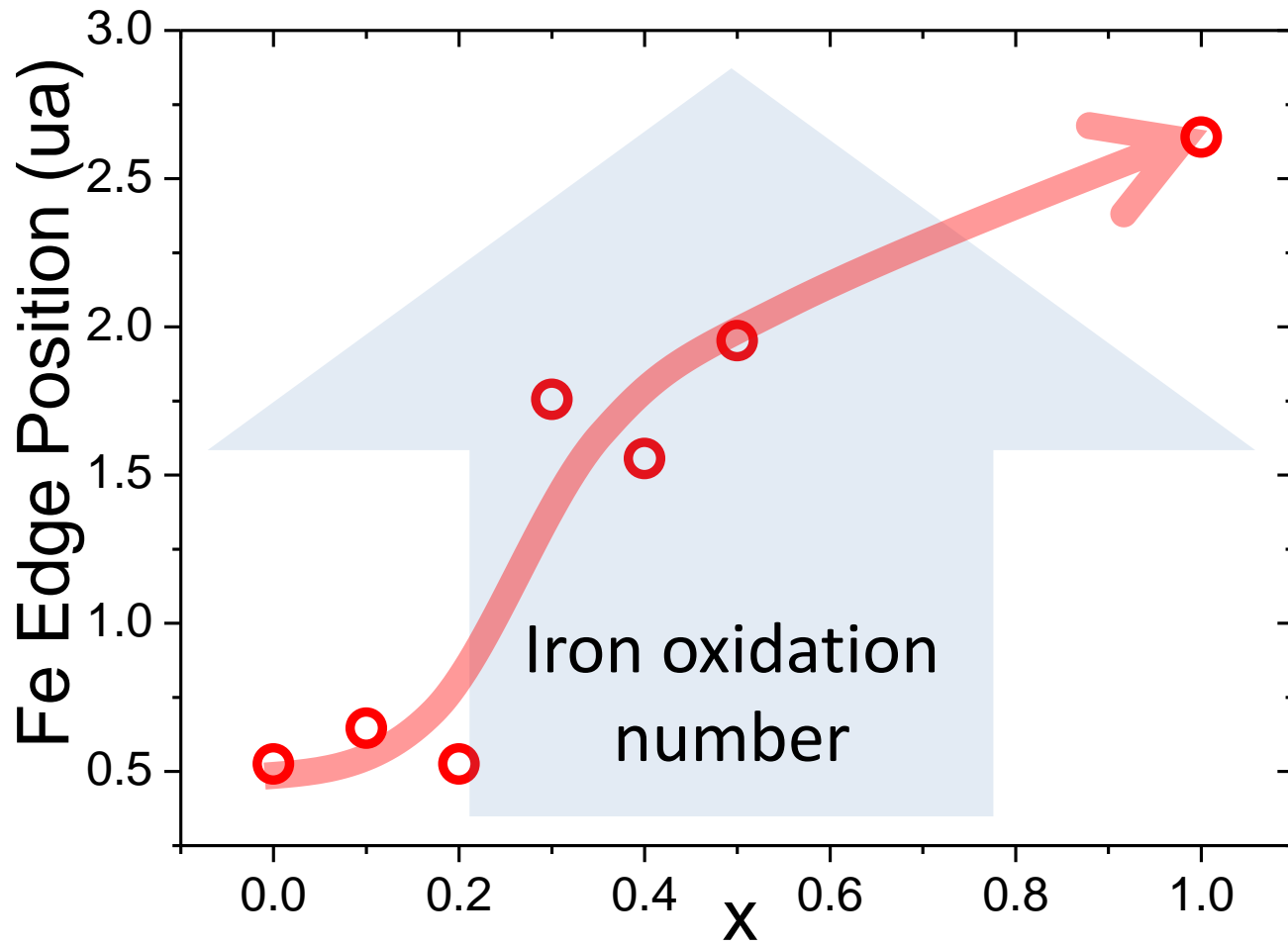
$$\tau_{Larmor} = \tau_0 e^{K_{eff} V / kT}$$

$V, \tau_{Larmor}$   
 $\tau_0 \sim 1 \times 10^{-9} \text{ s}$

$$K_{eff} \approx 1.9 \times 10^4 \text{ J / m}^3$$

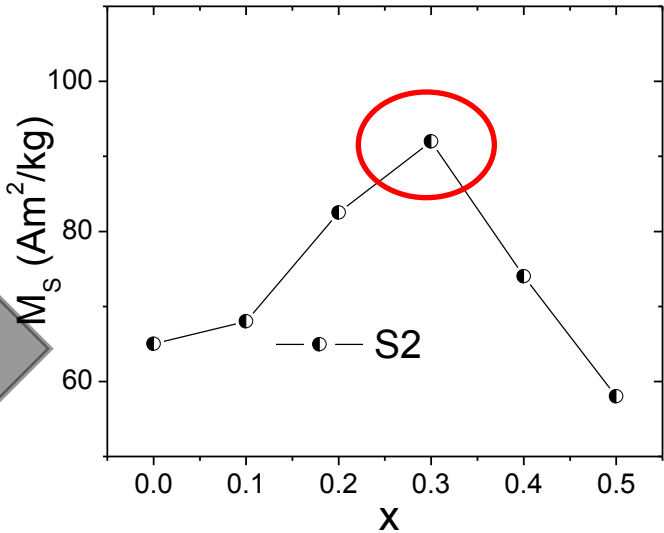
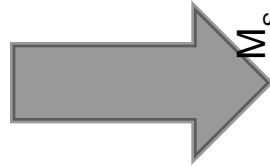
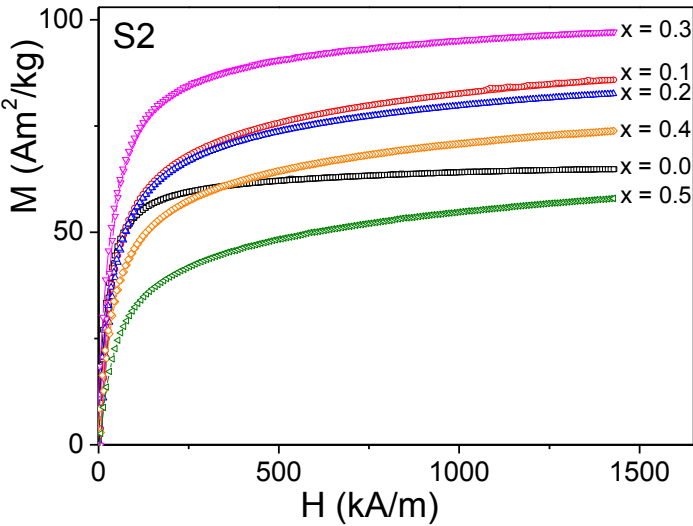
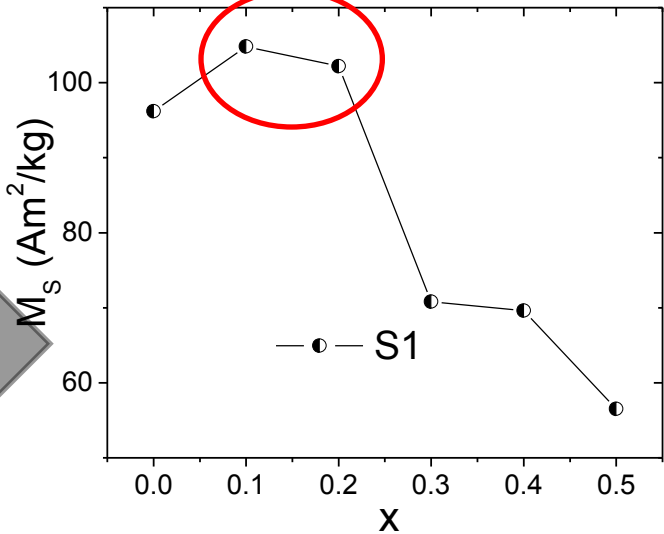
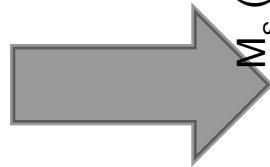
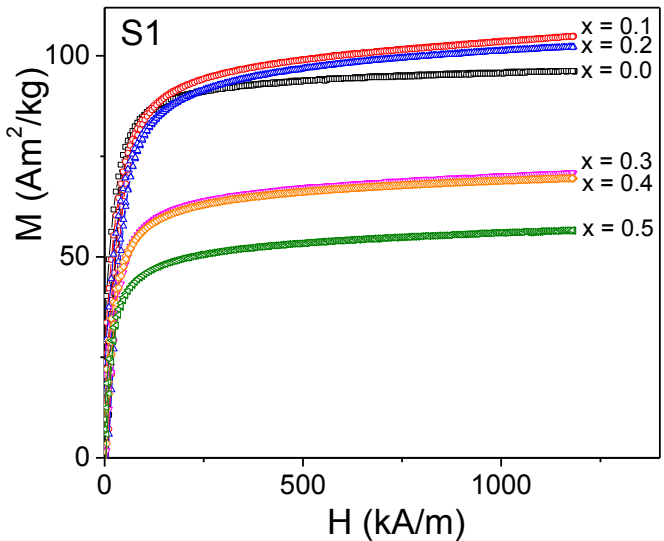
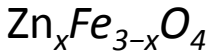
# XAS observation







# Magnetization results

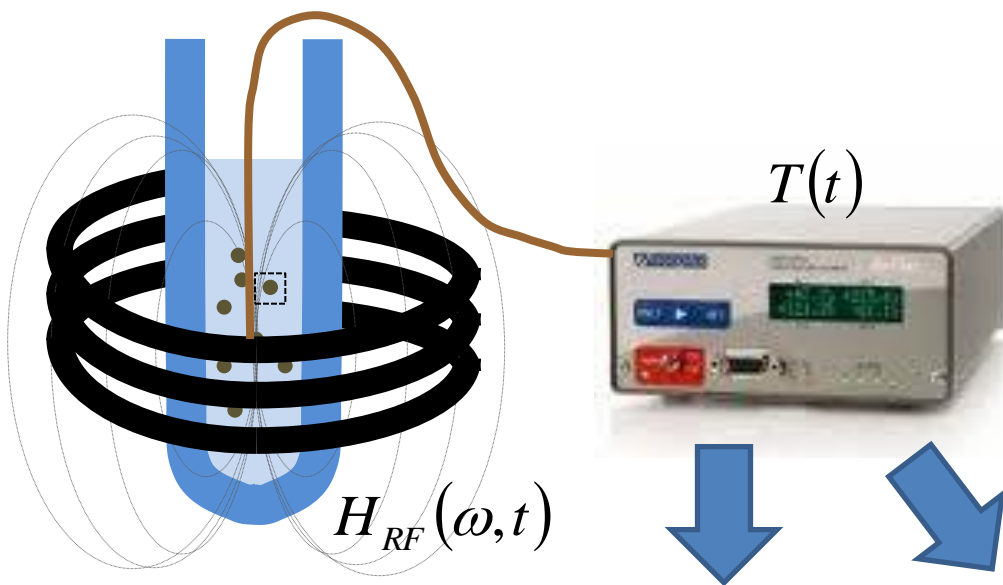


$\mu_{UF} \approx 4\mu_B$

$Zn \Rightarrow A \text{ site}$

$\mu_{UF} \approx 10\mu_B$

# SAR experiments

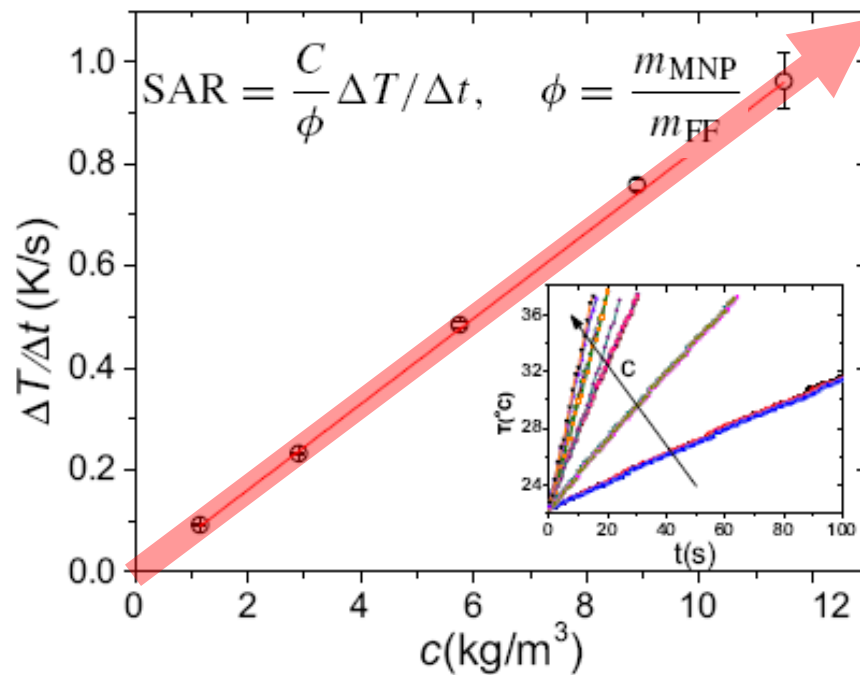
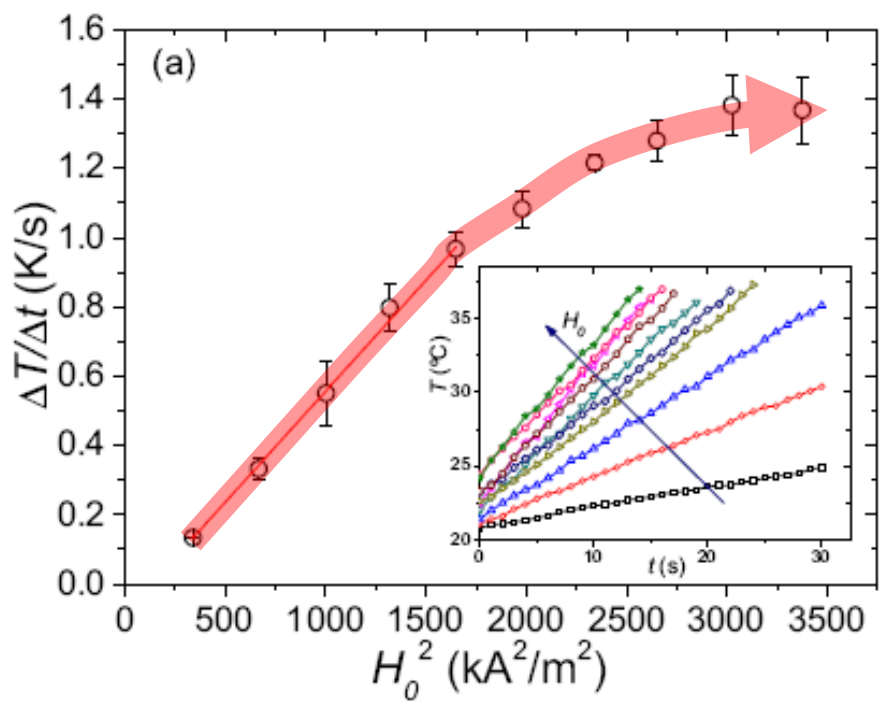


$$H_{RF}(\omega, t)$$

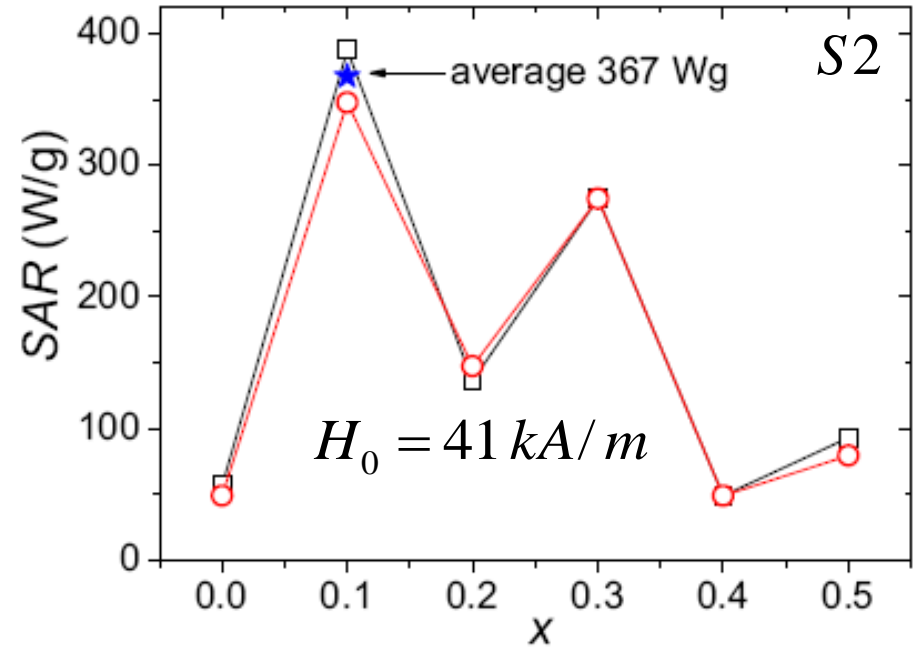
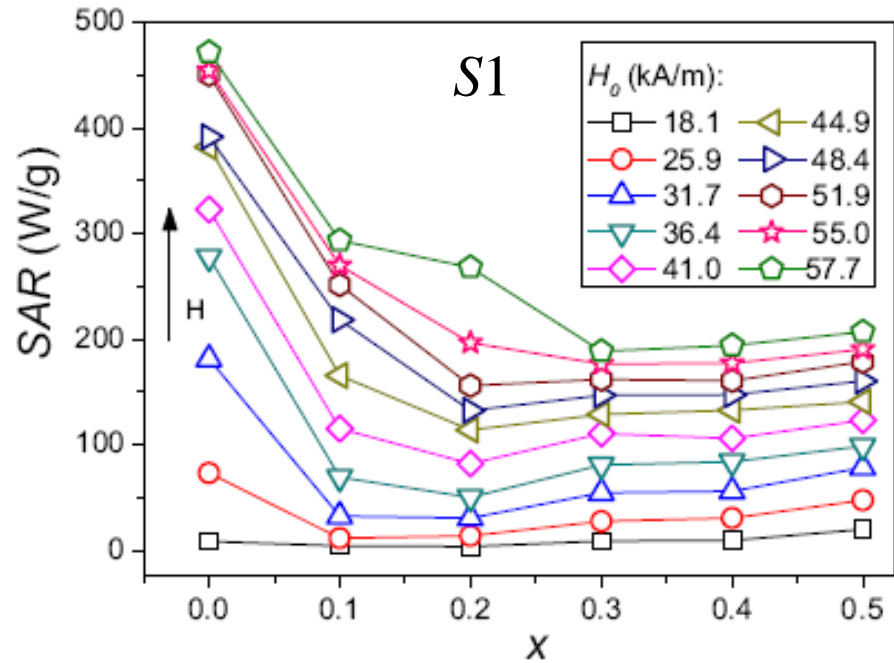
$$H_0 \leq 57.7 \text{ kA/m}$$

$$\omega = 2\pi f$$

$$f = 260 \text{ kHz}$$



# SAR results



$$SAR = \frac{\pi \mu_0 H_0^2 f}{\rho} \frac{\mu_0 M_S^2 V \rho^2}{3kT} \frac{\omega \tau}{1 + (\omega \tau)^2}$$

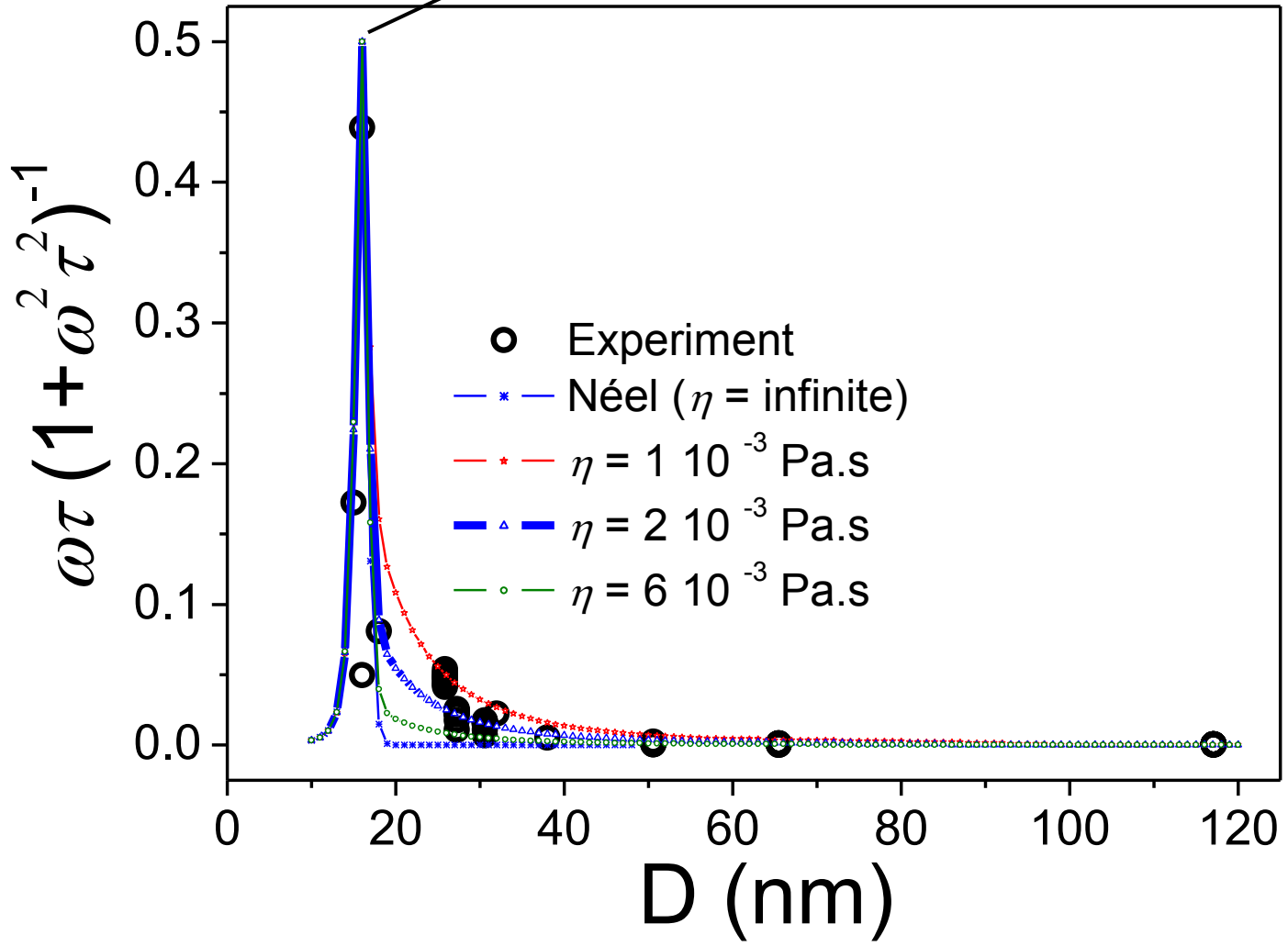
Normalizing all SAR data by  $H_0$ ,  $M_S$  and  $V$

$$\frac{SAR}{H_0^2 M_S^2 V} = \frac{\pi \mu_0^2 f \rho}{3kT} \frac{\omega \tau}{1 + (\omega \tau)^2} = C \frac{\omega \tau}{1 + (\omega \tau)^2}$$

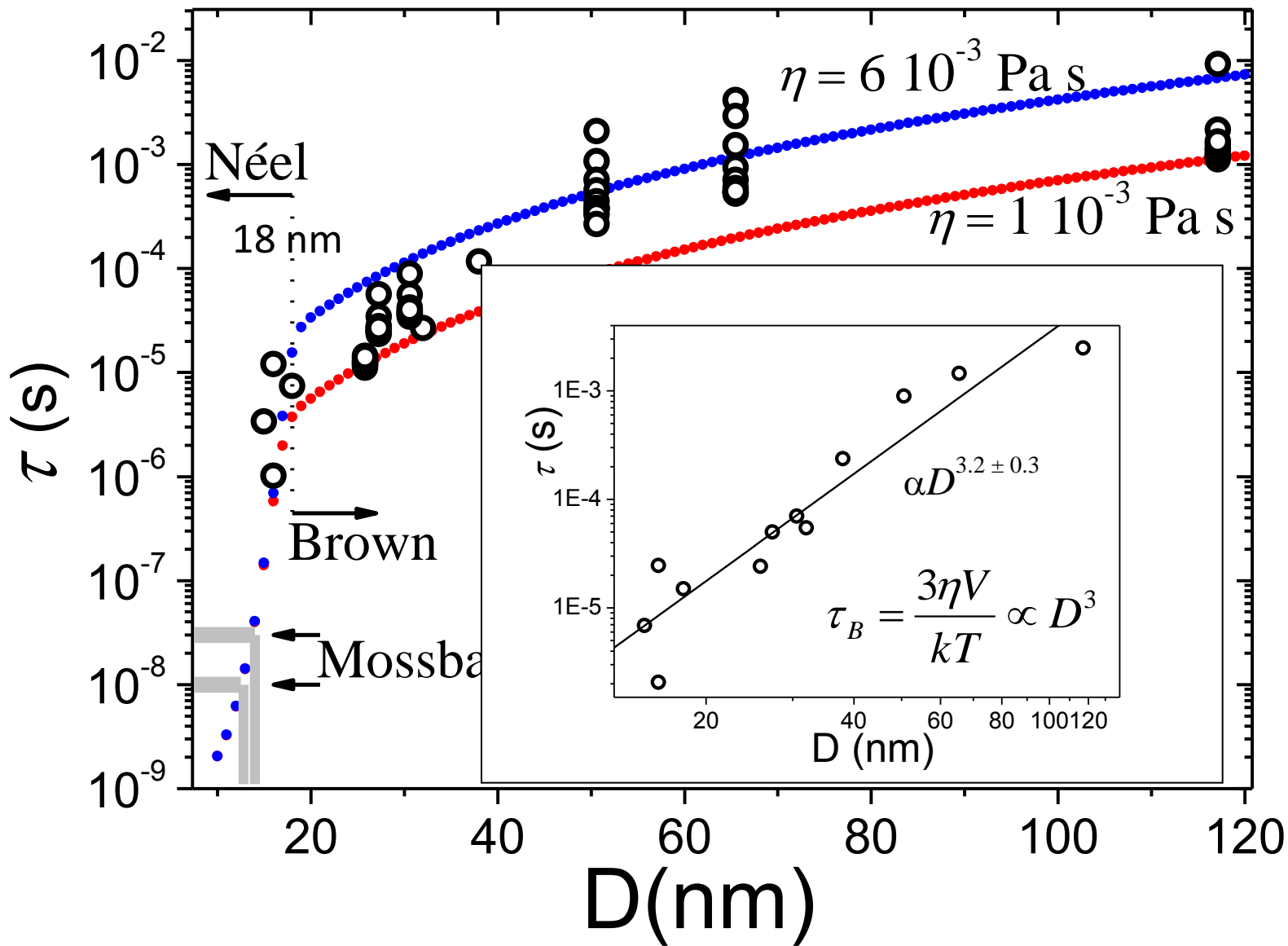
# SAR results – Relaxation time

$$\frac{SAR}{CH_0^2 M_s^2 V} = \frac{\omega\tau}{1 + (\omega\tau)^2}$$

16 nm



$$\frac{\omega\tau}{1+(\omega\tau)^2} \Rightarrow \tau$$



## Conclusions II

$M_s$  increases with Zn low doping of magnetite ( $x = 0.1-0.3$ )

High SARs of almost 400 W/g can be obtained

At 260 kHz crossover from Néel to Brown relaxation is at 18 nm

At 260 kHz frequency factor is maximum at 16 nm

SAR can be improved further by increasing  $M_s$  of 16 nm NPs and reducing polydispersity

# Citric Acid Coated Magnetite Nanoparticles for Magnetic Hyperthermia

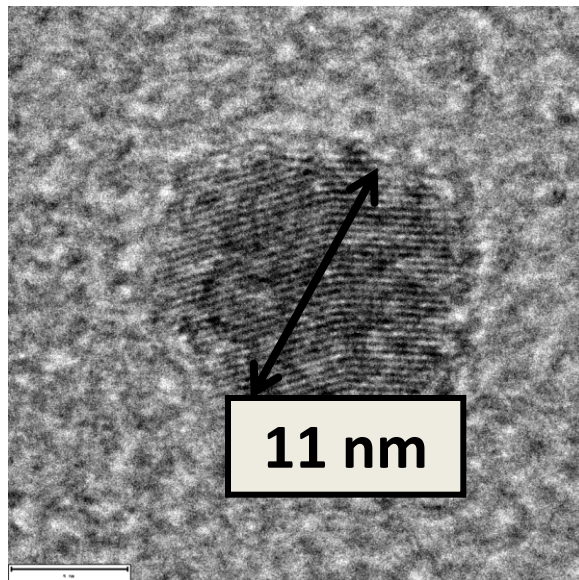
*M. Elisa de Sousa, Marcela B. Fernández van Raap, Patricia C. Rivas, Pedro Mendoza Zélis, Pablo Girardin, Gustavo A. Pasquevich, Jose L. Alessandrini, Diego Muraca, and Francisco H. Sánchez*

*Stability and relaxation mechanisms of citric acid coated magnetite nanoparticles for magnetic hyperthermia, J. Phys. Chem. C, (2013), 117 (10), pp 5436–5445. DOI: 10.1021/jp311556b.*

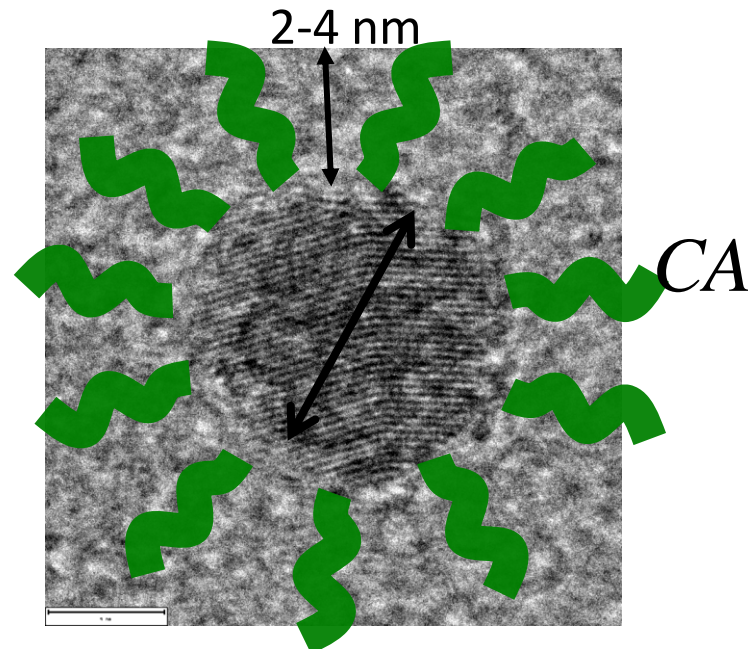
## Modified Masart Method

Chemical coprecipitation from ferric chloride and ferrous chloride in the presence of excess ammonia  $\text{NH}_4\text{OH}$  solution (AS)  $\rightarrow$  magnetite NPs.

Magnetite cores were negatively charged by Citric Acid (CA) adsorption over its surfaces. Both steps, co-precipitation and CA adsorption, were carried out under a  $\text{N}_2$  reflux at  $60^\circ\text{C}$ .



uncoated



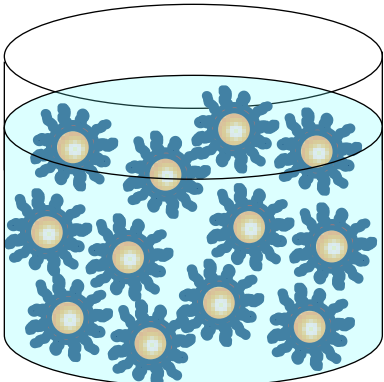
CA coated



# preparation

The pH at which CA was adsorbed to the MNP surface ( $pH_{ads}$ ) was varied from 4.58 to 7.08. AS (0.25% w/w) was used to adjust the suspension pH to  $pH_{sus}$  close to 7.

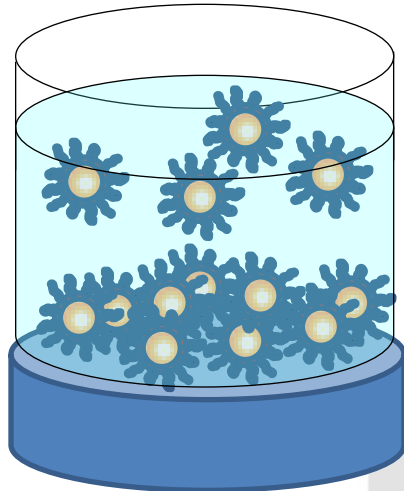
The suspensions were placed on a permanent magnet during 600 s. By this way six colloids  $C_i$  were obtained.



$C_i$



$pH_{ads}^i$



→  $CS_i$

→  $CP_i$

magnet  
(600 s)

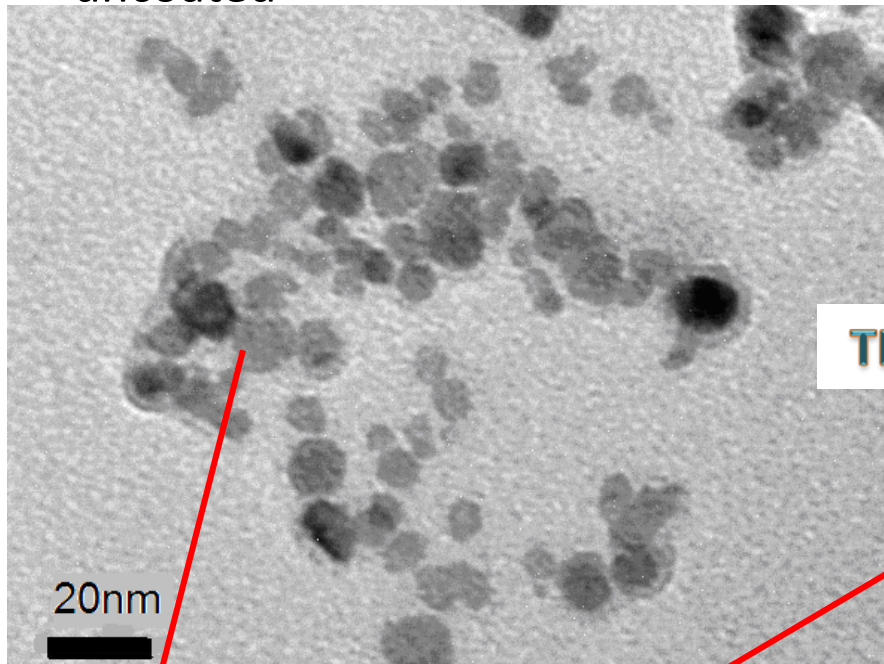
$$f = \frac{[CS_i]}{[C_i]}$$

CA adsorption efficiency and long term stability parameter

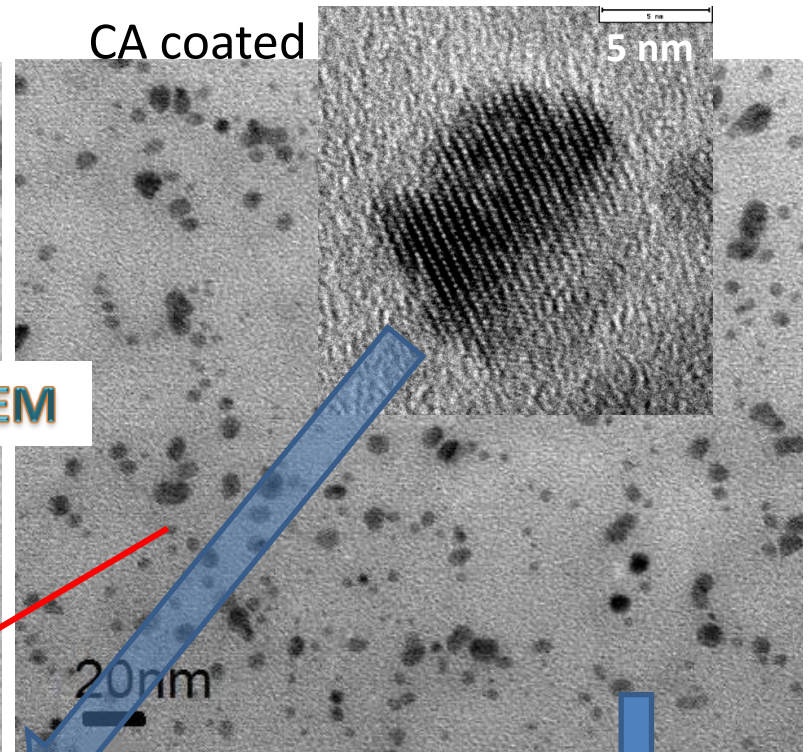
	[X] (g/L)	$pH_{ads} \pm 0.01$	$pH_{sus} \pm 0.01$
$CS_1$	13.4	4.58	7.44
$CS_2$	10.7	4.91	7.34
$CS_3$	18.3	5.50	6.97
$CS_4$	13.1	6.25	7.22
$CS_5$	5.6	6.88	7.10
$CS_6$	7.4	7.08	7.22

# Structural studies

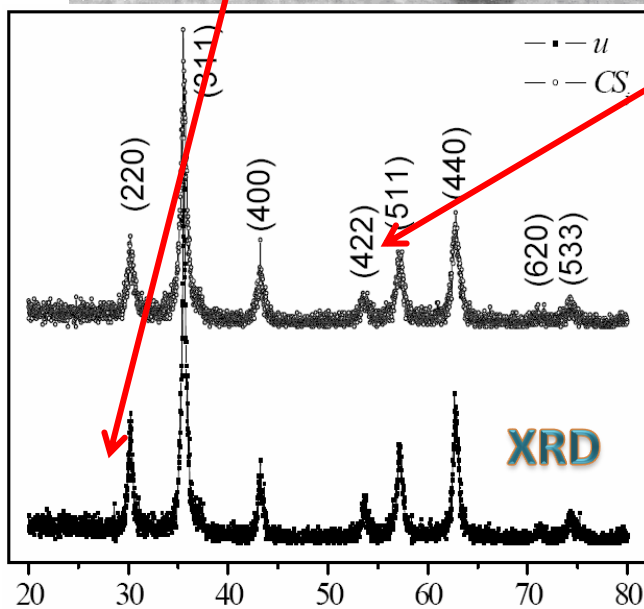
uncoated



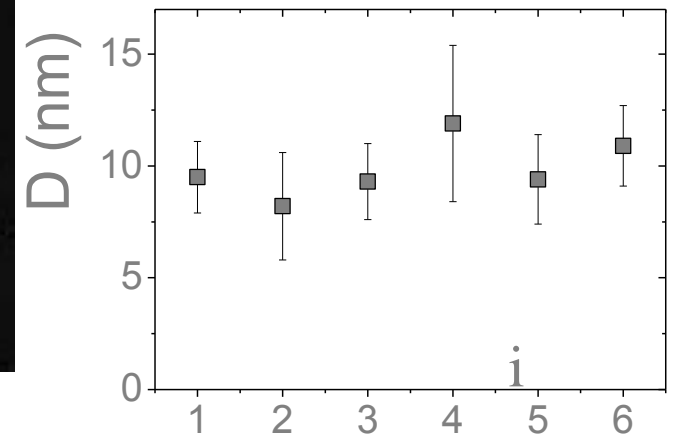
CA coated



TEM

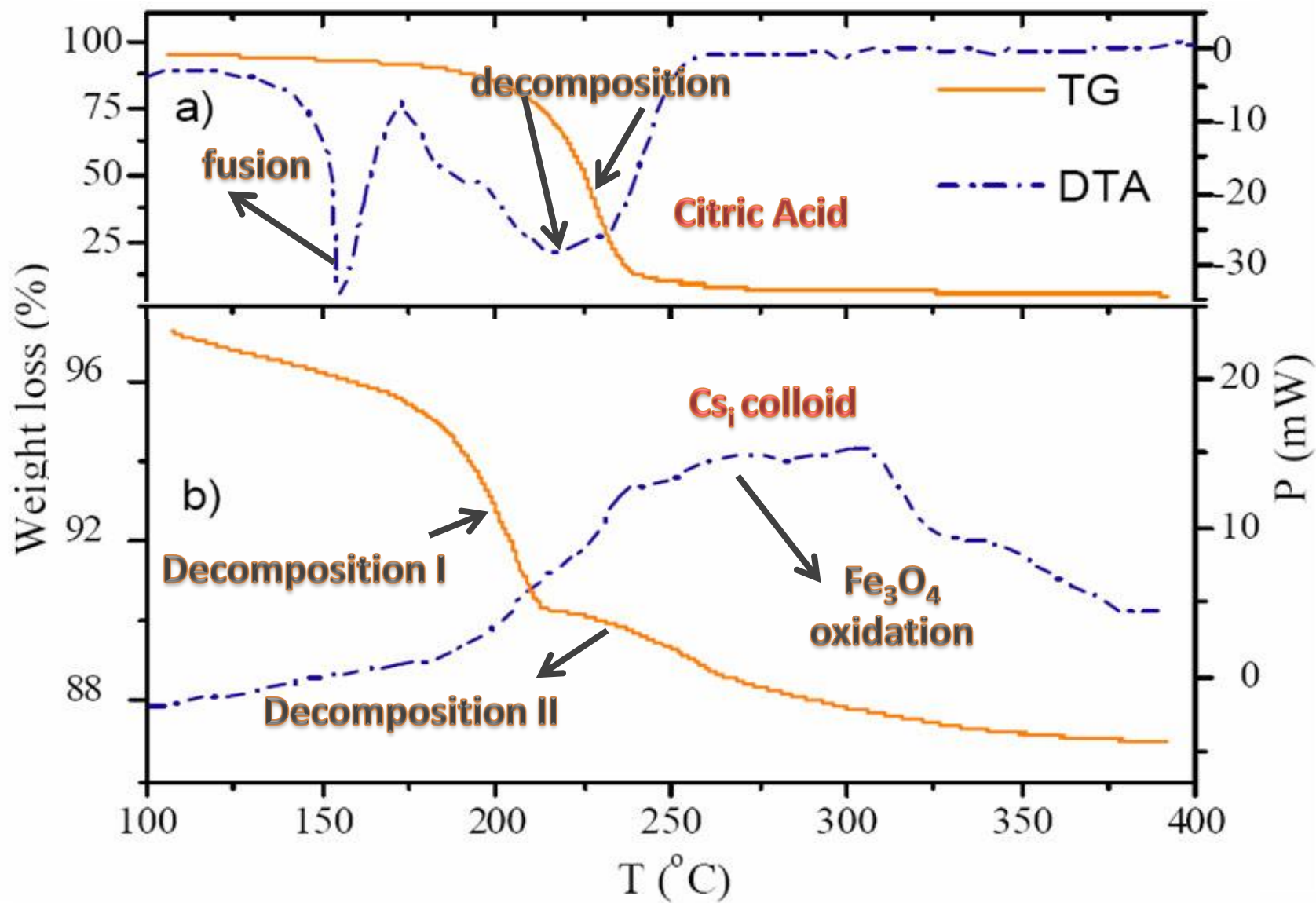


$2\theta$

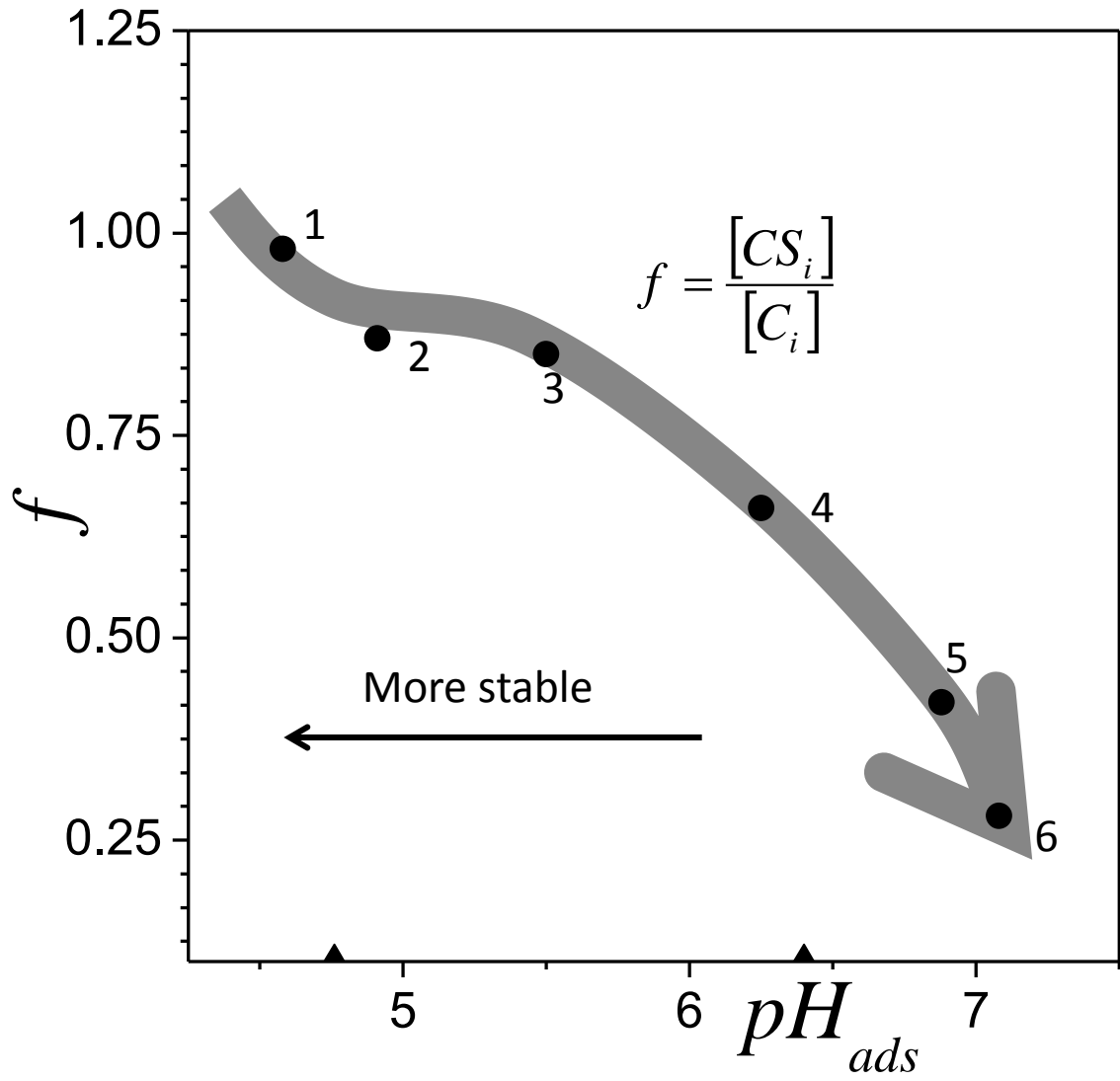


X-LAW3M

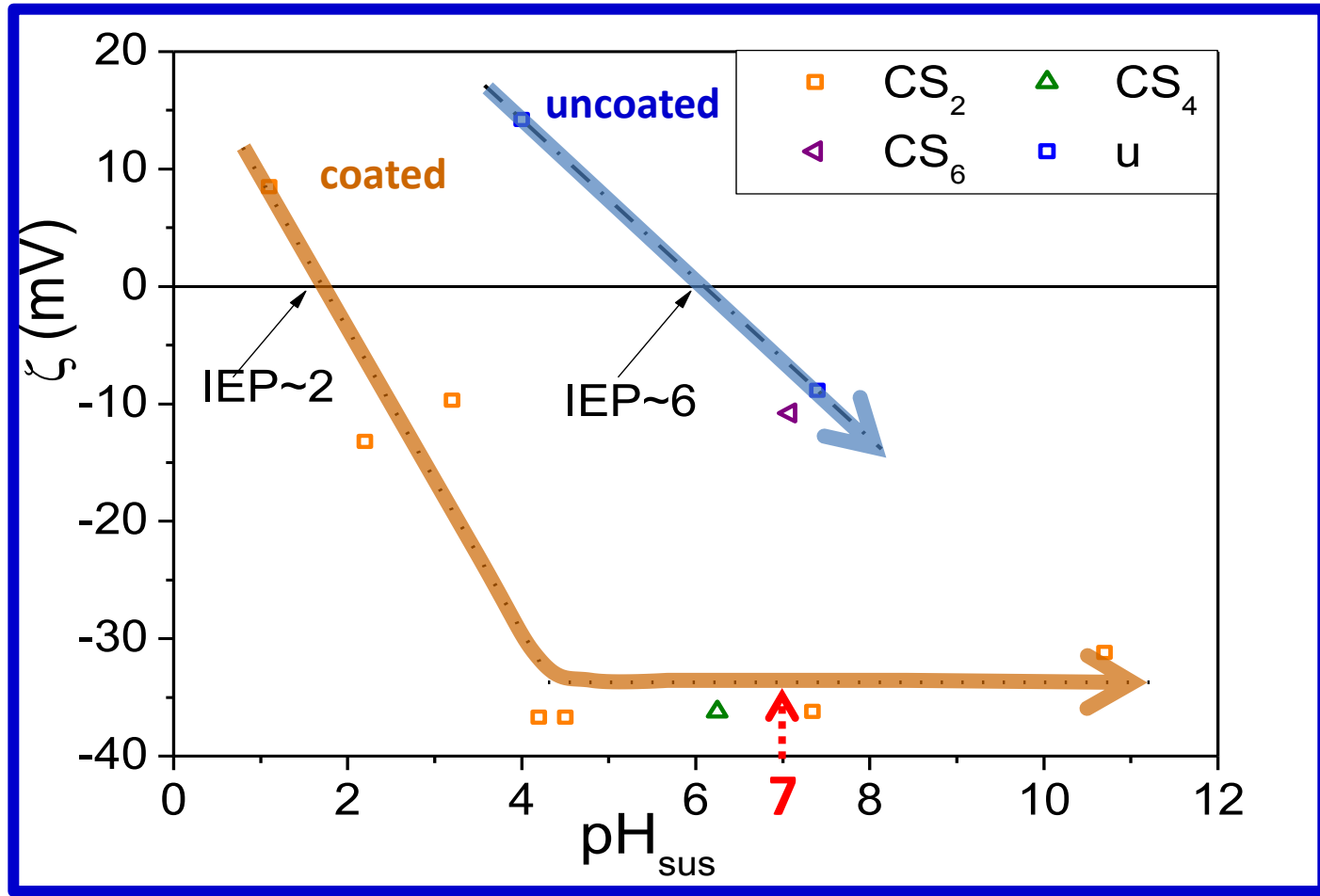
# Thermal stability, TG-DTA



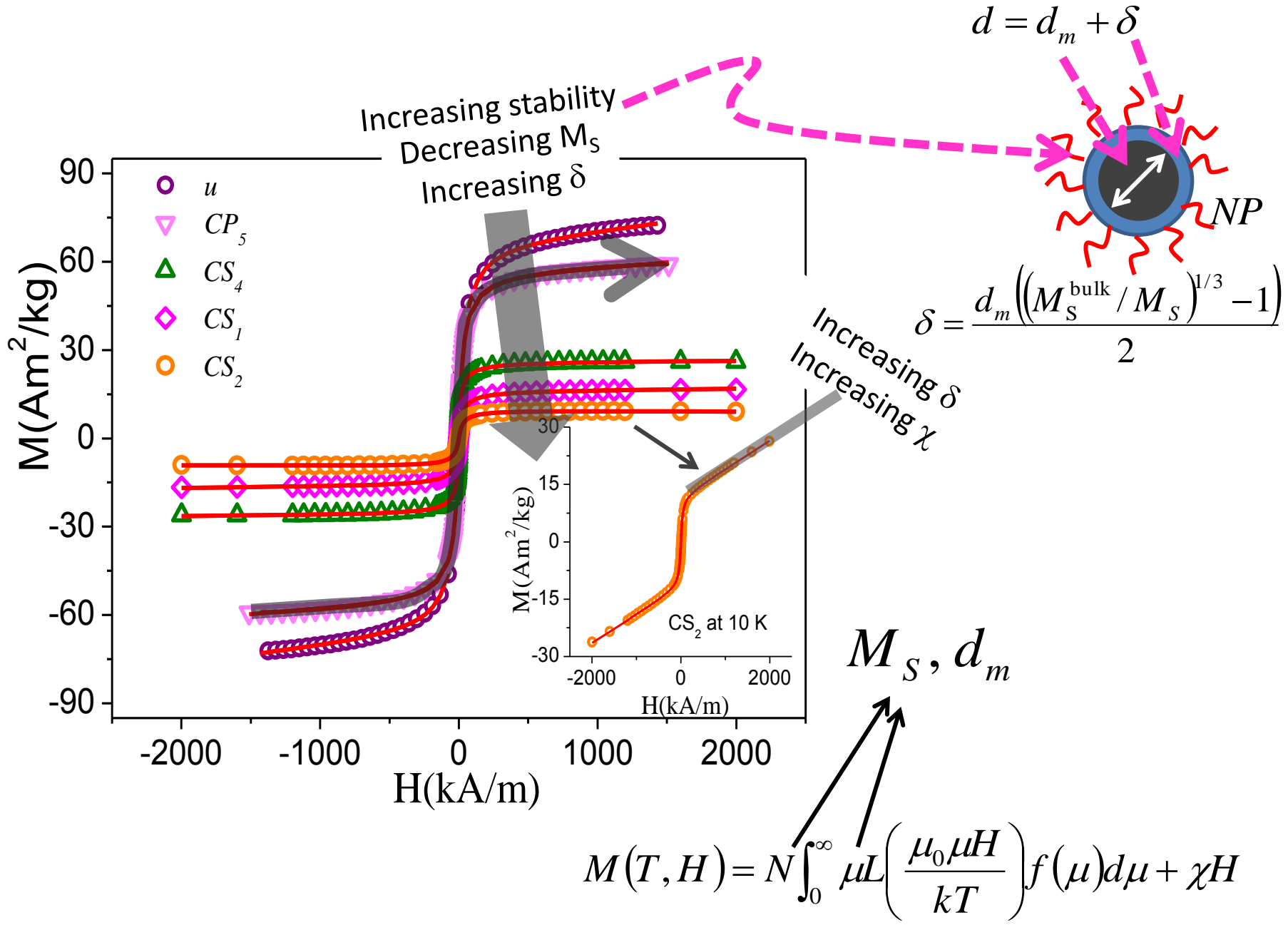
# FF stability vs adsorption pH



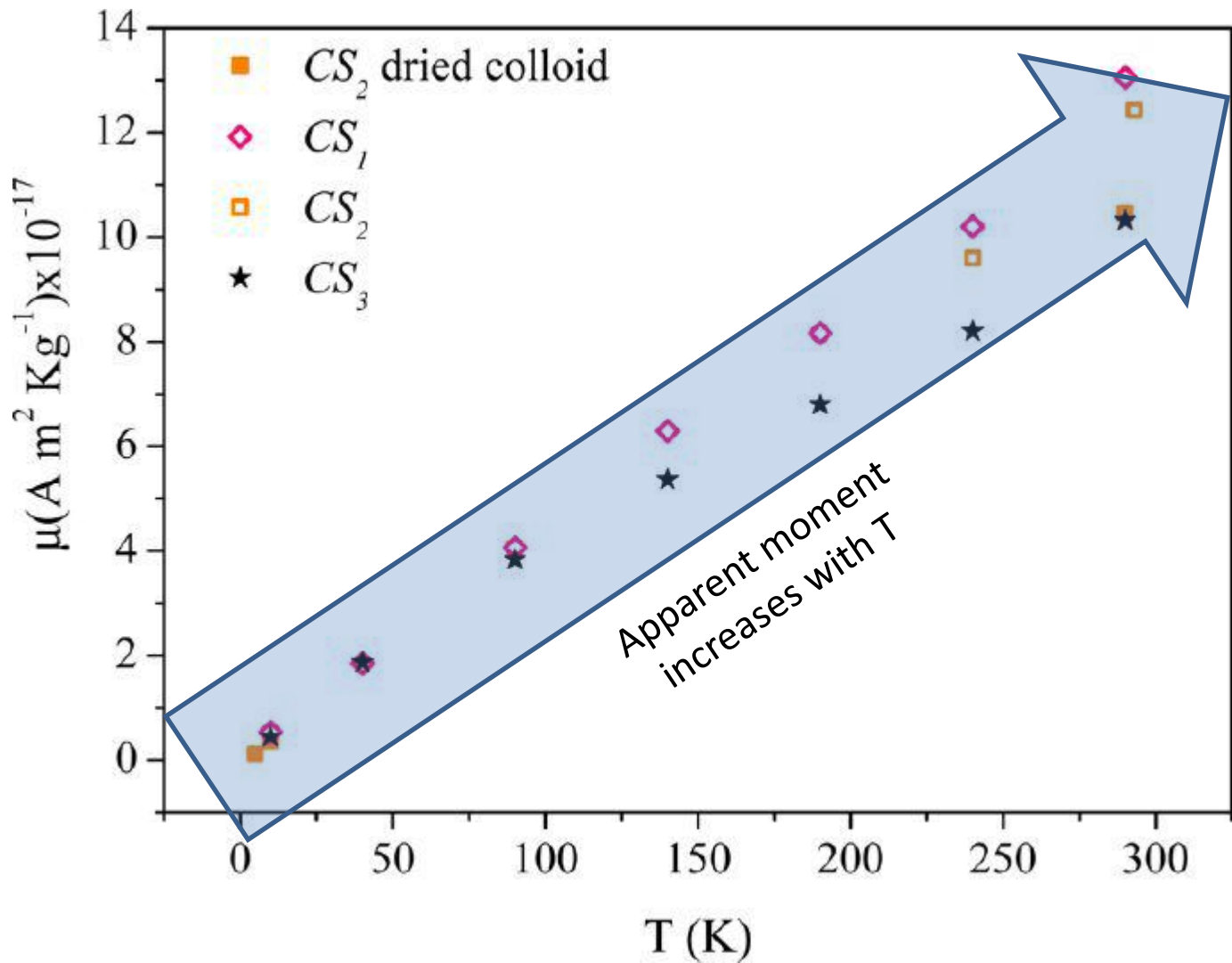
# Z potential, surface charge



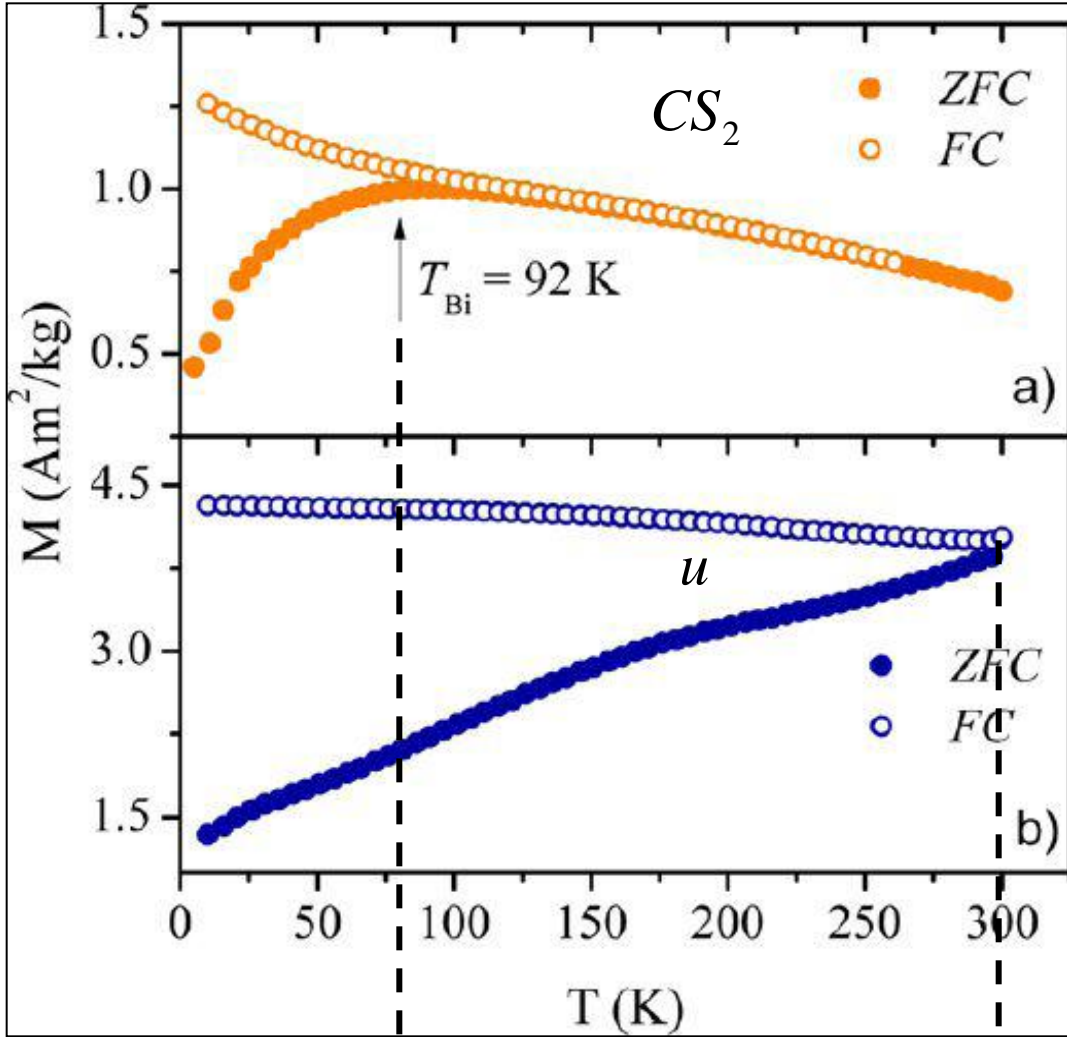
# M vs H on frozen FF, stability – dead layer



# $\mu$ vs T, from M vs H curves on frozen FF, dipolar interactions



# M vs T on frozen FF, ZCF-FC, dipolar interactions



$$E_b \approx K_{\text{eff}} V + \varepsilon$$

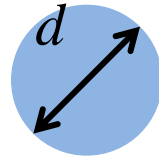
Dormann et al., Advances in Chemical Physics 98; Wiley: New York, 2007.



# M vs T on frozen FF, ZCF-FC, dipolar interactions

$$K_{eff} = K + \frac{6}{d} K_S$$

$$K_S = K_S^0 \tanh(d / \lambda)$$



$$K_S^0 \approx 2 \times 10^{-5} \text{ J / m}^2$$

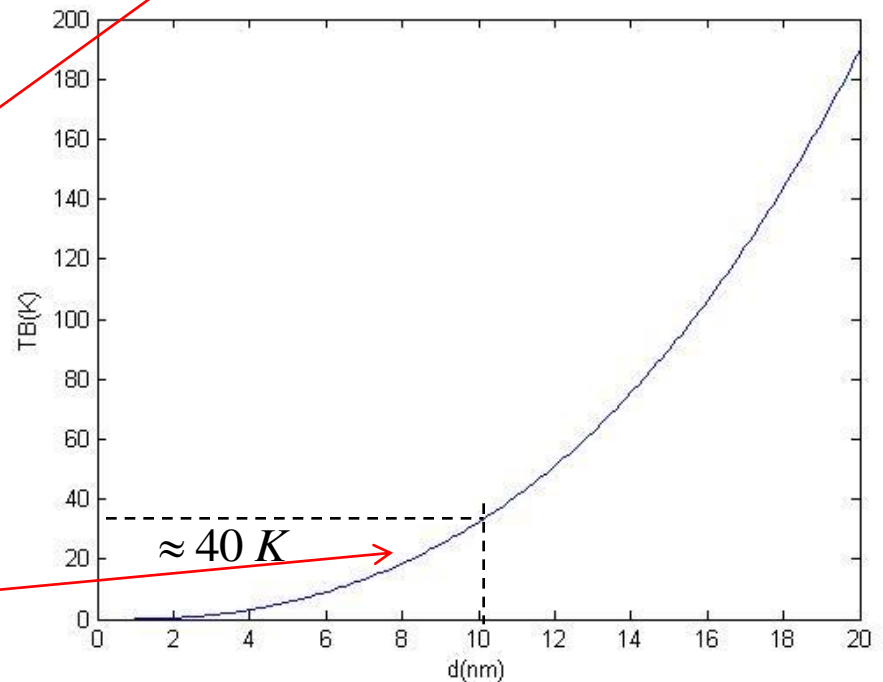
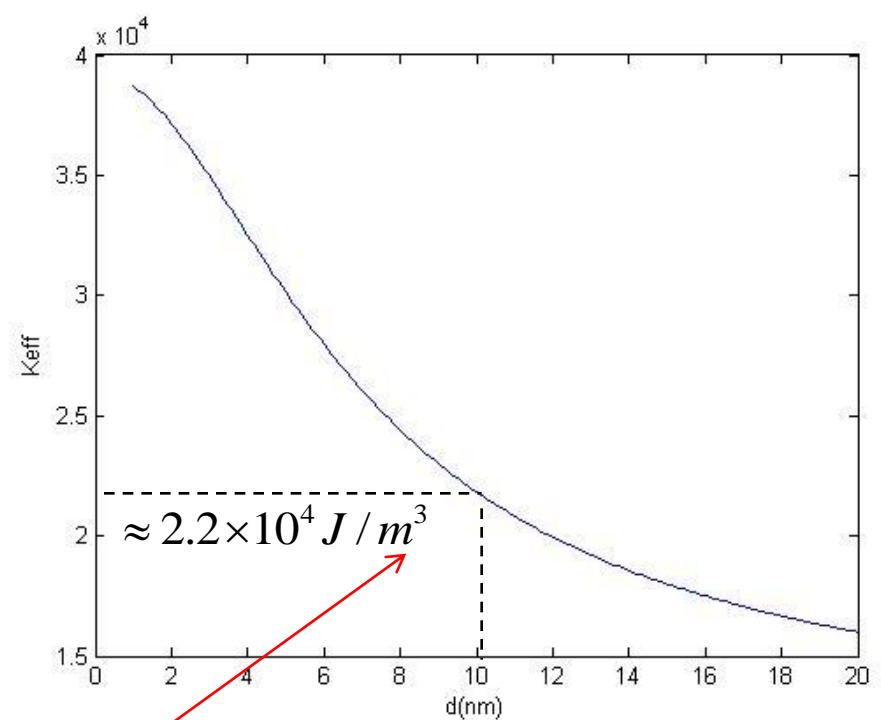
$$\lambda \approx 4.1 \times 10^{-9} \text{ m}$$

Gilmore *et al.* J. Appl. Phys. 97, 10B301 (2005)

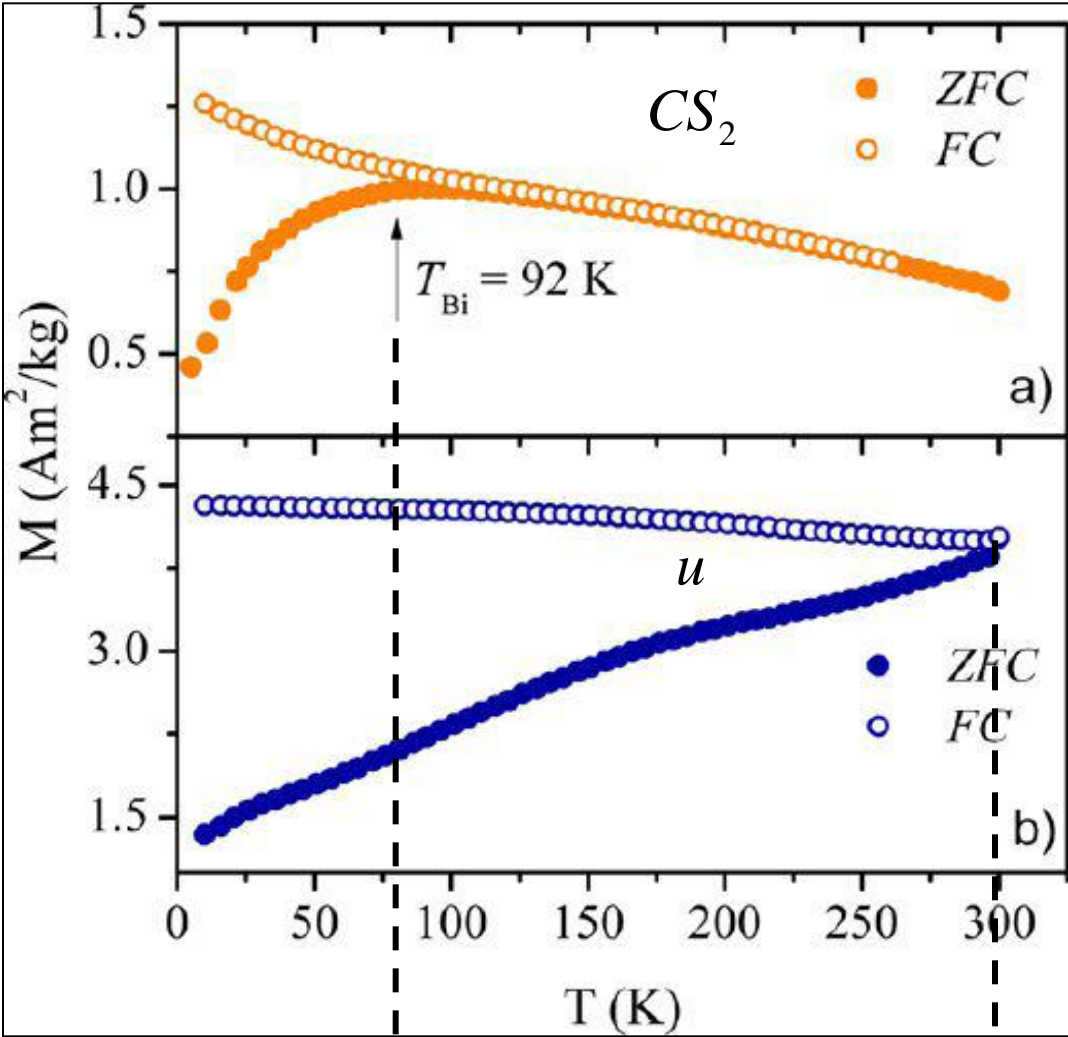
bulk  $K = 1 \times 10^4 \text{ J / m}^3$

$$K_{eff} \approx 2 \times 10^4 \text{ J / m}^3$$

$$T_B^{ni}(d) \approx \frac{K_{eff}(d)V}{25.6k}$$



# M vs T on frozen FF, ZCF-FC, dipolar interactions



$$\epsilon \approx k\Delta T_B$$

$$\epsilon_{\text{CS}_2} \approx 0.7 \times 10^{-21} \text{ J / NP}$$

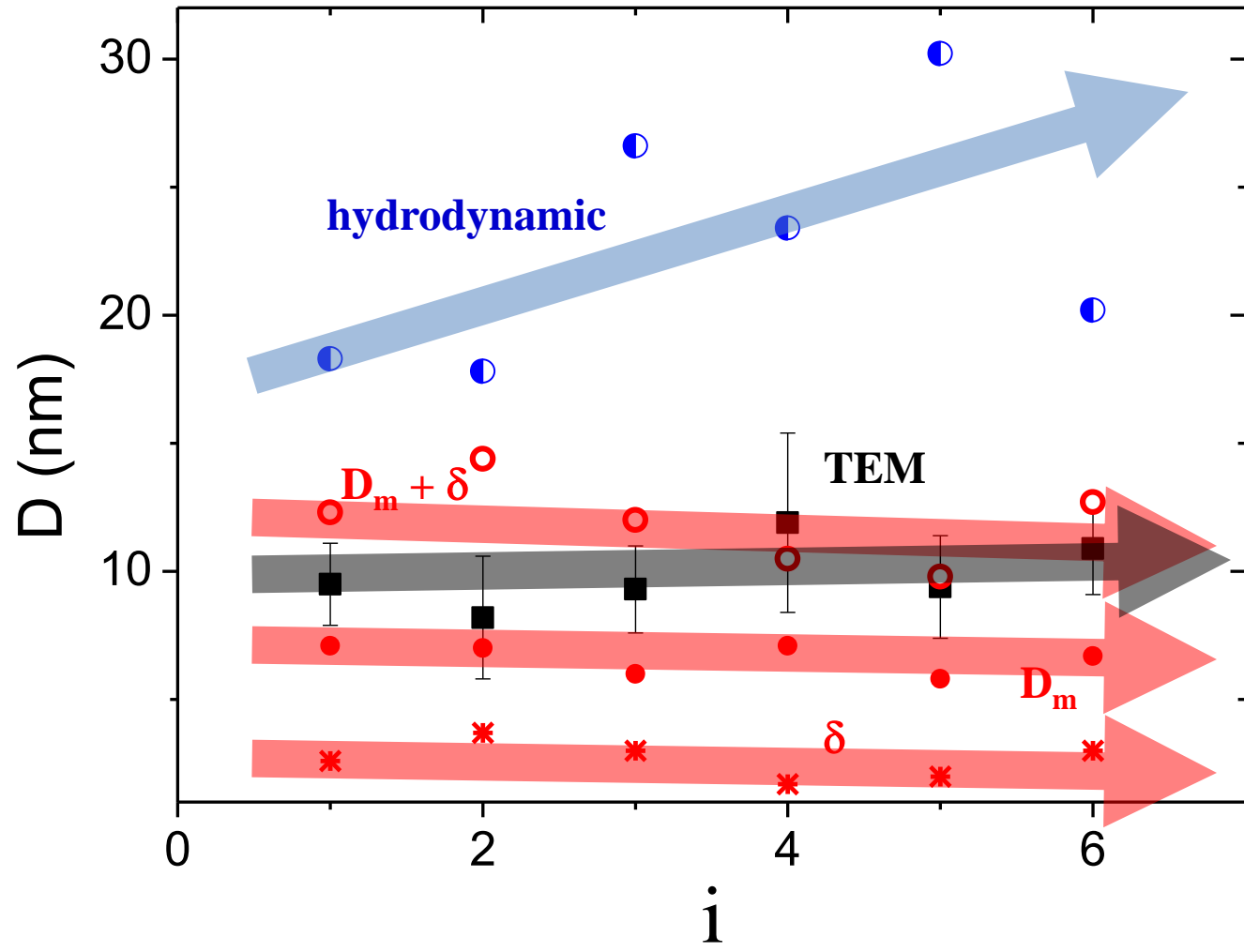
$$\epsilon_u \approx 3.6 \times 10^{-21} \text{ J / NP}$$

at RT

$$kT \approx 4.1 \times 10^{-21} \text{ J}$$

$$\Delta T_B^{u-CS} \approx 208 \text{ K}$$

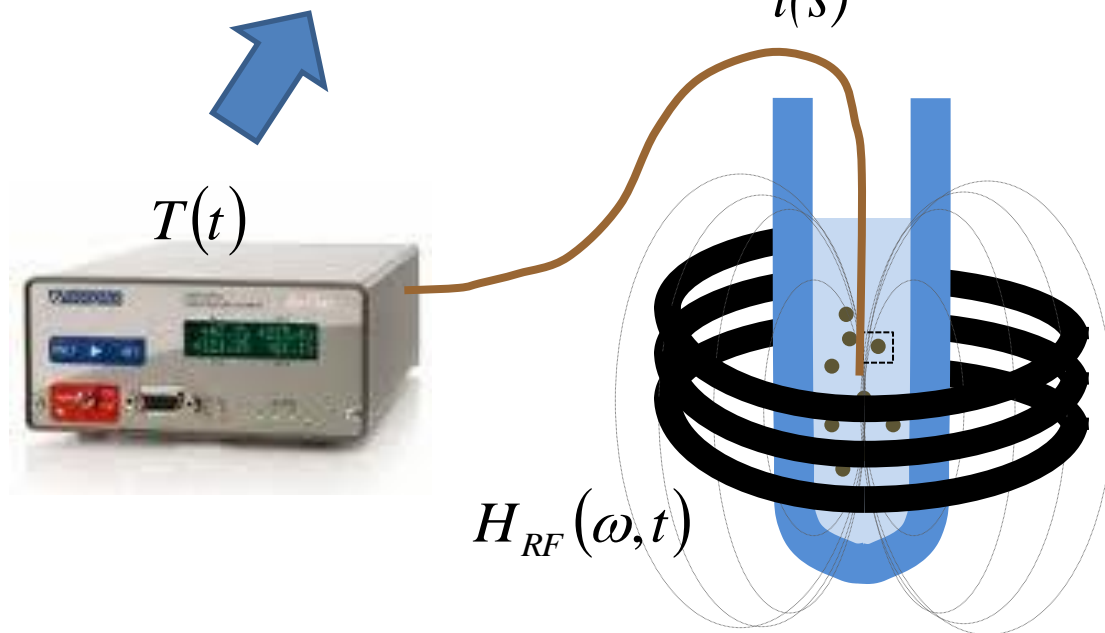
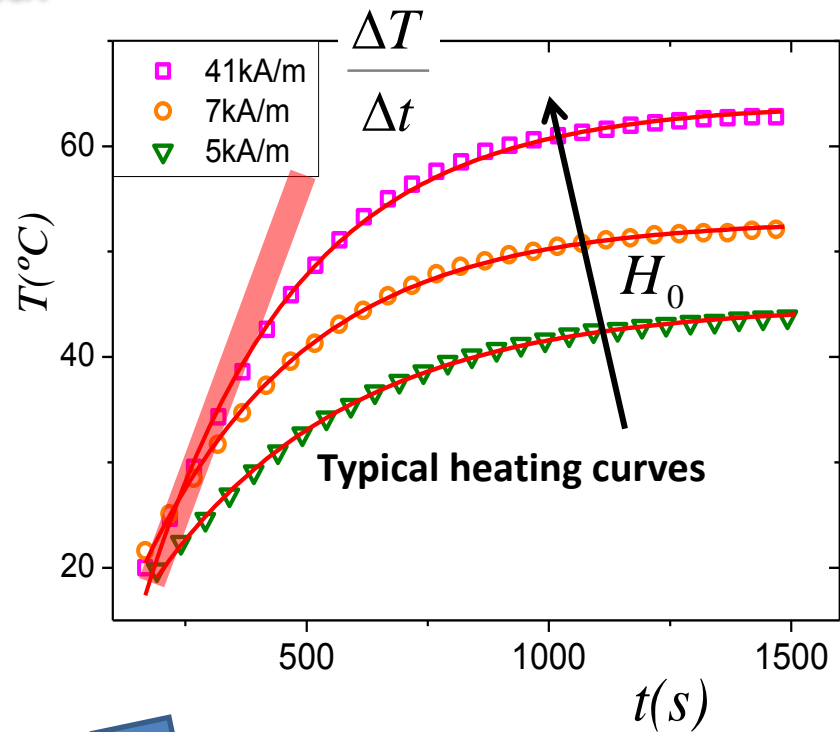
# NP sizes



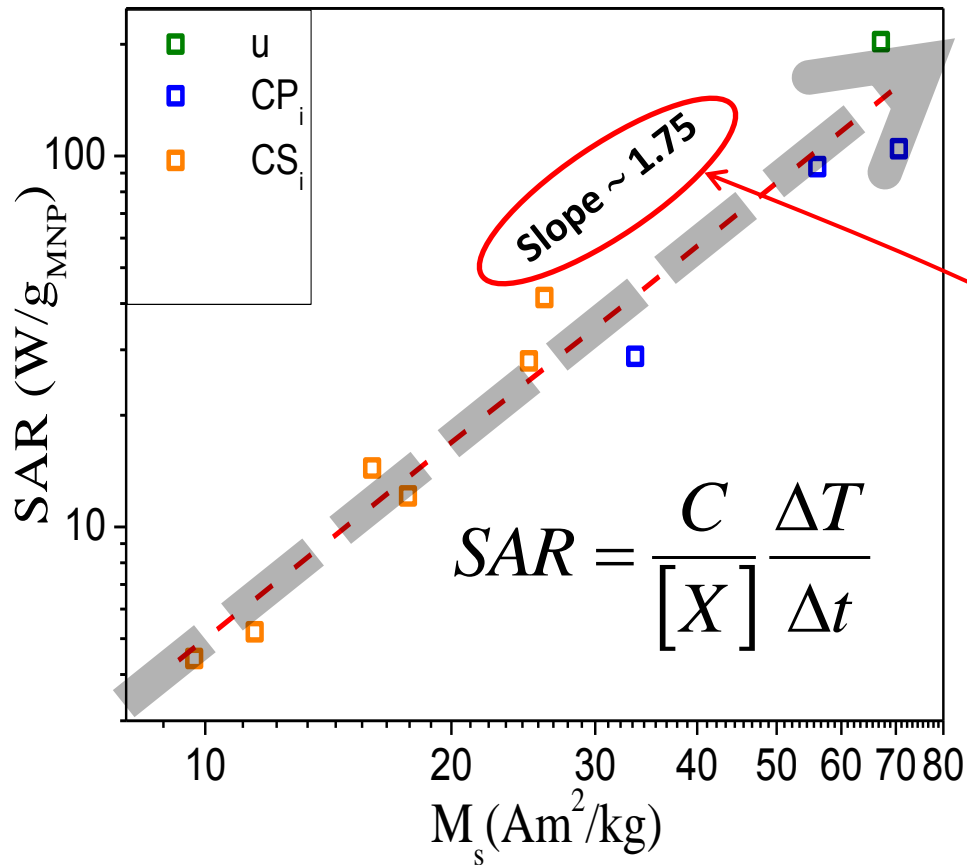
# SAR, experimental



# SAR, experimental



# SAR, Ms dependence



$$SAR = \frac{\pi \mu_0 H_0^2 f_{RF}}{\rho} \int_0^\infty \chi''(\tau(d)) f(d) dd$$

$$\chi''(\omega) = \chi_0 \frac{\omega \tau}{1 + (\omega \tau)^2}$$

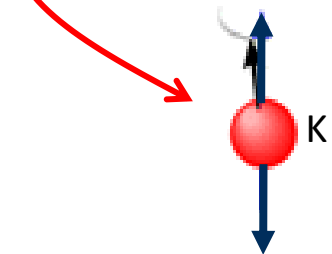
$$\chi_0 \approx \frac{\mu_0 \rho^2 M_s^2 V}{3kT}$$

# SAR, moment relaxation mechanism

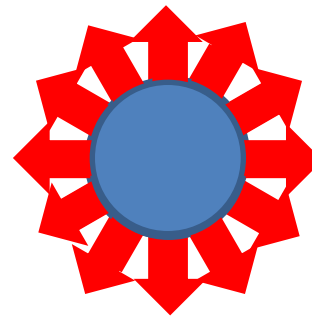
$$SAR \propto \chi''(\omega) = \chi_0 \frac{\omega\tau}{1 + (\omega\tau)^2}$$

$$\tau = \begin{cases} \tau_N \approx \tau_0 e^{\sigma}, & \sigma = E_b / kT \\ \tau_B \approx 3\eta V_H / kT \end{cases}$$

$$\tau^{-1} = \tau_N^{-1} + \tau_B^{-1}$$



*Néel*



*Brown*

# Conclusions I

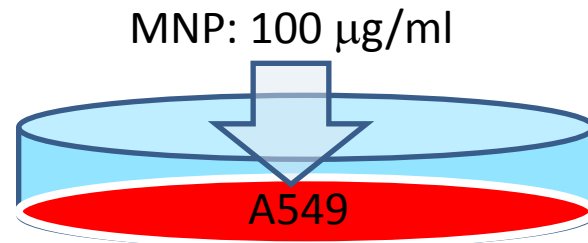
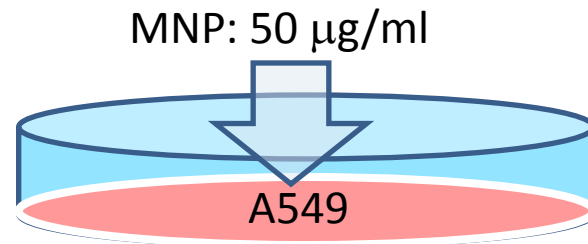
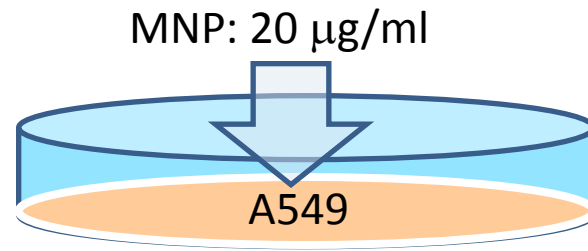
Citric acid coating: highly stable FFs

High influence of coating on dead layer thickness and Ms

High influence of dipolar interactions on SAR

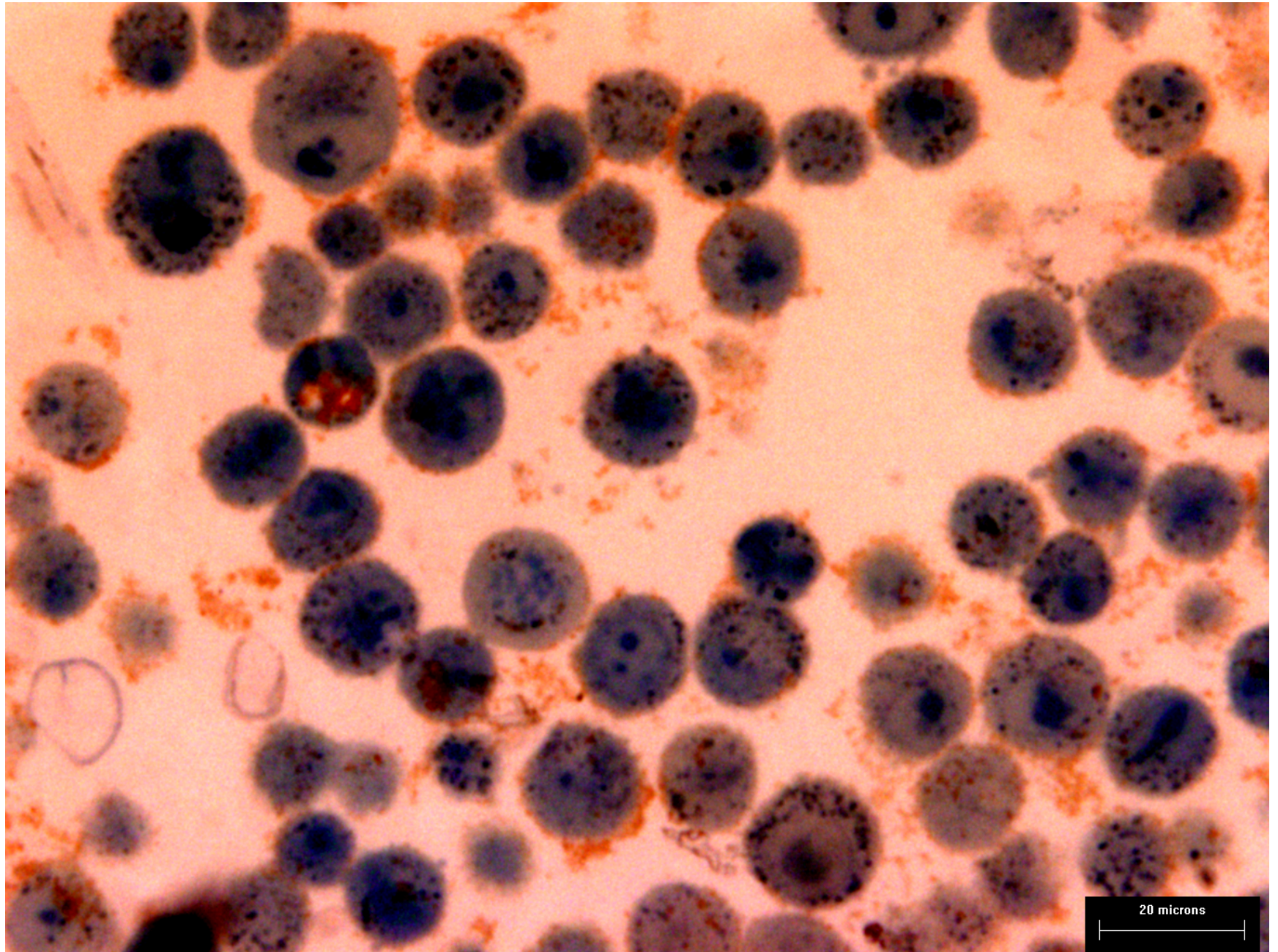


# In vitro experiments – A549

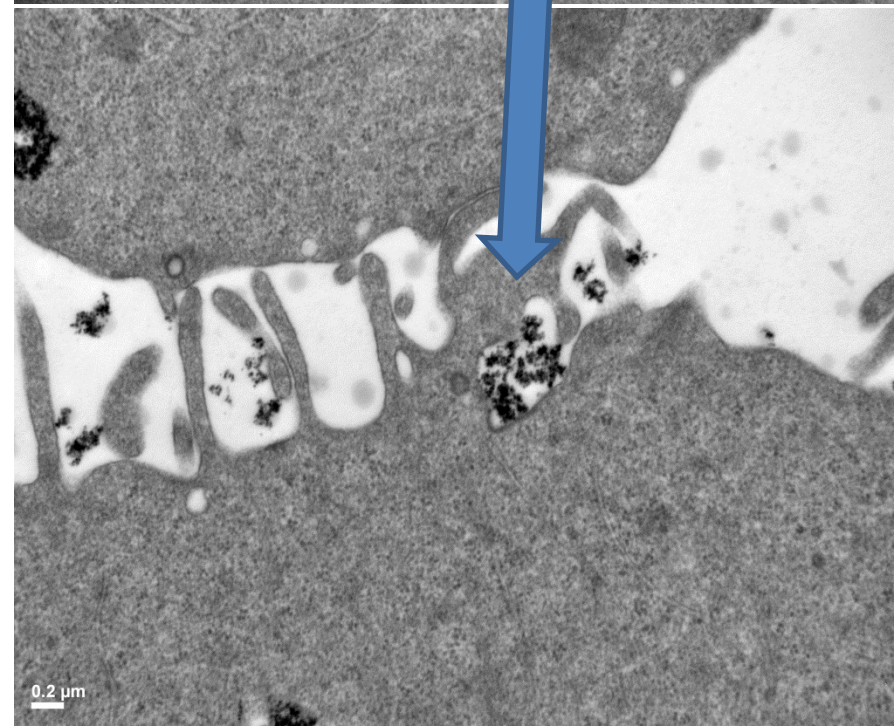
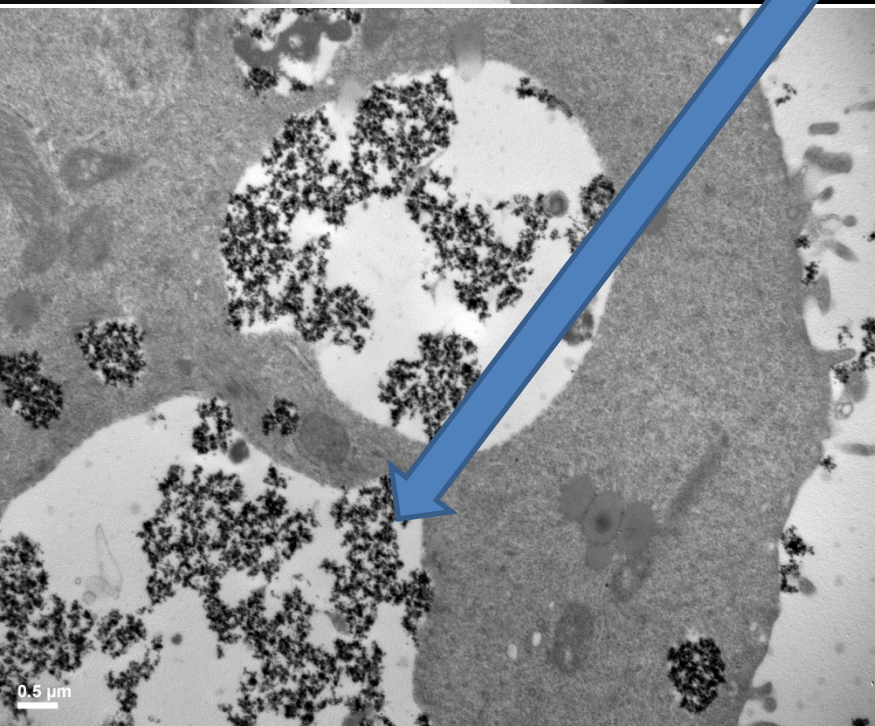
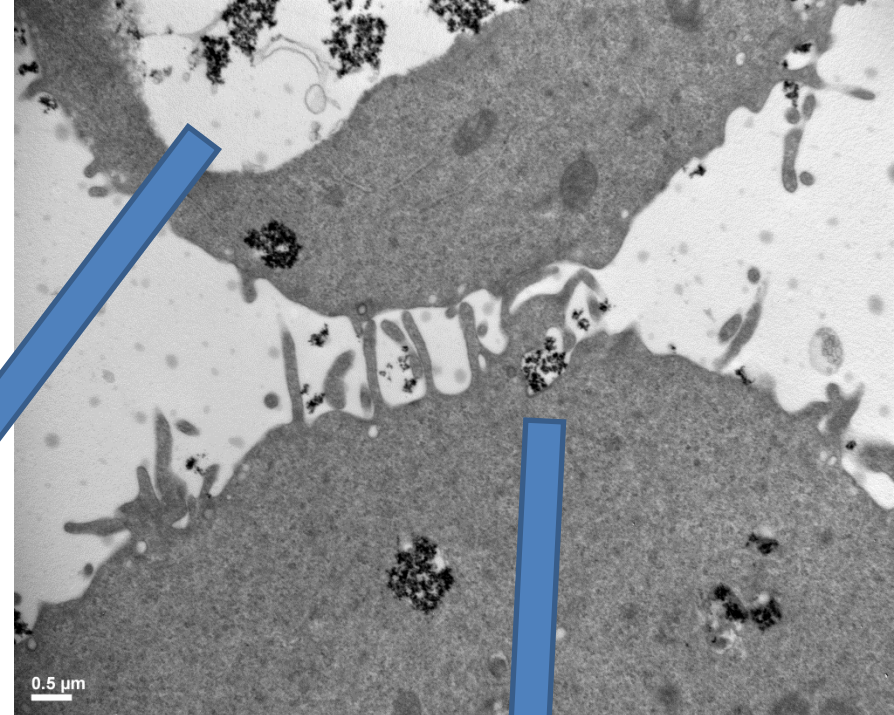
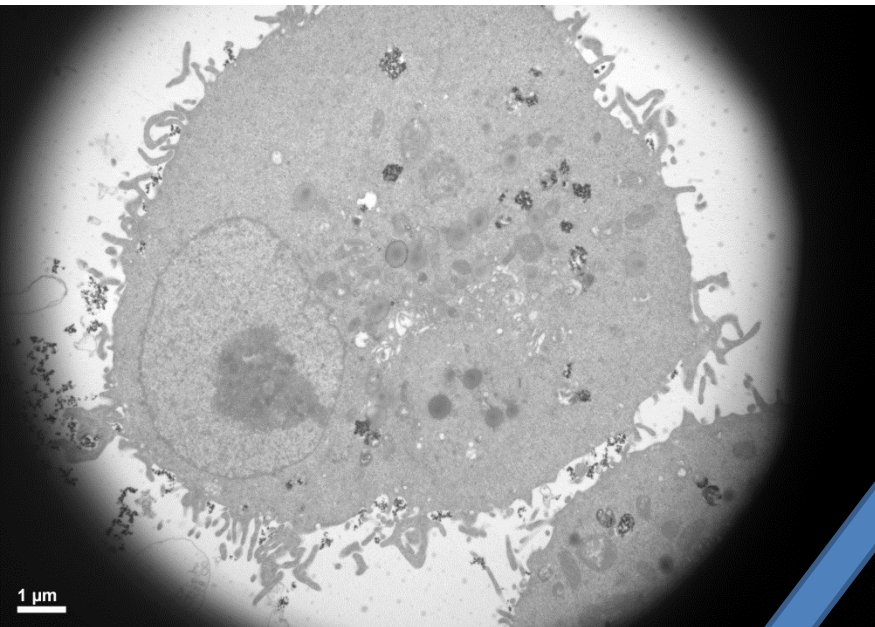


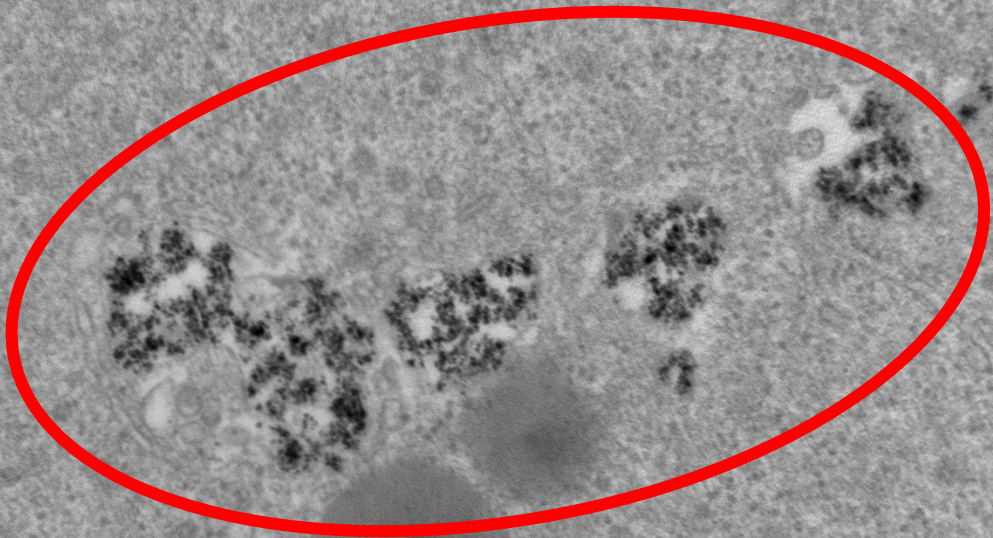
$t \approx 12 h$

# In vitro experiments – A549

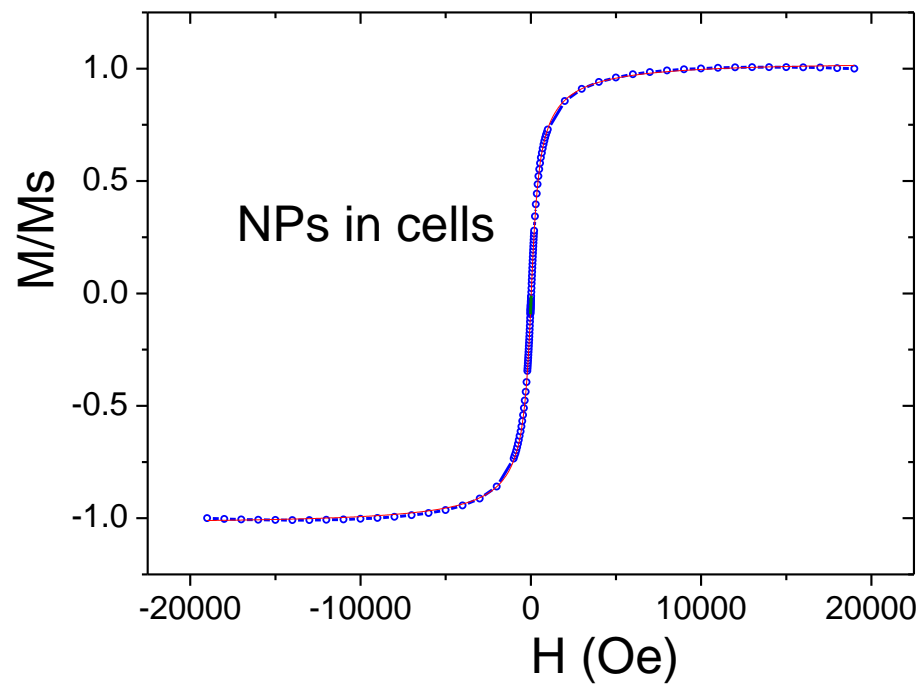
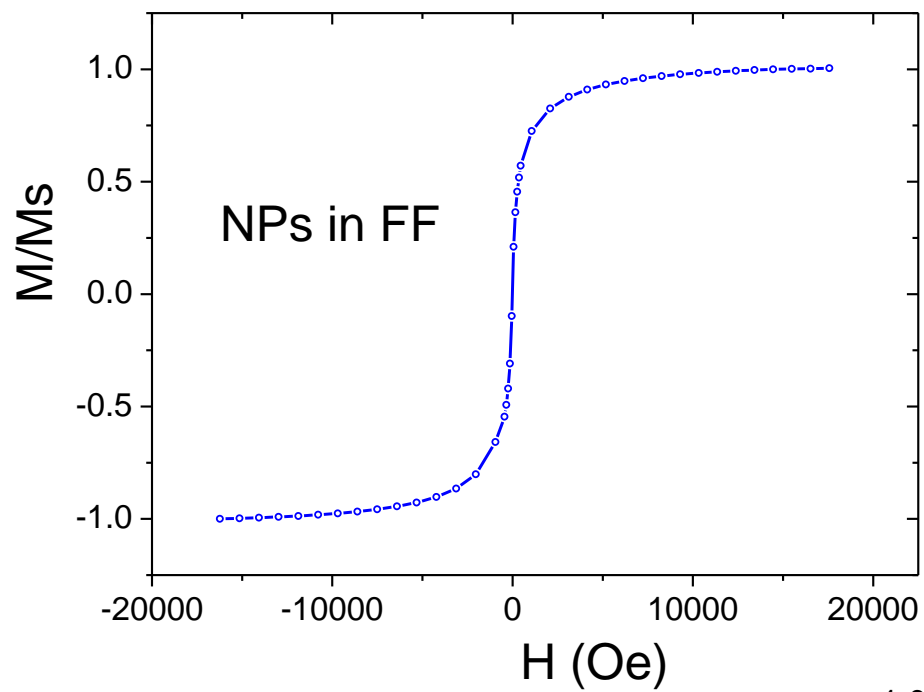


# In vitro experiments – A549

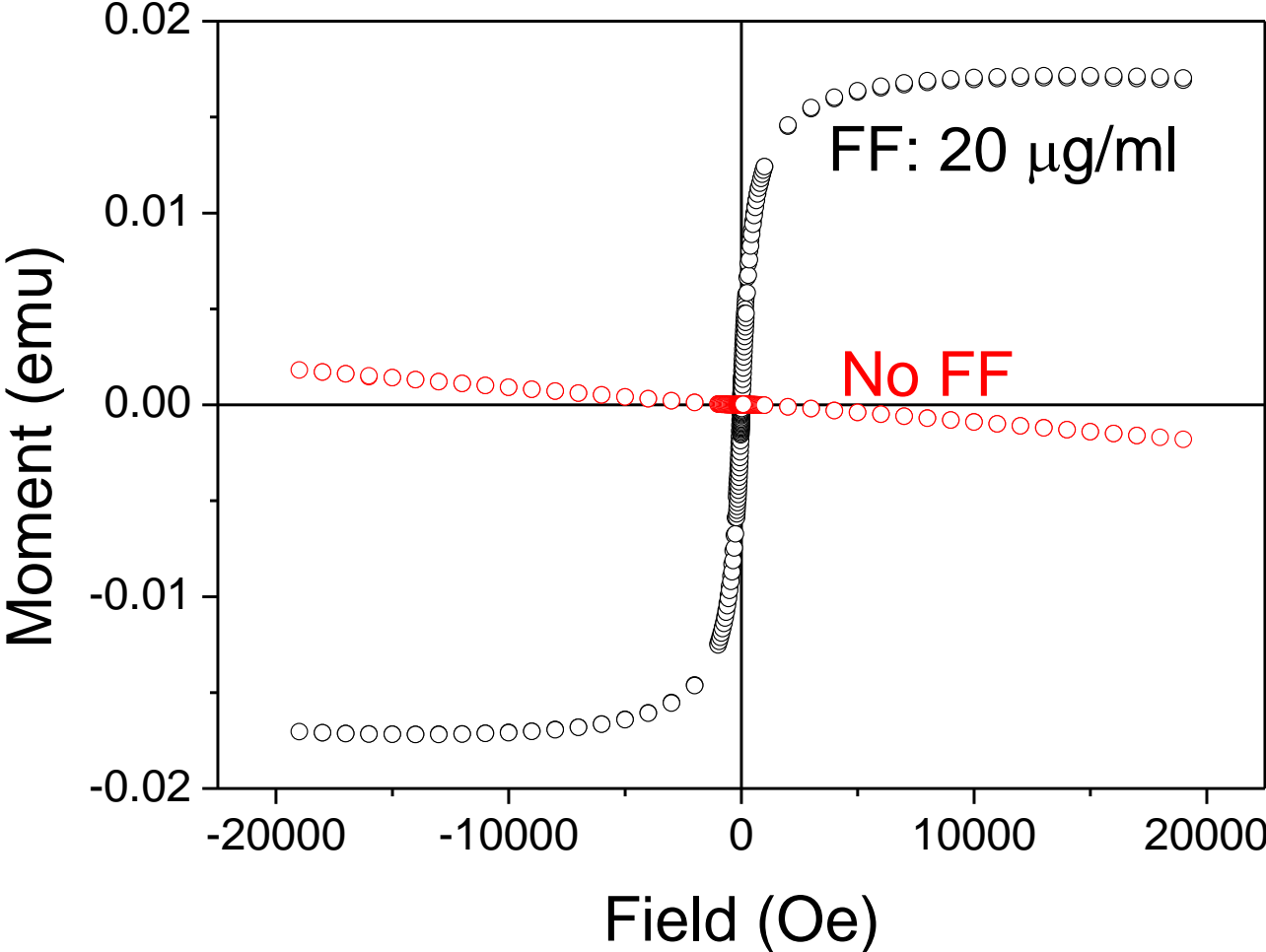
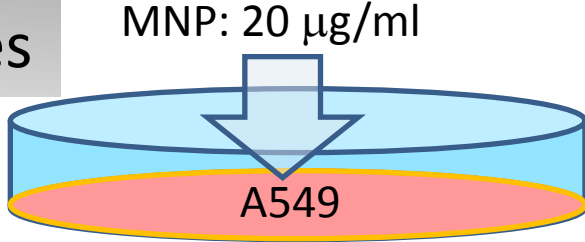




0.2  $\mu\text{m}$

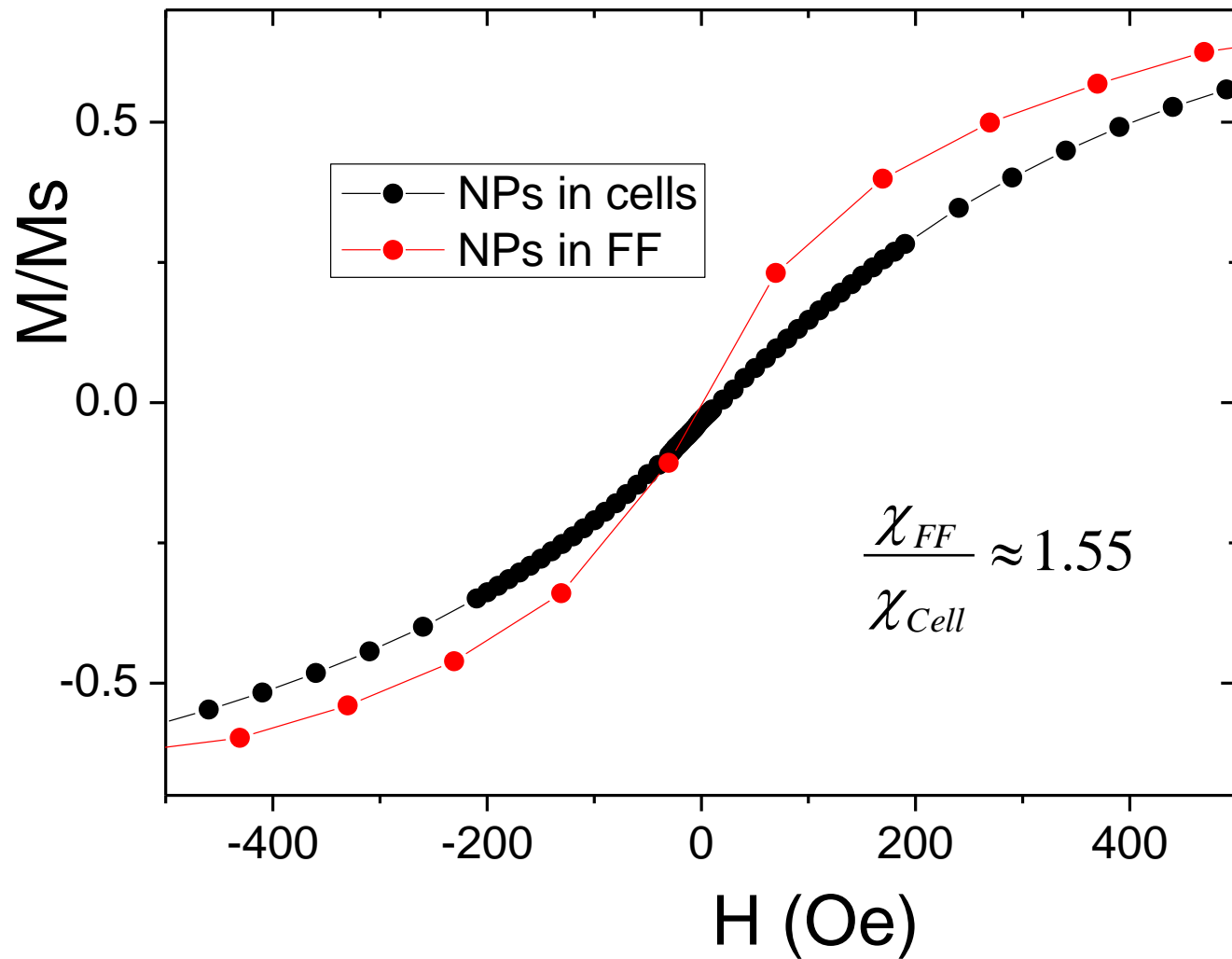


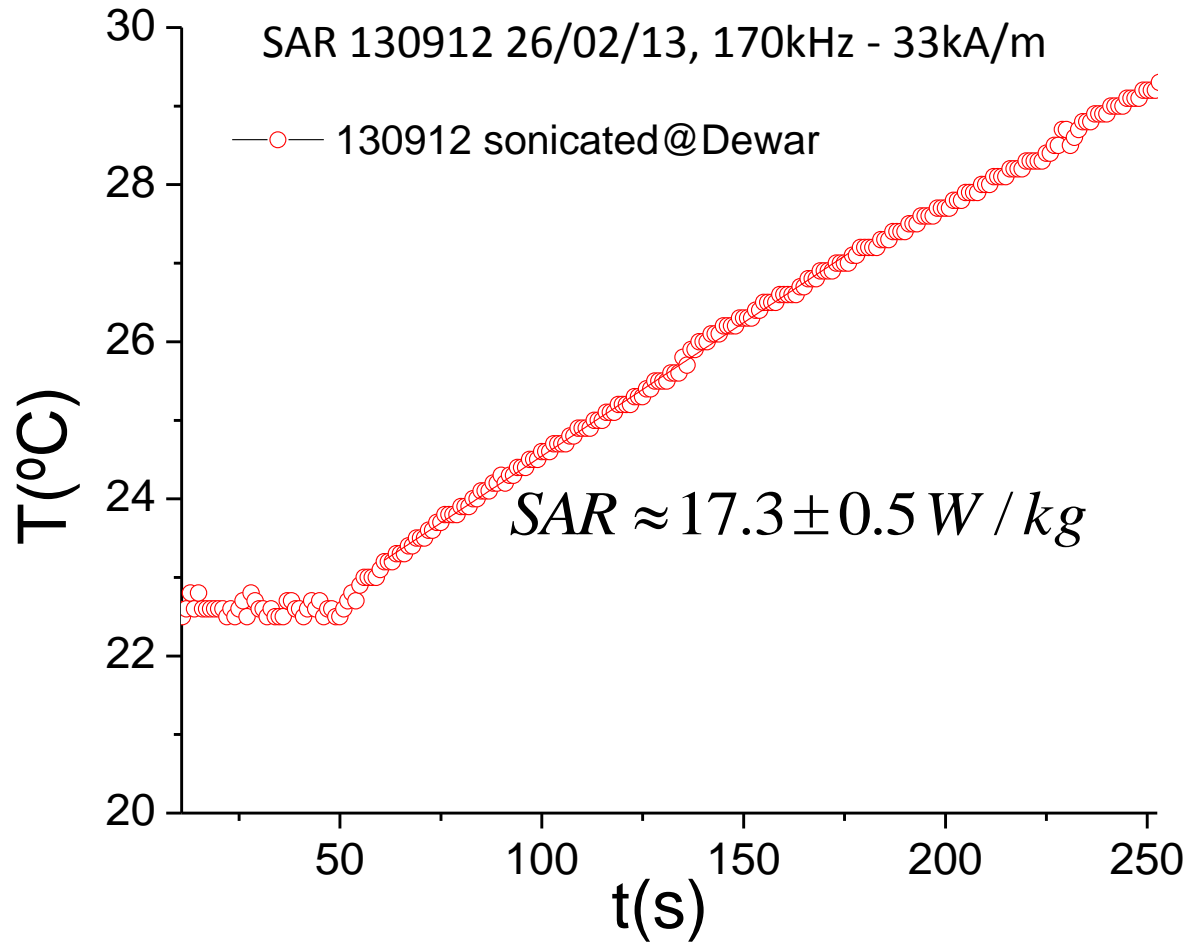
# Internalization efficiency of NPM in cell cultures



15 x 10<sup>6</sup> cells

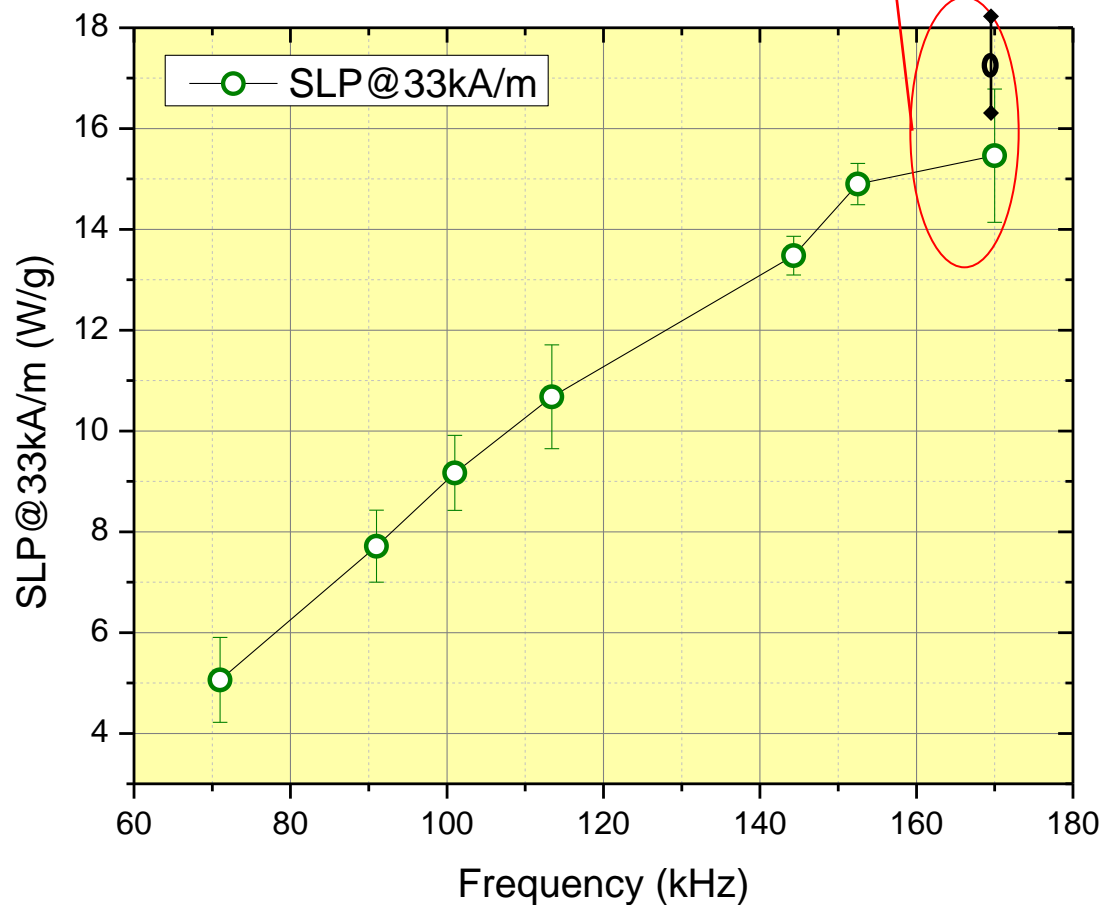
29  $\text{pg}_{\text{mag}}/\text{cell}$







*SAR 130912 (170kHz , 33 kA/m), 15 – 17 W/g*



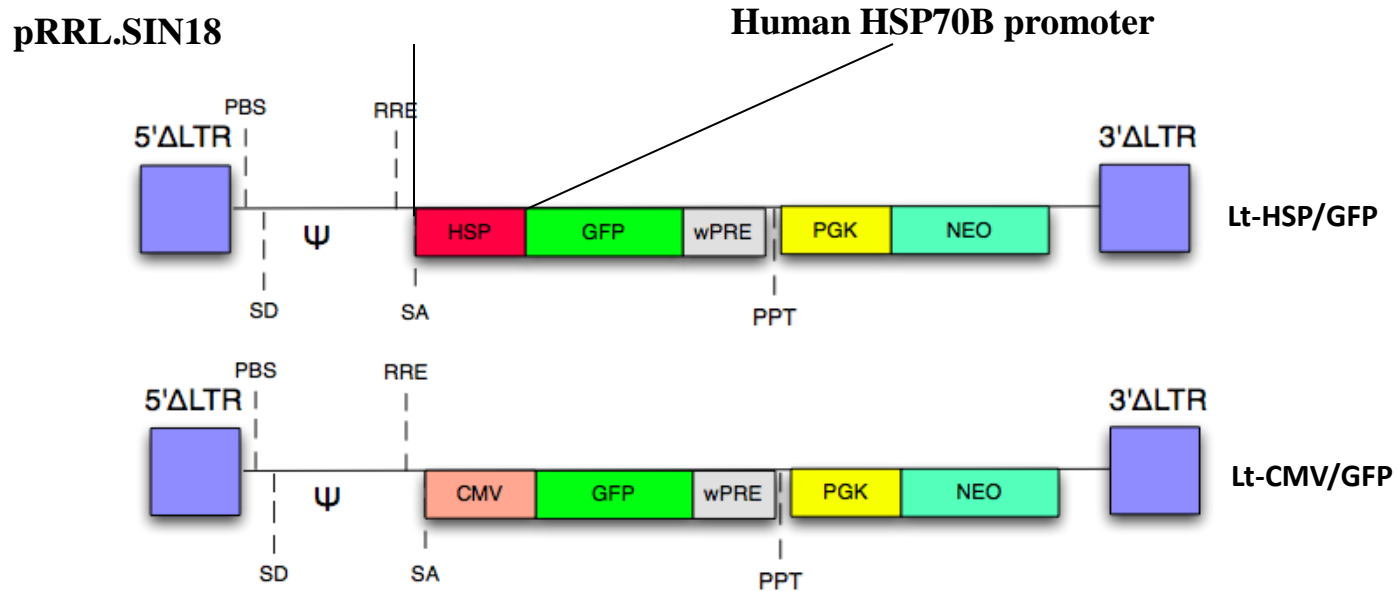
Sample	[X](mg/ml)	Suspension pH	Z Potencial	MS (emu/g)	SAR(W/g) F= 170kHz H=33kA/m
130912pt	8.3	7.44	-13.1mV	27.9	15-17 W/g

# Cellular Thermometer for hyperthermia

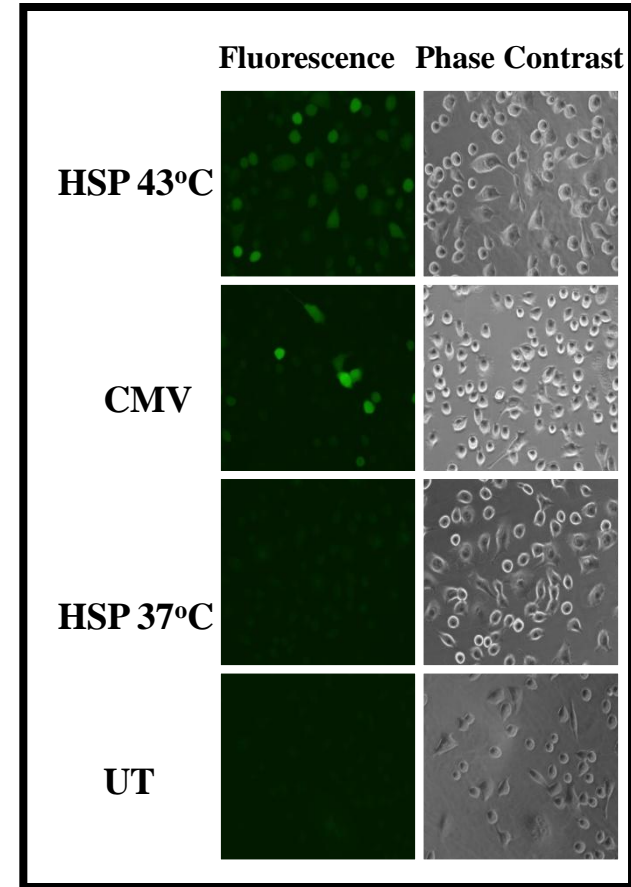
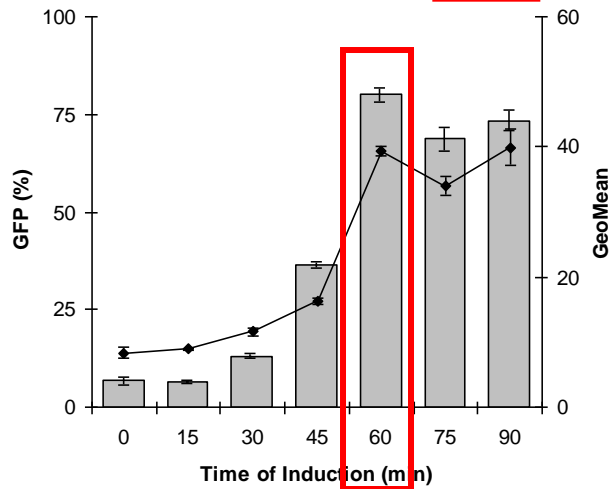
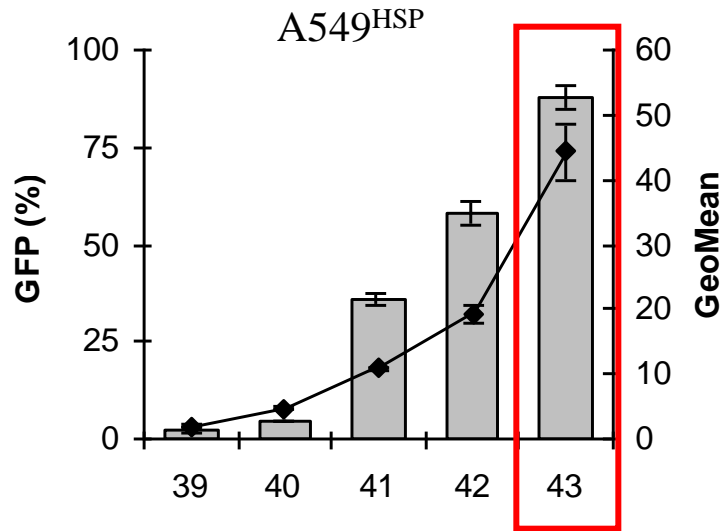
## Lentivectors encoding GFP under the human HSP70b promoter

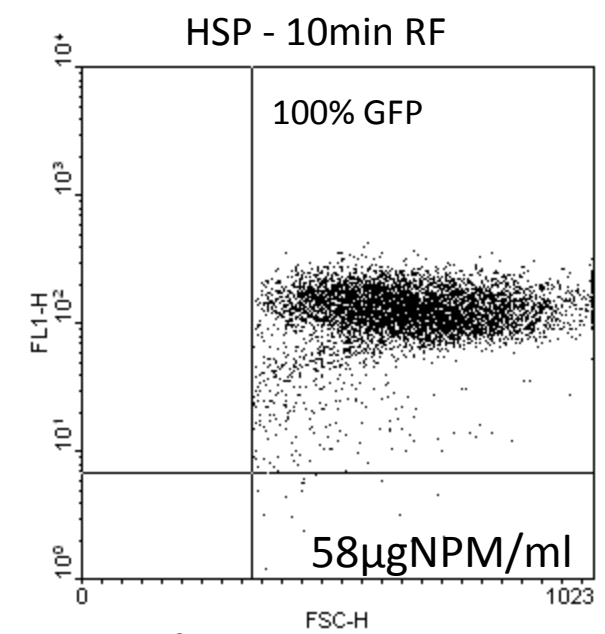
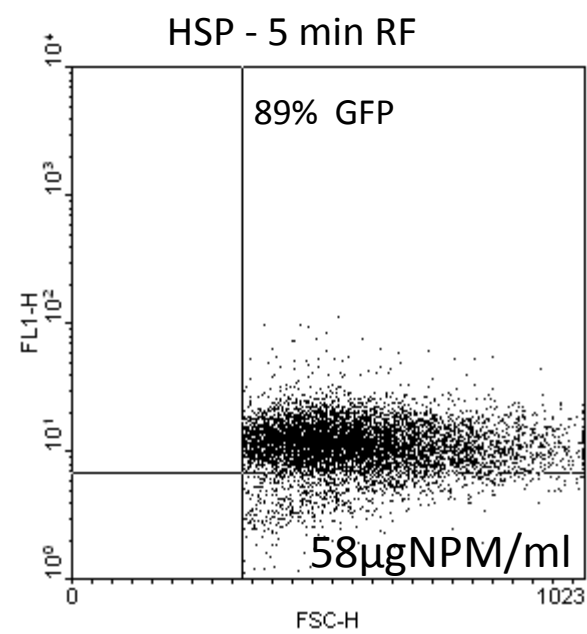
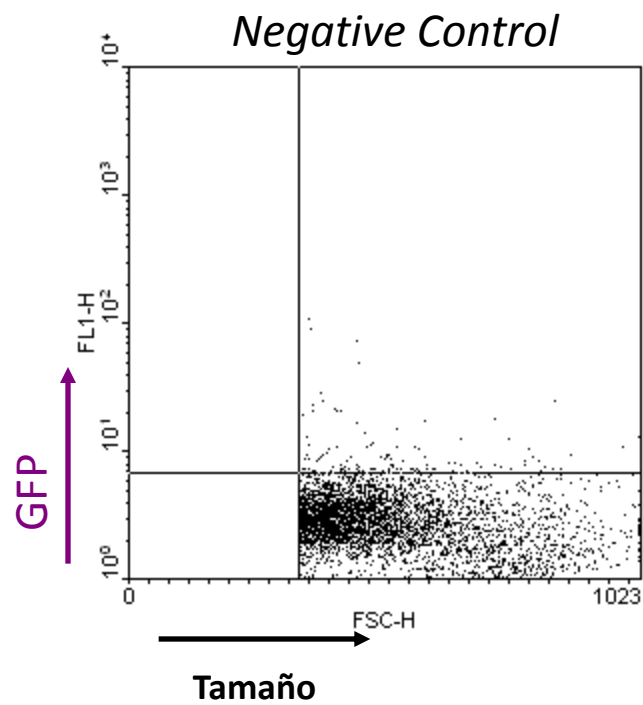
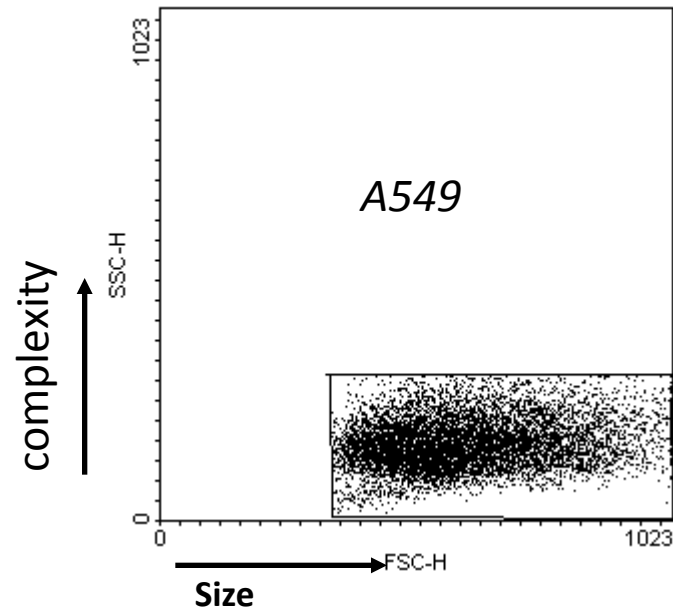
HSP: Same cell line A549 infected with a lentiviral vector containing an inducible promotor (under stress) named HSP60B. It leads to the expression of GFP.

CMV: Same cell line A549 infected with a lentiviral vector containing a constitutive promotor named CMV. Always expresses GFP.



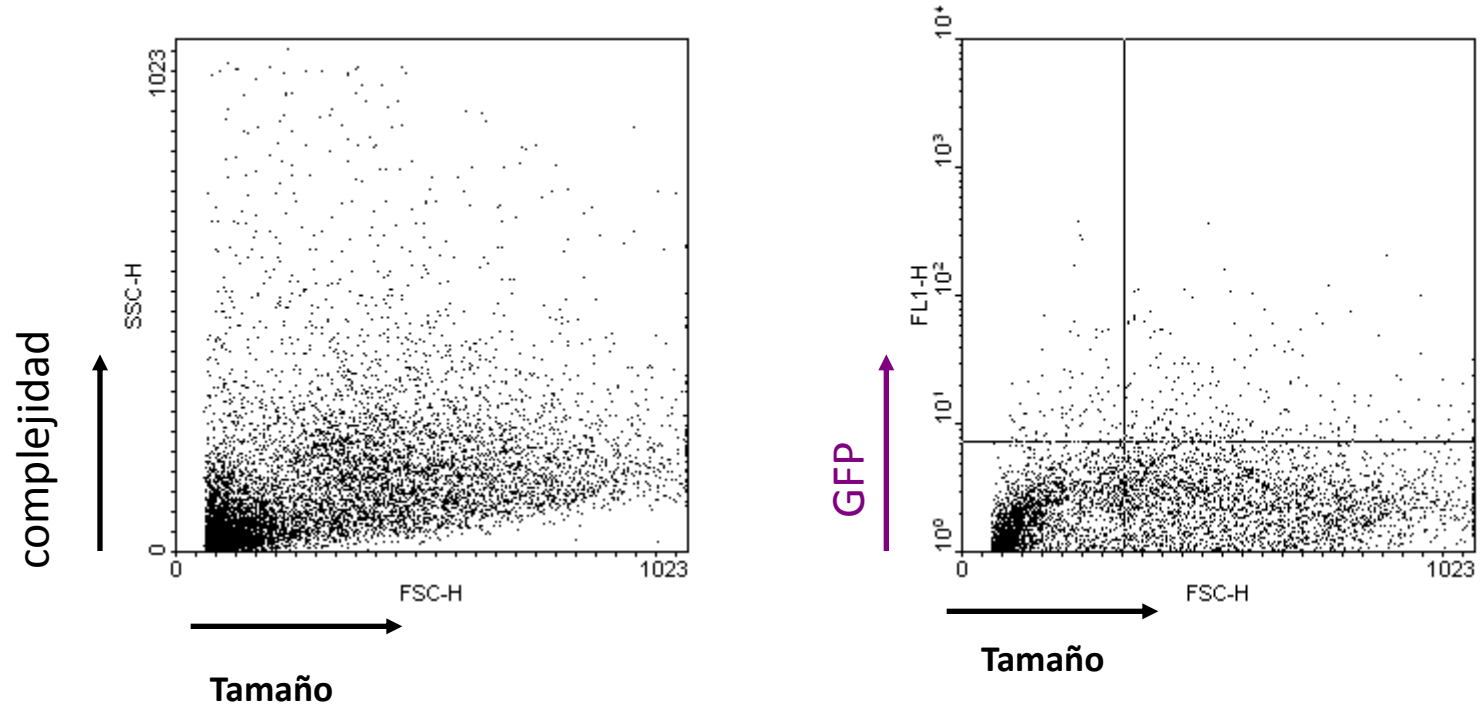
# In vitro Inducibility properties of HSP70b promoter by hyperthermia





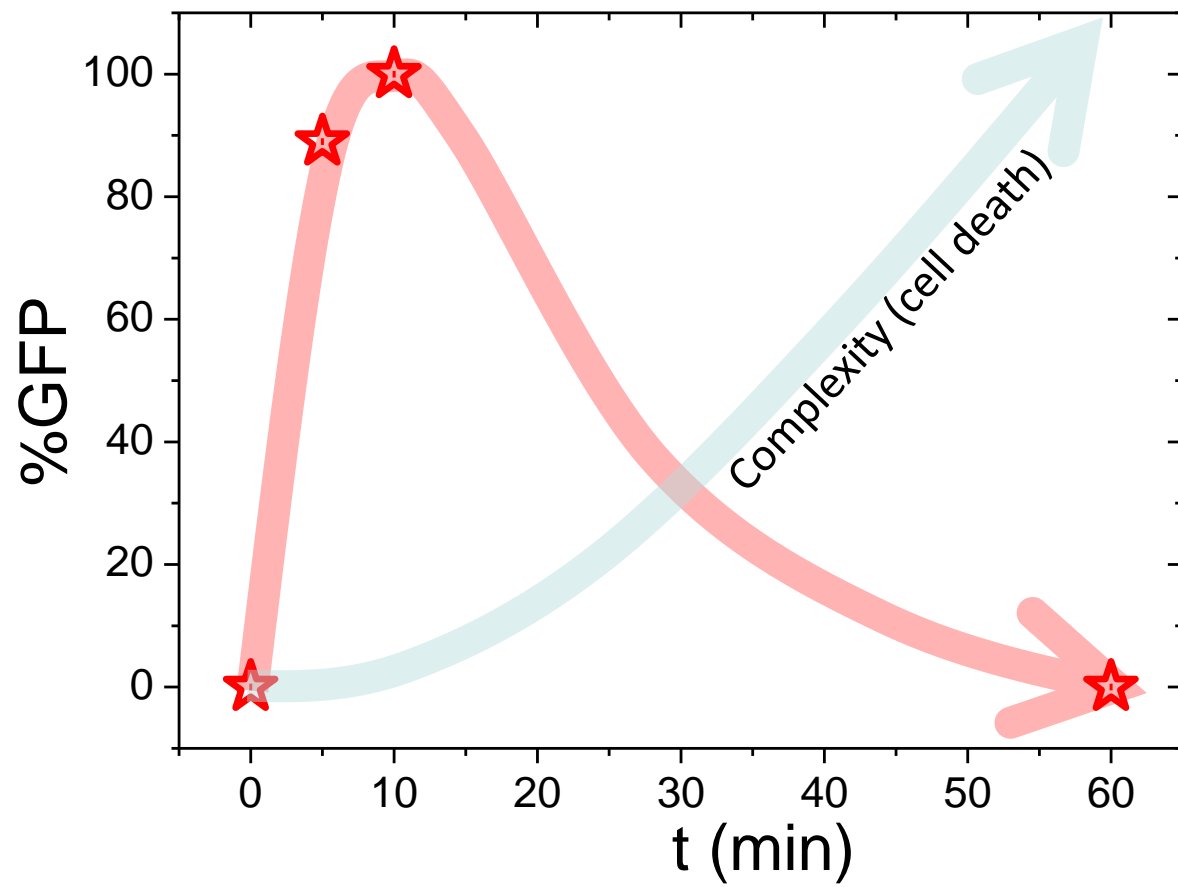
**170kHz H= 34 kA/m**

HSP(58 $\mu$ gNPM/ml)  
y 60 min de RF



**170kHz H= 34 kA/m**

Se observa que la expresión de GFP bajo mucho ya que, como se ve en el grafico de la izquierda, estas células están muy dañadas y al lisarse la membrana se pierde la GFP.



## **Class 2b**

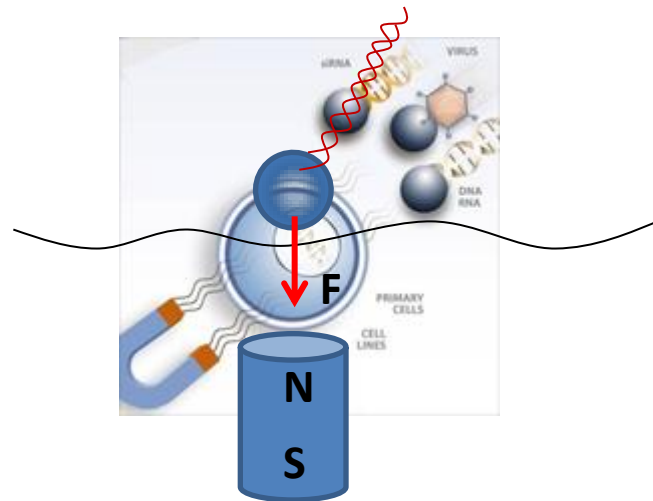
In Vitro Magnetofection: Magnetic Force Influence

Ferrogels PVA/Fe oxide

# In Vitro Magnetofection: Magnetic Force Influence

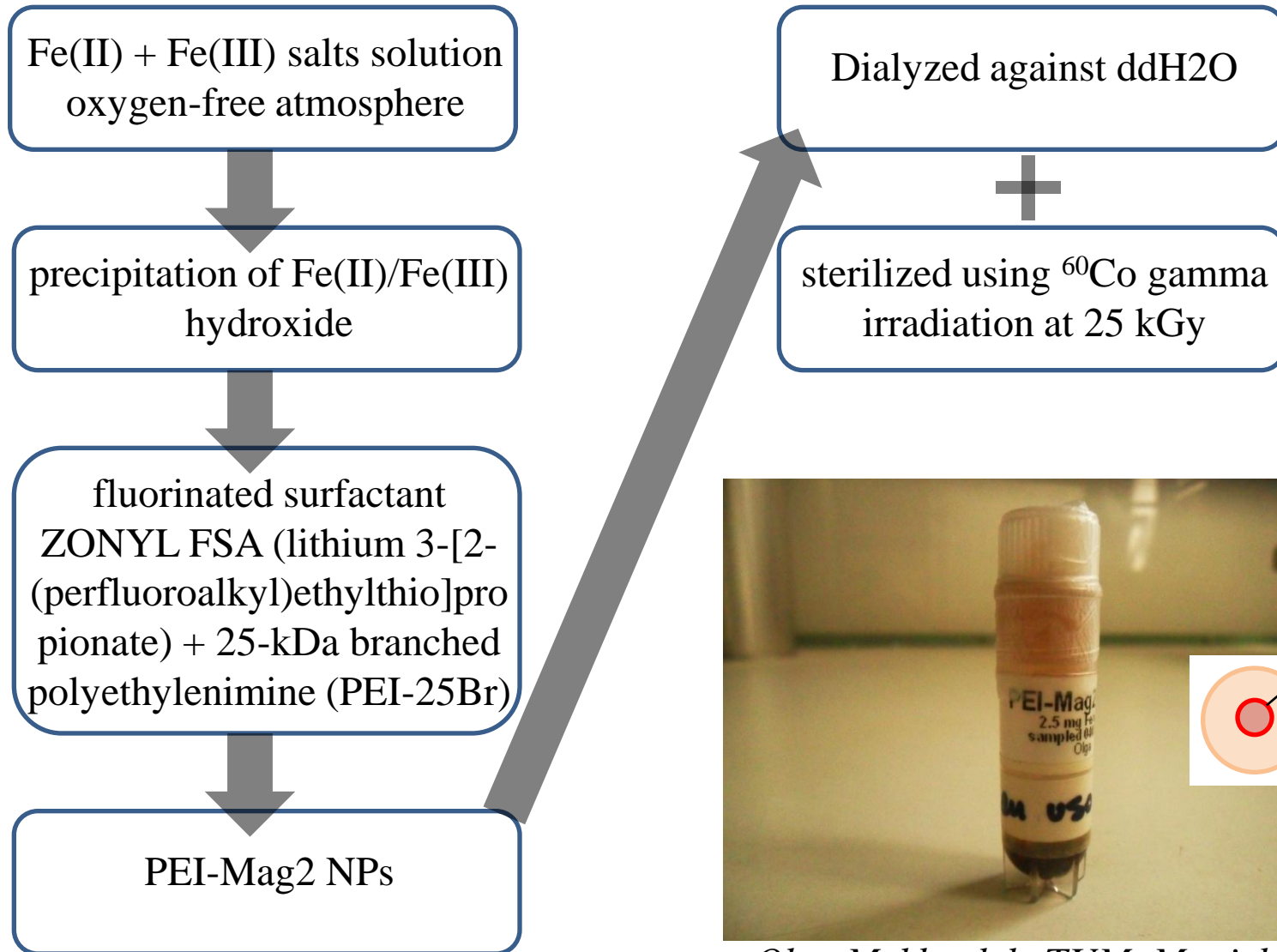
*Arciniegas L, Pardo J, Pasquevich GA, Goya RG,  
Mykhaylik O, Sánchez FH*

*Magnetofection is a simple and highly efficient transfection method that uses magnetic fields to concentrate particles containing nucleic acid into target cells*





# PEI-Mag2 Nanoparticles



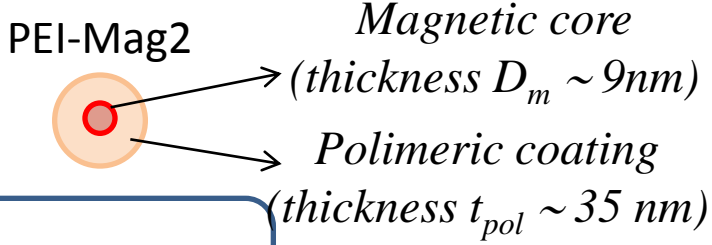
*Olga Mykhaylyk, TUM, Munich*

# PEI-Mag2 RAd-GFP Complex



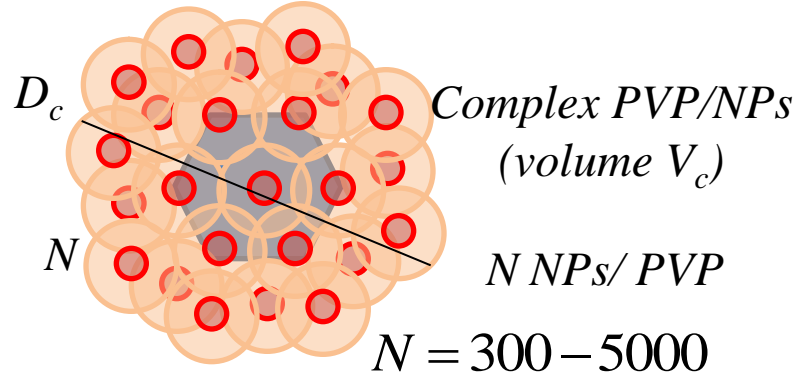
*Virus (PVP, thickness  $D_V \sim 80 \text{ nm}$ )*

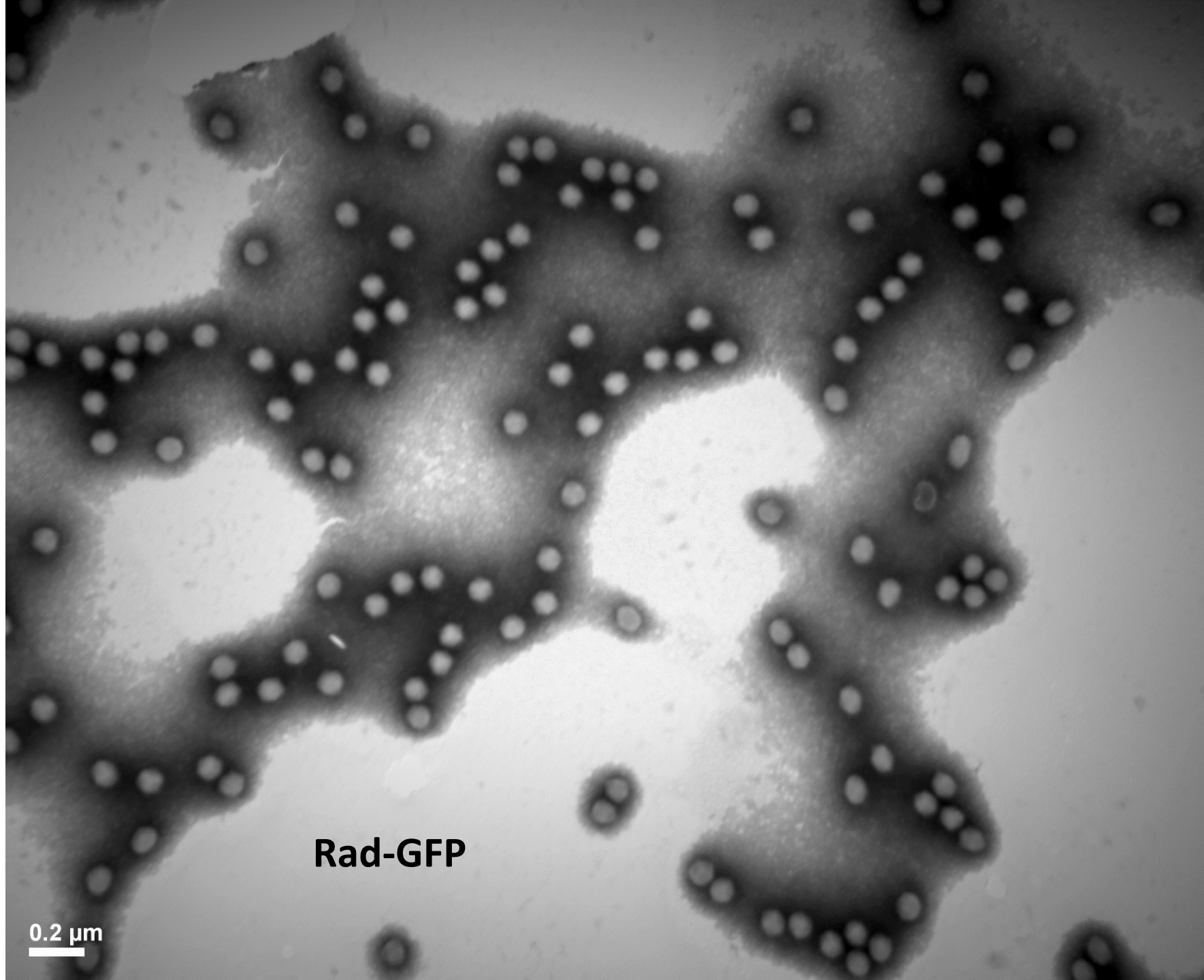
Rad-GFP: Adenovector which expresses GFP gen (*Aequorea victoria*) under citomegalovirus murino (mCMV) promotor control.



PEI-Mag2 NPs

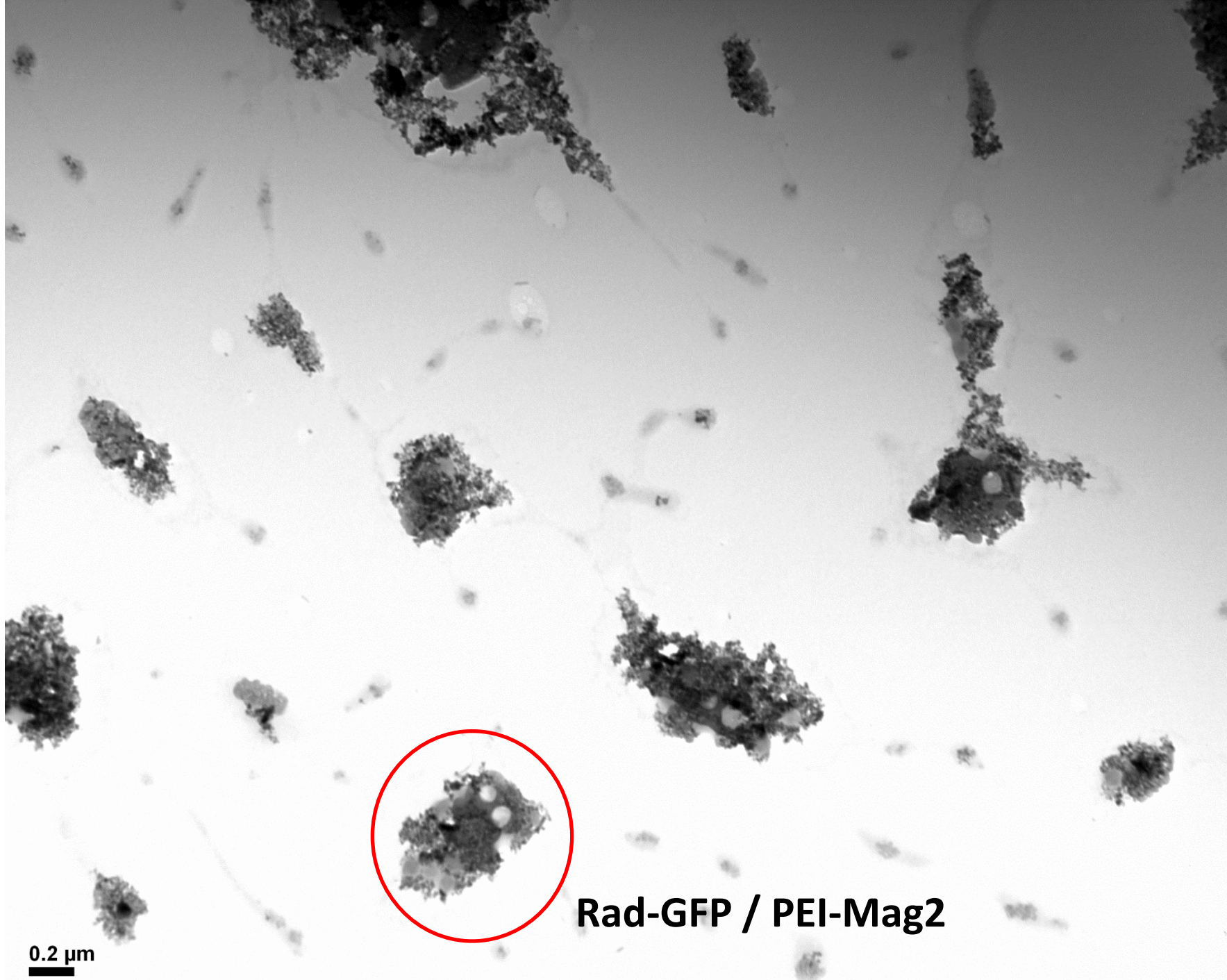
PEI-Mag2 RAd-GFP Complex





**Rad-GFP**

0.2  $\mu\text{m}$



0.2  $\mu\text{m}$

**Rad-GFP / PEI-Mag2**

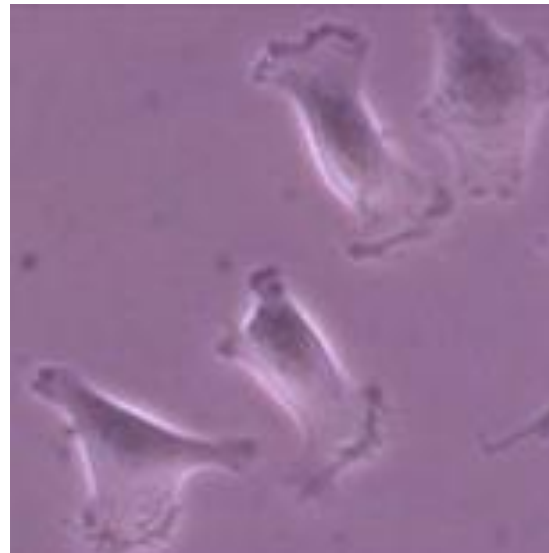


**Rad-GFP / PEI-Mag2**

**0.2  $\mu$ m**

# B92 cell culture conditions

B92 cells (mouse glial cell)  
MEM (Minimum Essential Media),  
10% FBS (Foetal Bovine Serum)  
100 U de penicillin/ml  
0,1 mg de streptomycin/ml  
0,25  $\mu$ g de anphotericin B/ml  
5% CO<sub>2</sub> atmosphere  
37 °C



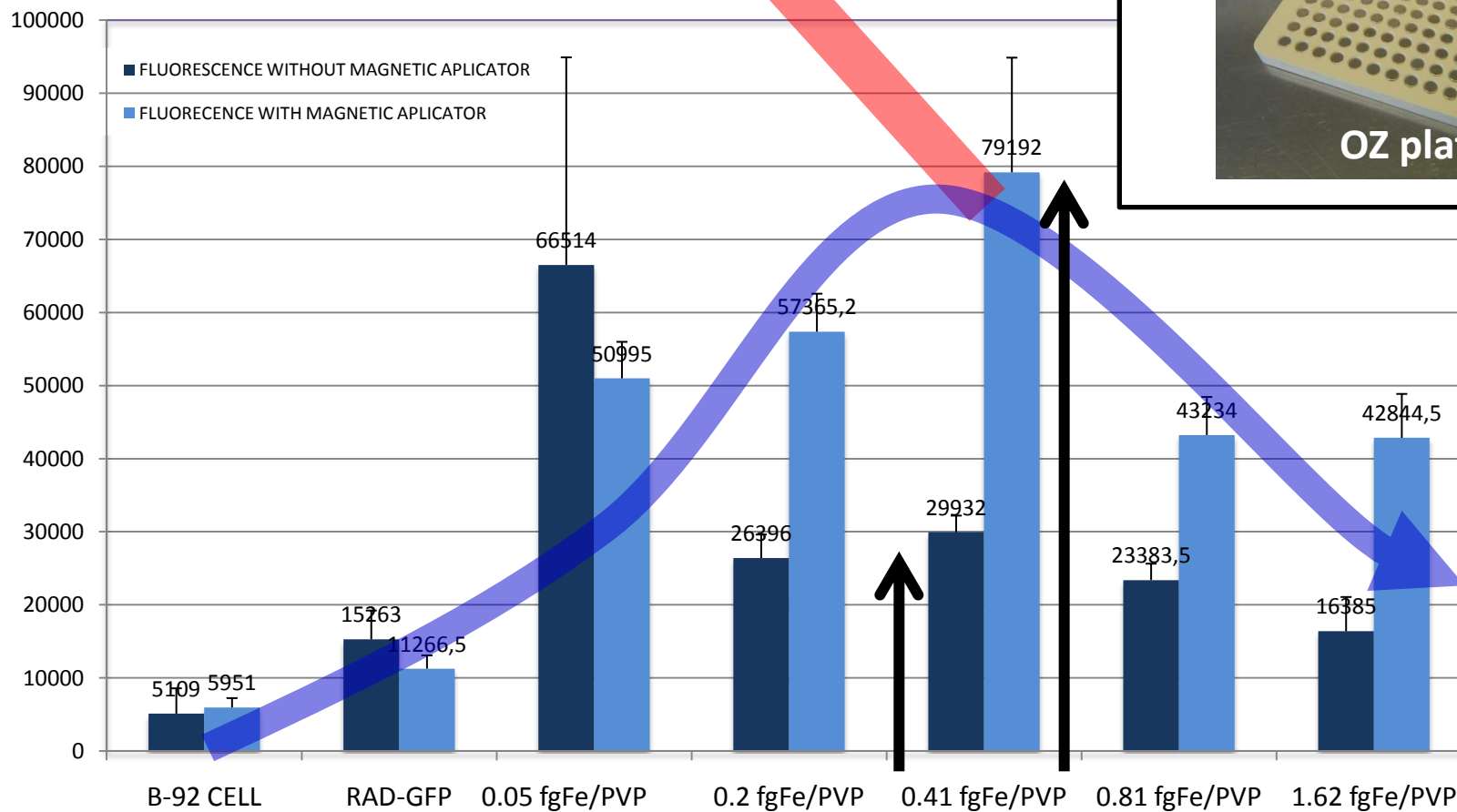
# PEI-Mag2 RAd-GFP Complex / optimum ratio

Acta Bioquím Clín Latinoam 2013; 47 (2): 399-406

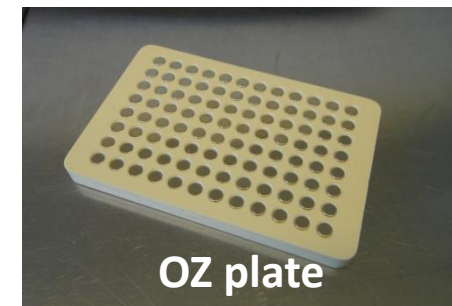
Joaquín Pardo<sup>1a\*</sup>, Yolanda Elena Sosa<sup>2a\*</sup>, Paula Cecilia Reggiani<sup>3a</sup>,  
Magda Lorena Arciniegas<sup>4b</sup>, Francisco Homero Sánchez<sup>5b</sup>, Rodolfo Gustavo Goya<sup>6a</sup>

Selected Ratio 0.5

## RELATIONSHIPS DETERMINATION



Wells plate



OZ plate

# One word on magnetic applicators

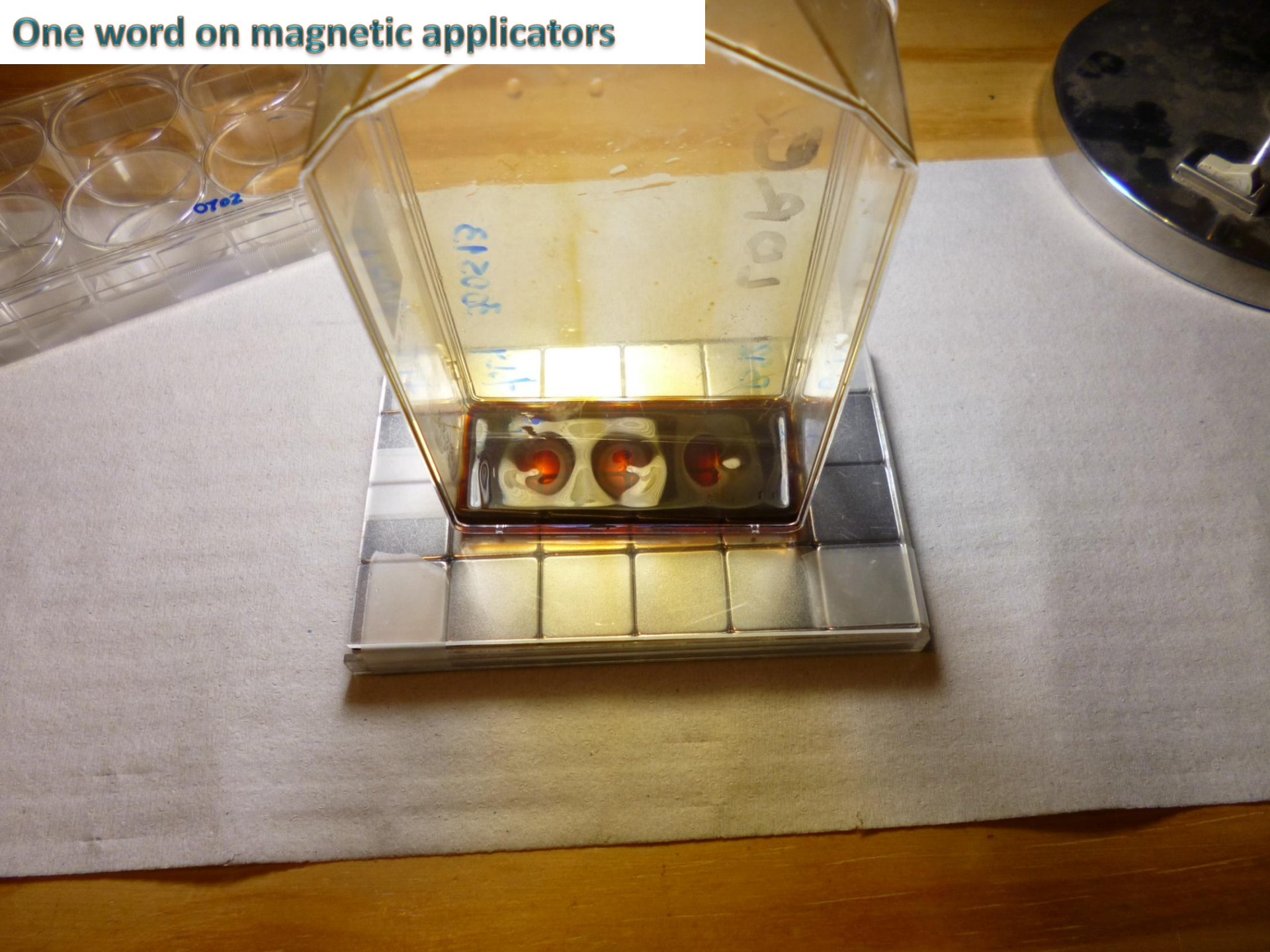




# One word on magnetic applicators



# One word on magnetic applicators



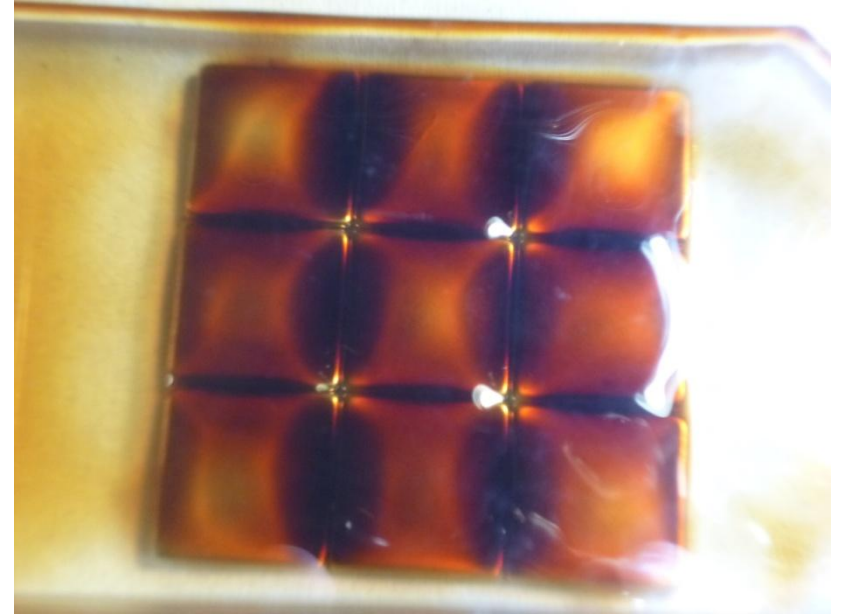
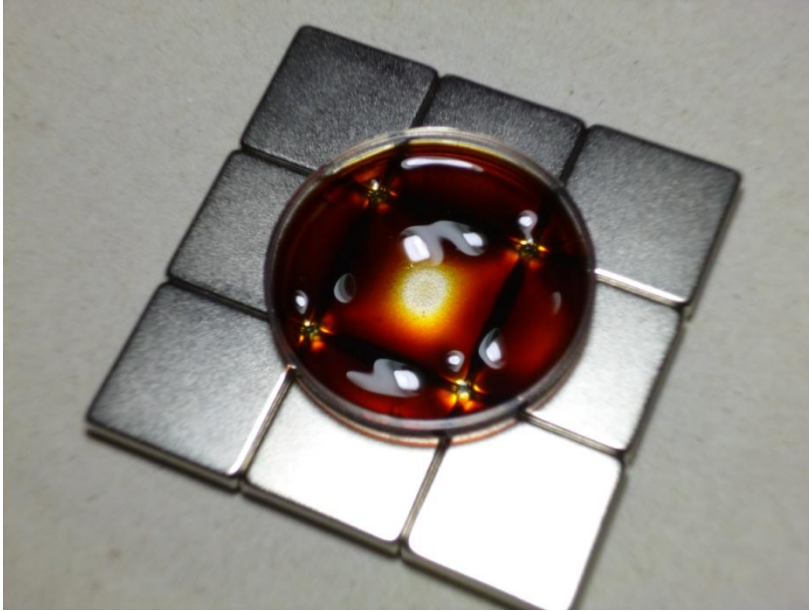
# One word on magnetic applicators



# One word on magnetic applicators



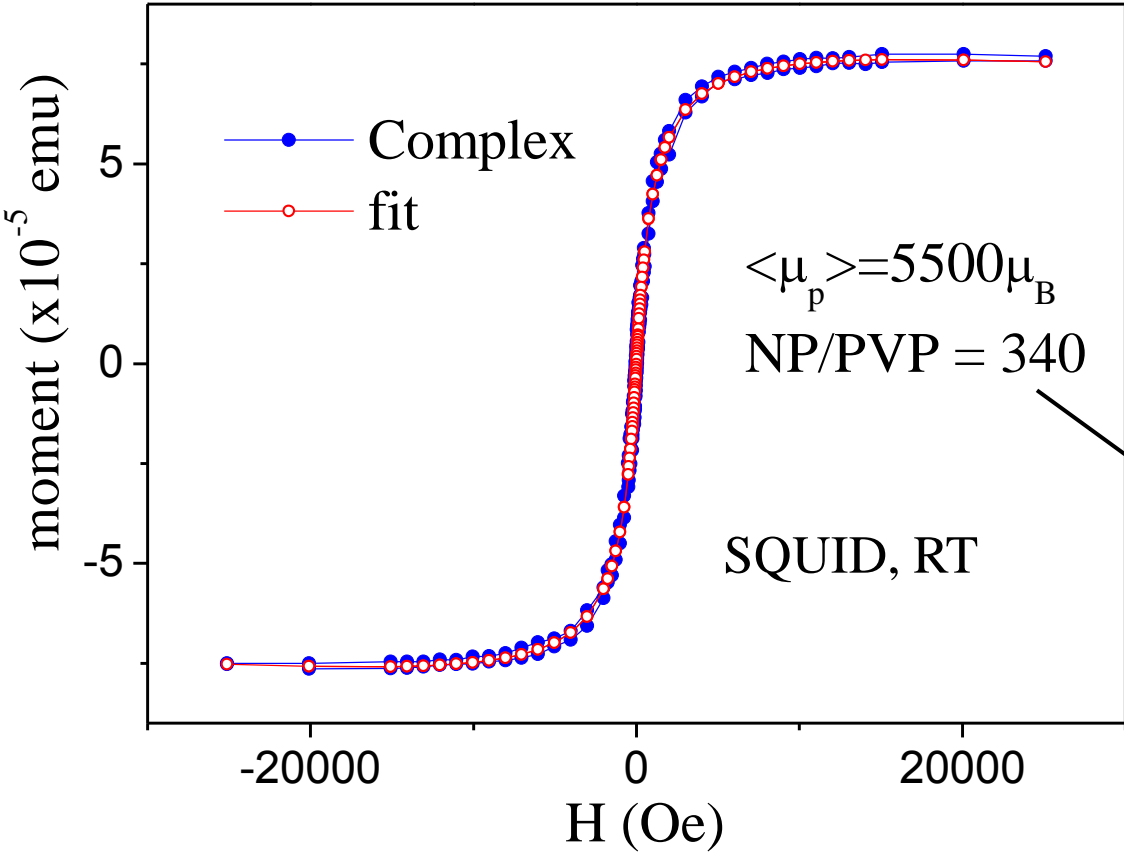
# One word on magnetic applicators



# One word on magnetic applicators

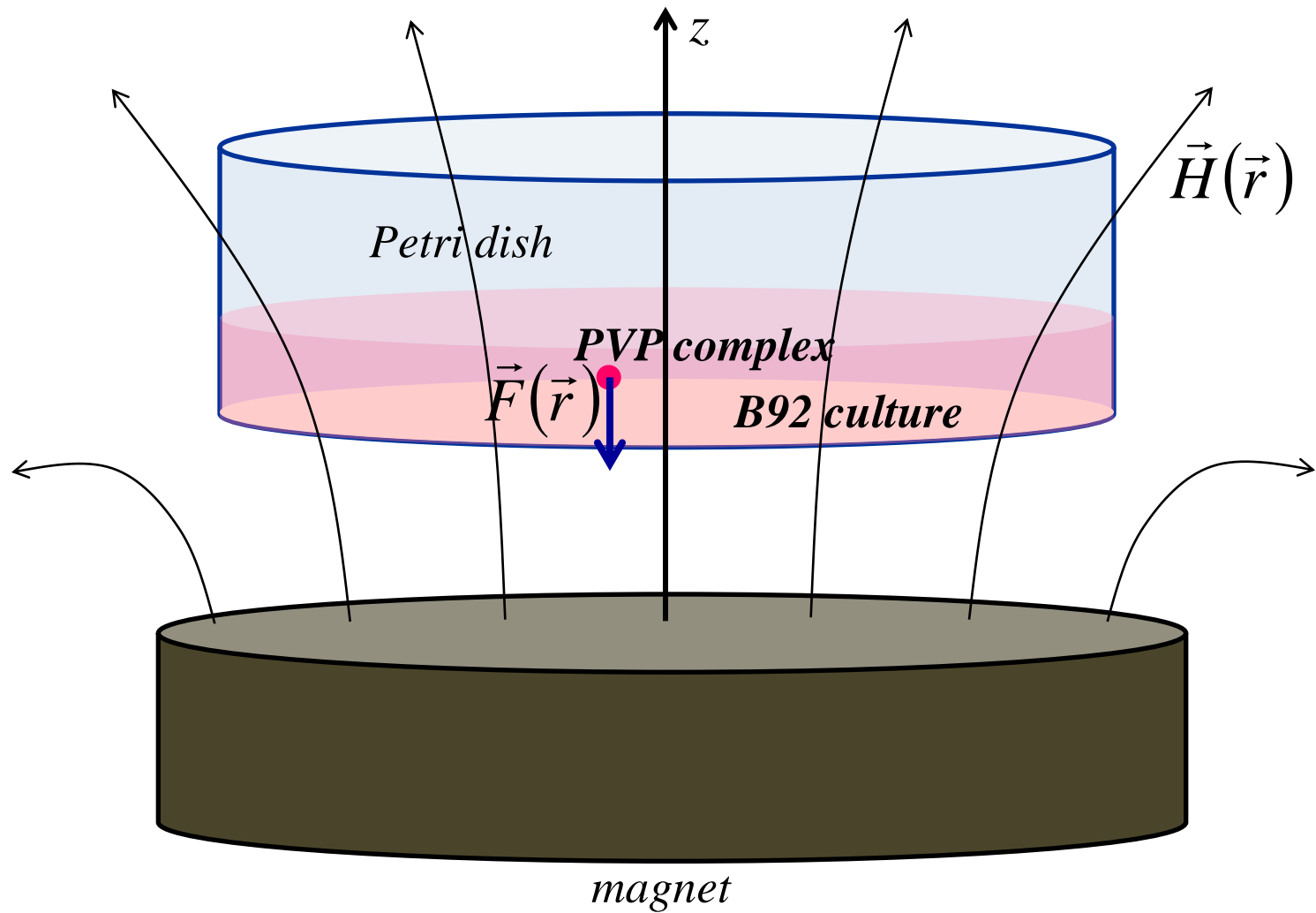


# PEI-Mag2 RAd-GFP Complex / magnetic response



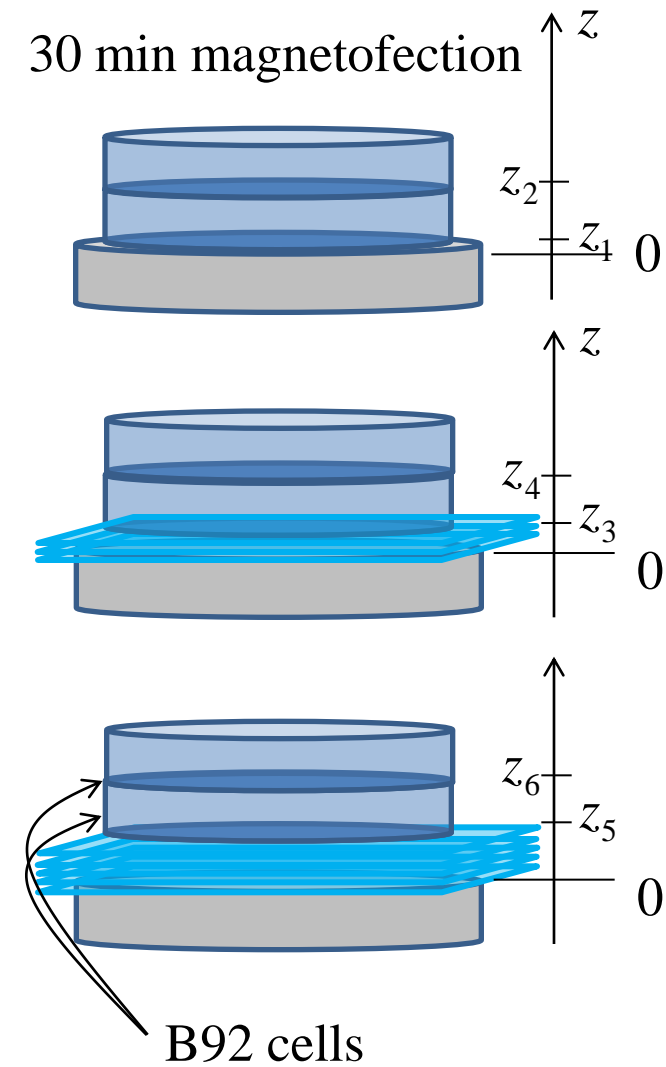
Consistent with selected 0.5 Ratio

# Magnetofection experimental design





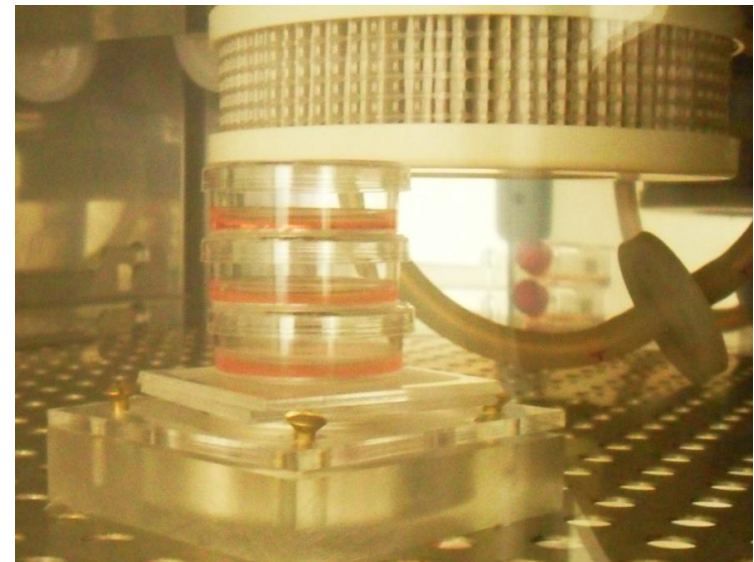
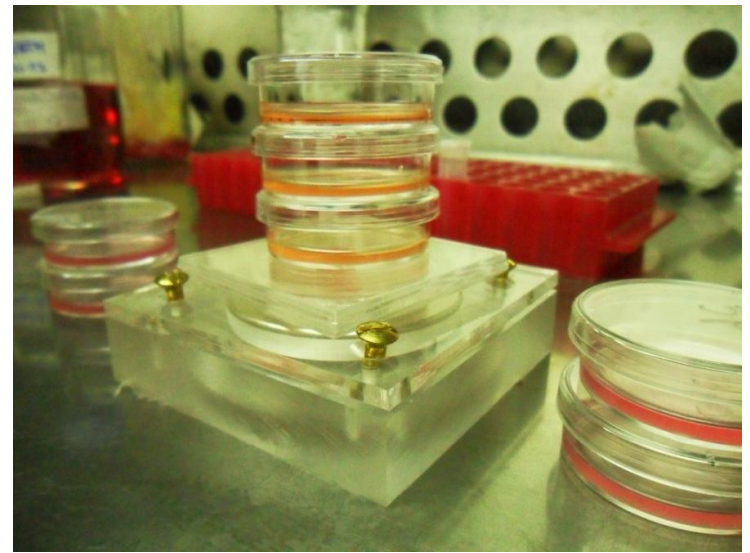
# Magnetofection experimental design



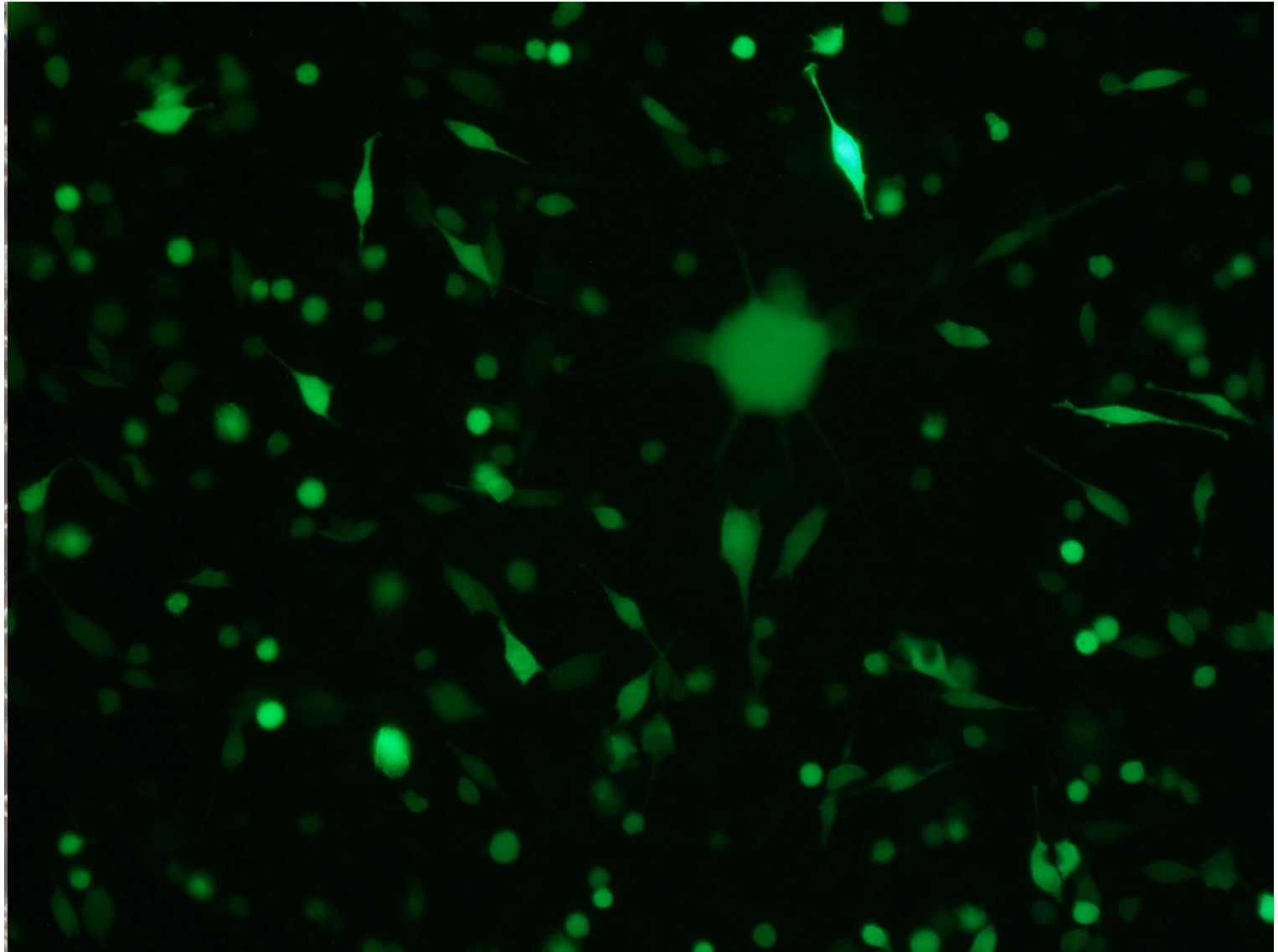
washing

72 h with  
MEM +  
FBS

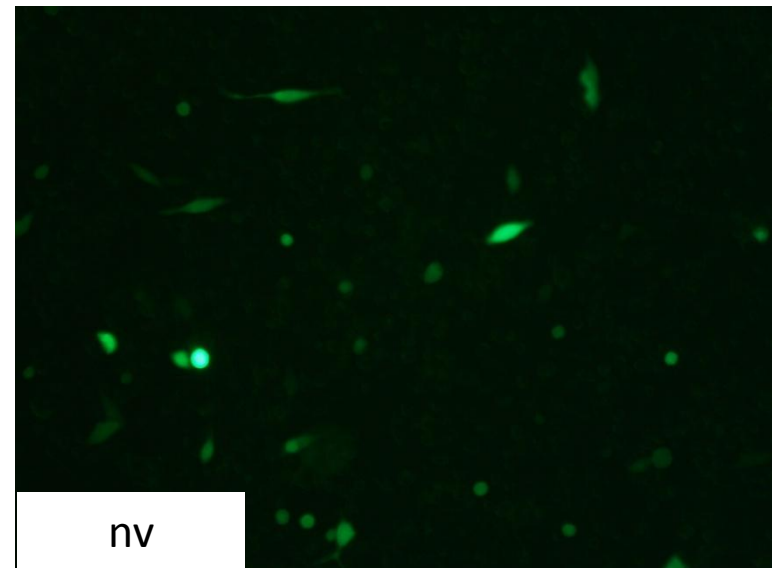
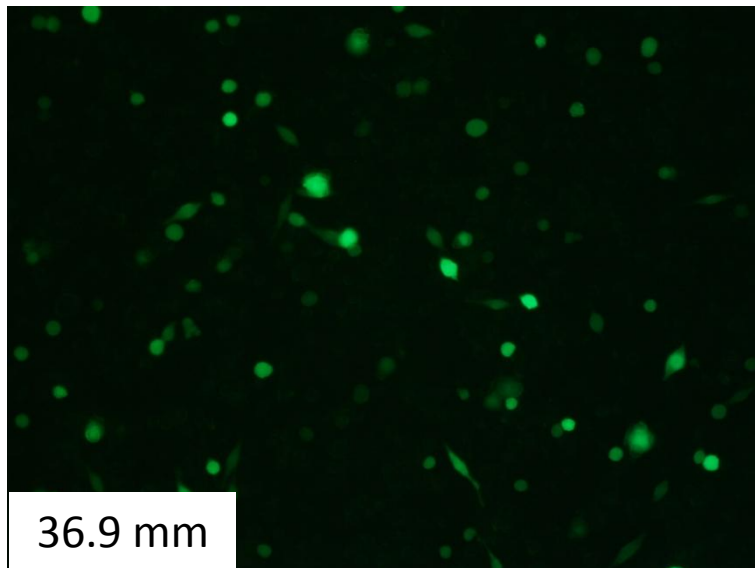
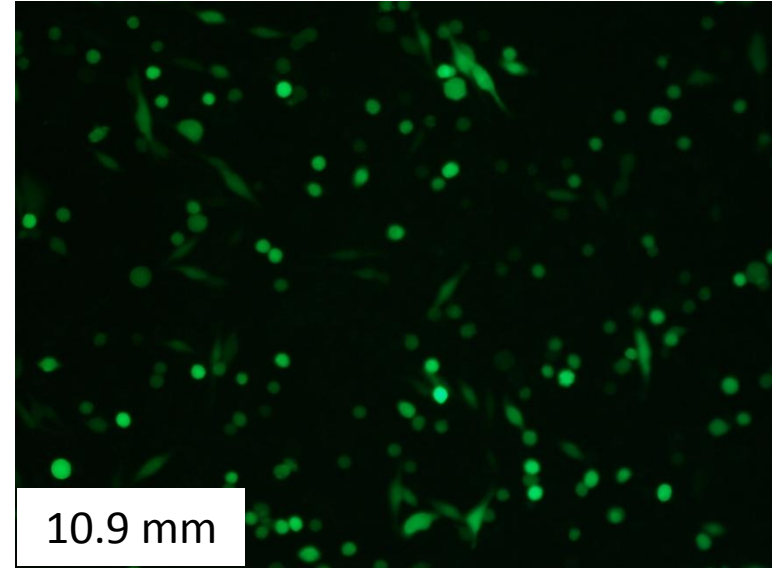
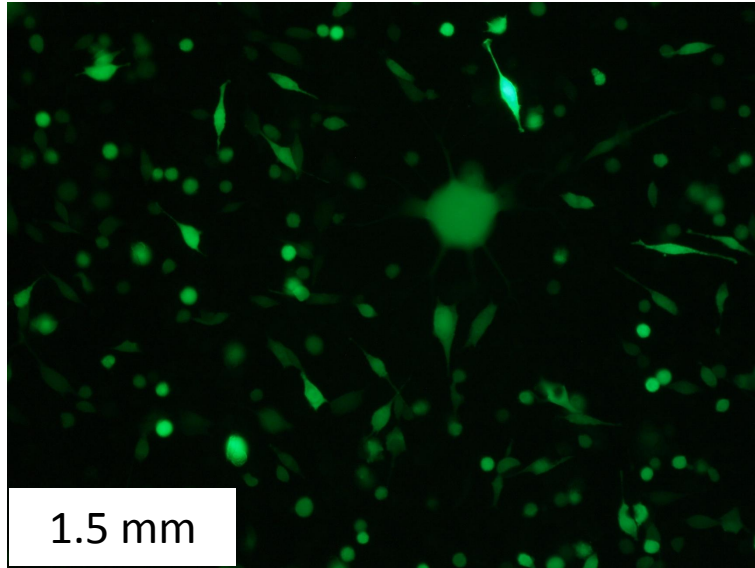
Magnetofection  
fluorescence vs  $z$



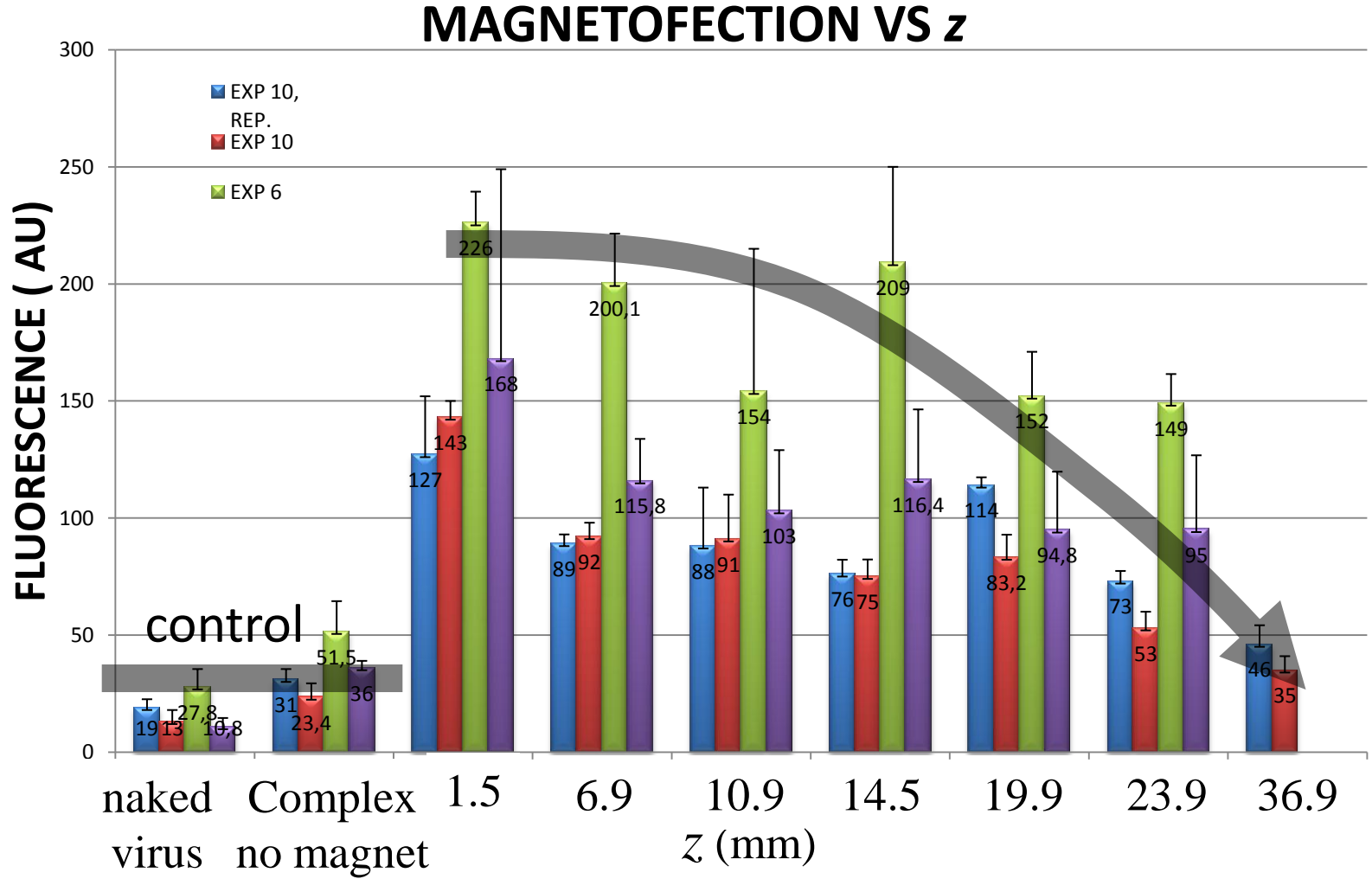
# Magnetofection results – phase and fluorescence obs



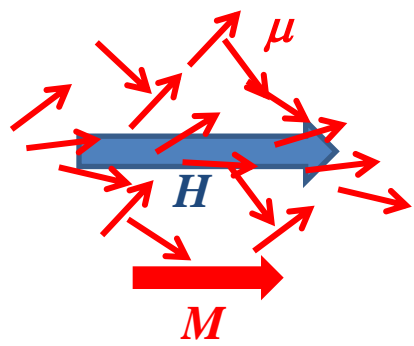
# Magnetofection results – phase and fluorescence obs



# Magnetofection results – Elisa spectrometer



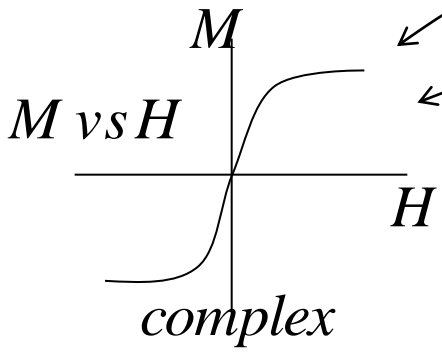
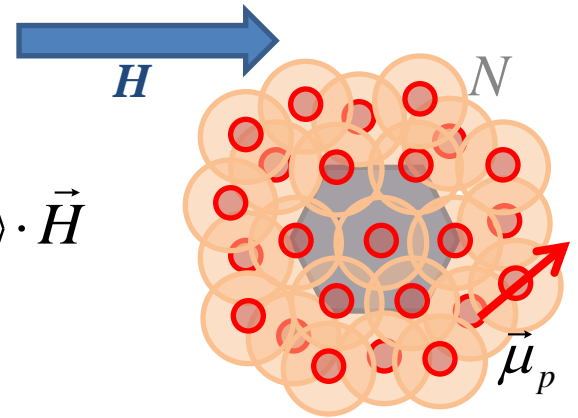
# Magnetic Force analysis



$$U_{\vec{B}} = -\sum \vec{\mu}_{pi} \cdot \vec{B} = -N\mu_0 \langle \vec{\mu}_p \rangle \cdot \vec{H}$$

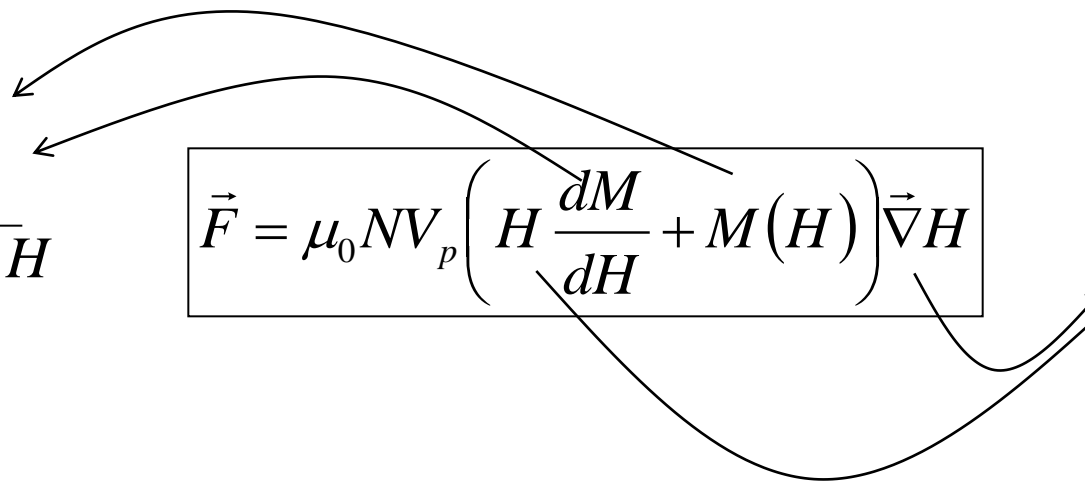
$$\langle \vec{\mu}_p \rangle = V_p \vec{M}(\vec{H}) // \vec{H}$$

$$\vec{F} = -\vec{\nabla} U_{\vec{B}} = N\mu_0 V_p \vec{\nabla}(MH)$$



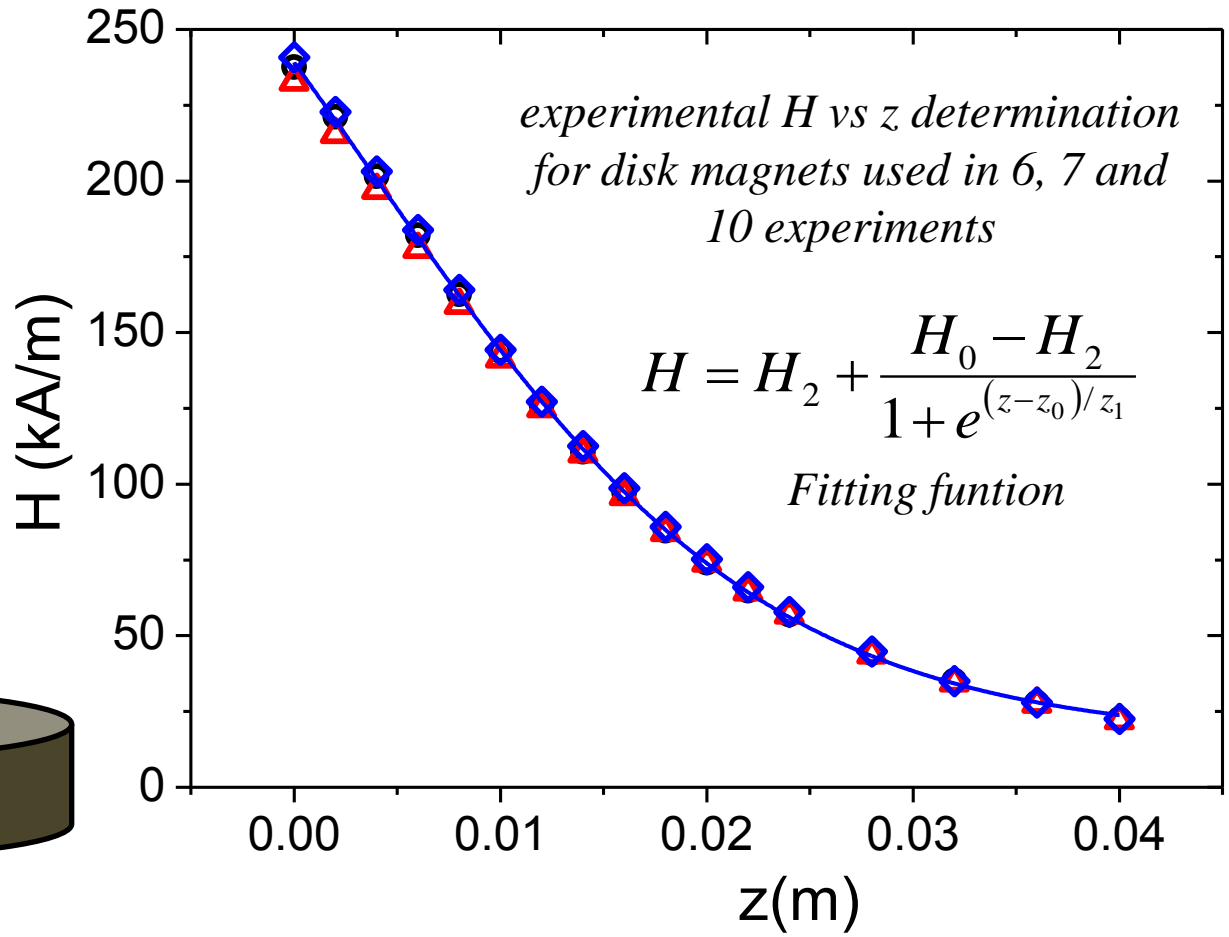
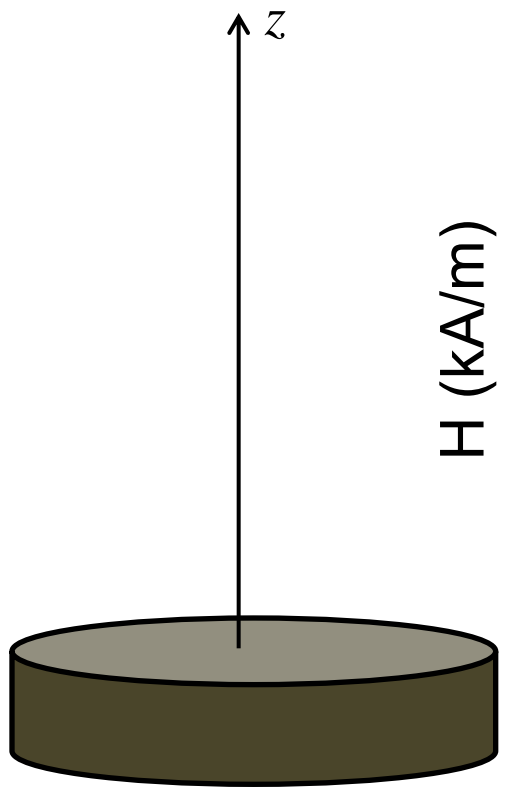
$$\vec{F} = \mu_0 N V_p \left( H \frac{dM}{dH} + M(H) \right) \vec{\nabla} H$$

*magnets*  
*H vs position*  
*exp. data*



# Magnetic Force analysis, experimental

H(z) measurement



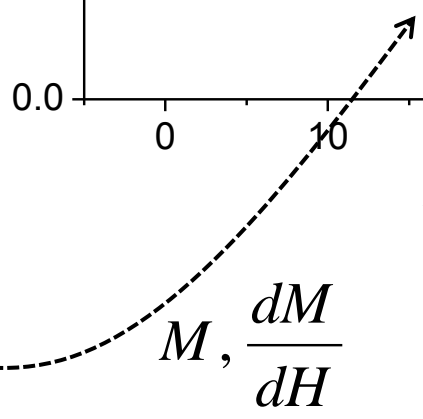
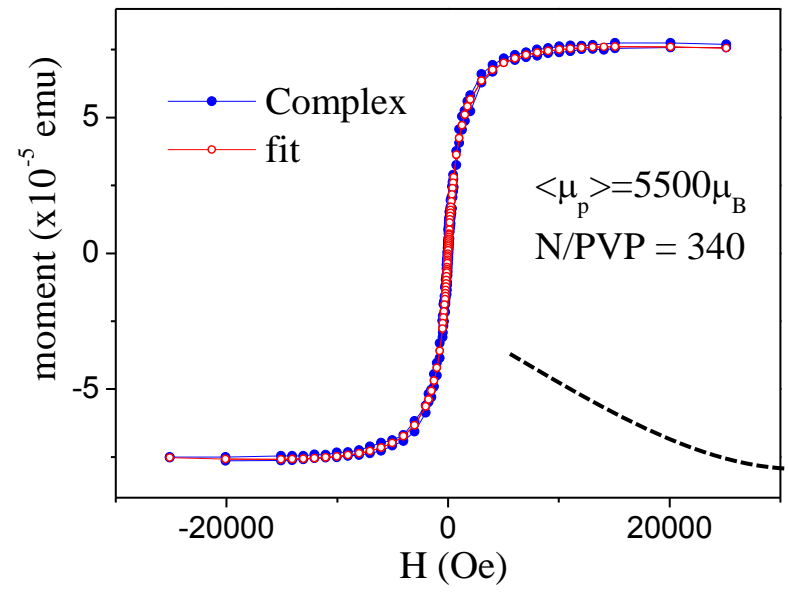
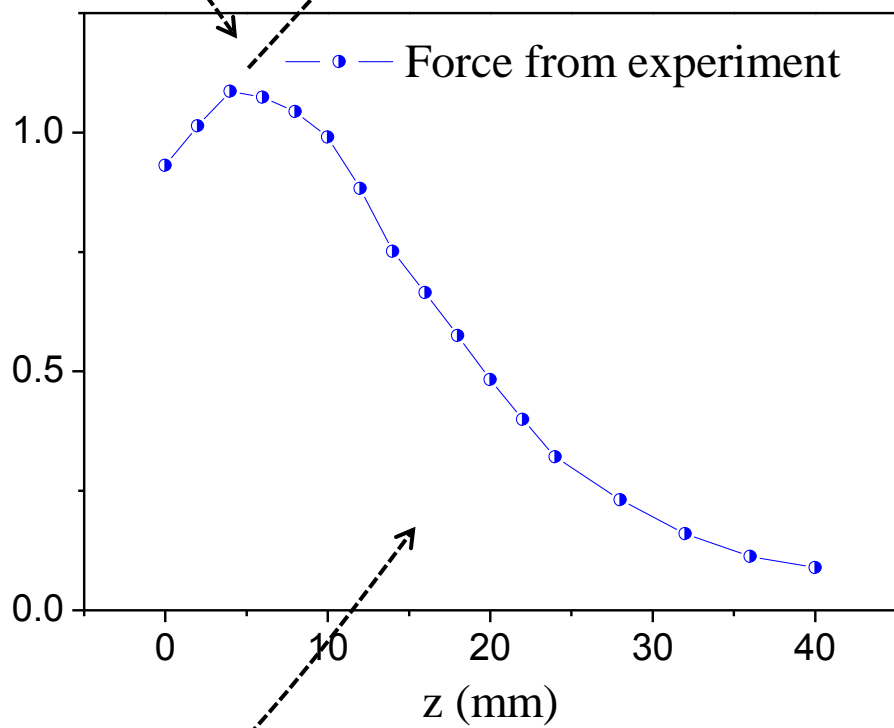
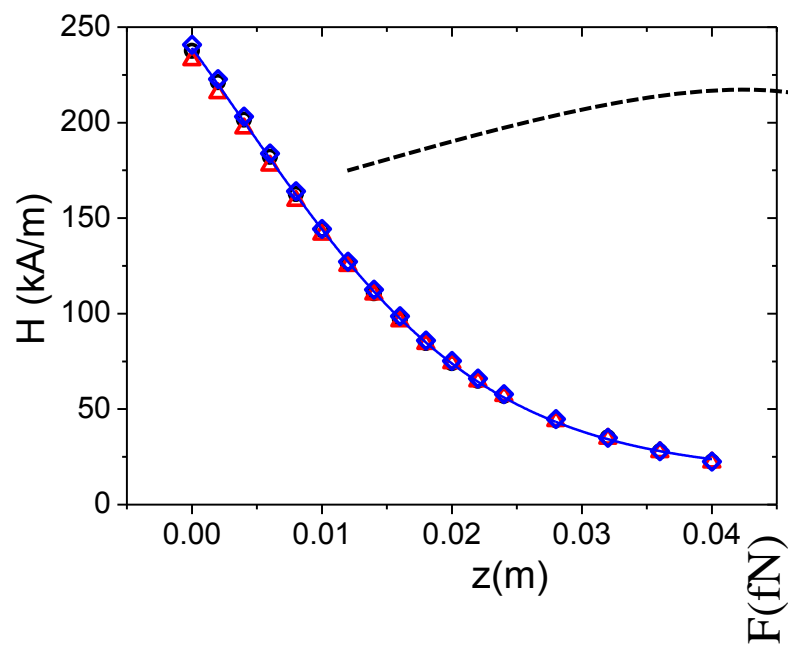
From M vs H



$$\mu_p \approx 5500 \mu_B$$

$$N \approx 350 \text{ NP / PVP}$$

# Magnetic Force analysis, experimental



# Magnetic Force analysis, what does it do?

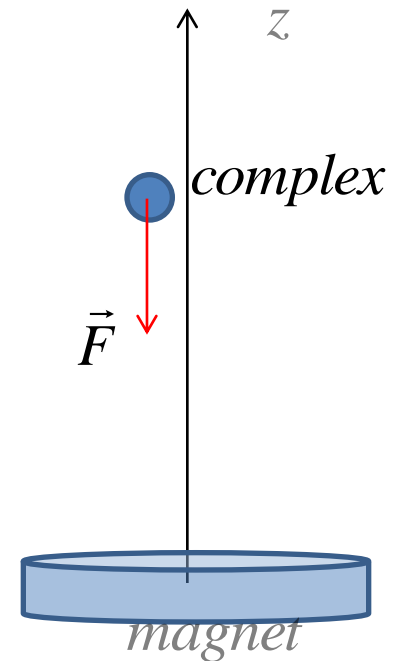
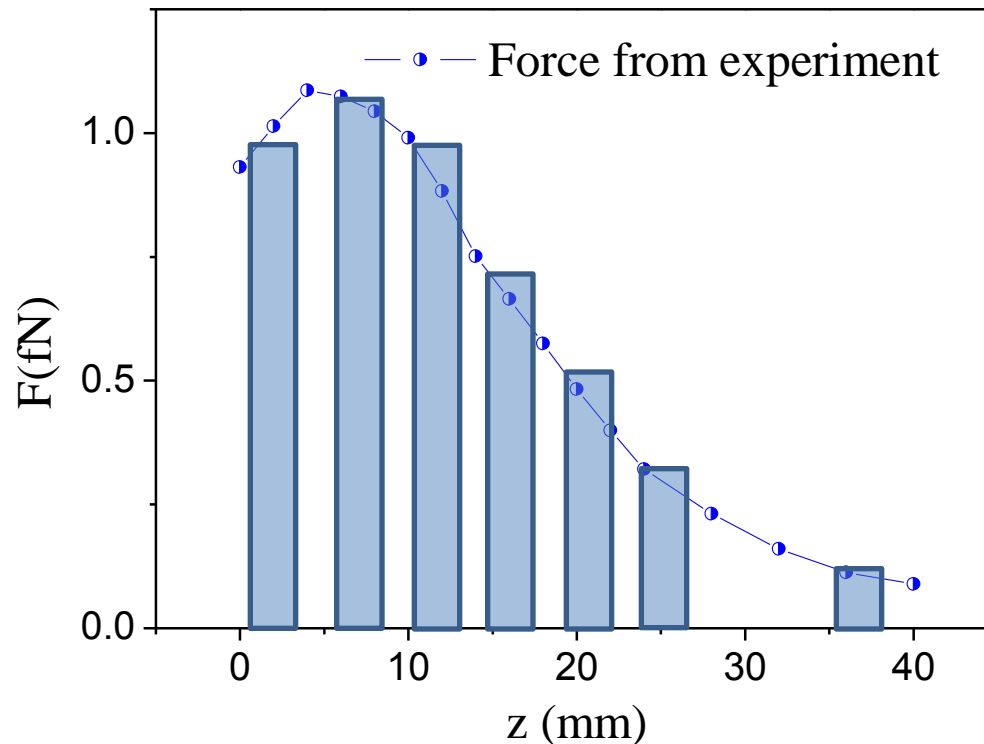
It changes state of motion of complexes

$$v(z, t)$$

It changes distribution of complexes in the fluid

$$c(z, t)$$

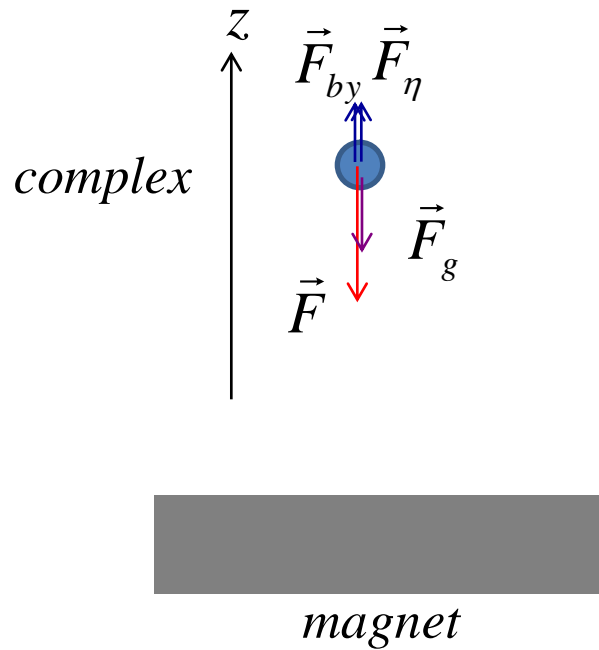
Constant Force approximation: force variation within each PD can be neglected





# Magnetic Force analysis, dynamics

Complex dynamics



$$\vec{F}_\eta = 6\pi\eta D_c / 2$$

$$F^* = F_{by} - (F + F_g)$$

$$F_{by} = \rho_s V_c; \quad F_g = \rho_p V_c$$

*solvent*                      *particle*

# Magnetic Force analysis, dynamics

Complex volume and mass

$$V_c \approx V_V + N(V_m + V_{pol}) \approx \frac{\pi}{6} (D_V^3 + ND_{pol}^3)$$

$$m_c \approx m_V + N(m_m + m_{pol}) \approx \frac{\pi}{6} (\rho_V D_V^3 + N(\rho_{pol}(D_{pol}^3 - D_m^3) + \rho_m D_m^3))$$

USING

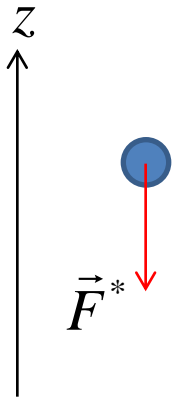
$$\left\{ \begin{array}{l} D_m \approx 9 \text{ nm} \\ D_{pol} \approx 70 \text{ nm} \\ D_V \approx 80 \text{ nm} \\ N \approx 350 \end{array} \right.$$

$$V_c \approx 1.3 \times 10^{-19} \text{ m}^3 = 1.57 \times 10^8 \text{ nm}^3 \Rightarrow D_c \approx 600 \text{ nm}$$

$$m_c \approx 1.4 \times 10^{-16} \text{ kg} = 0.14 \text{ fg}$$

$$P_c \approx 1.370 \times 10^{-15} \text{ N}; \quad F_{by} \approx 1.27 \times 10^{-15} \text{ N}$$

# Magnetic Force analysis, space distribution of complexes

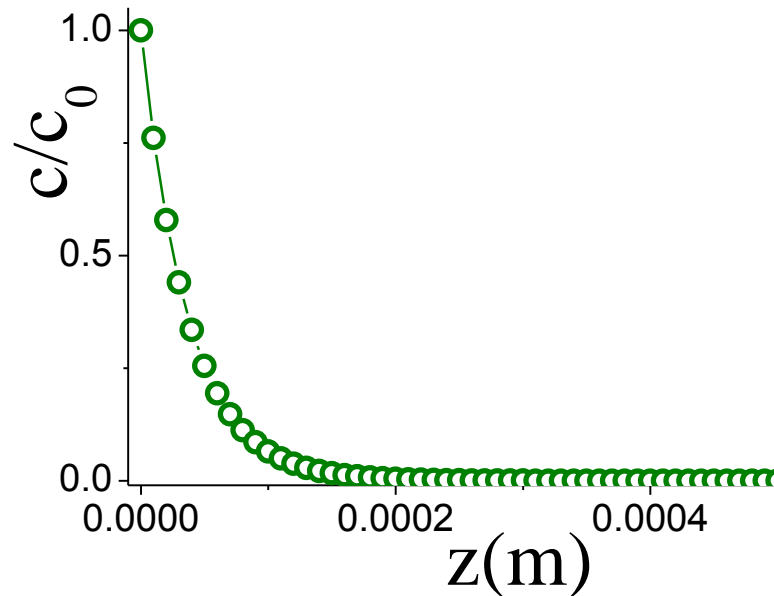


*Energy  $U$*

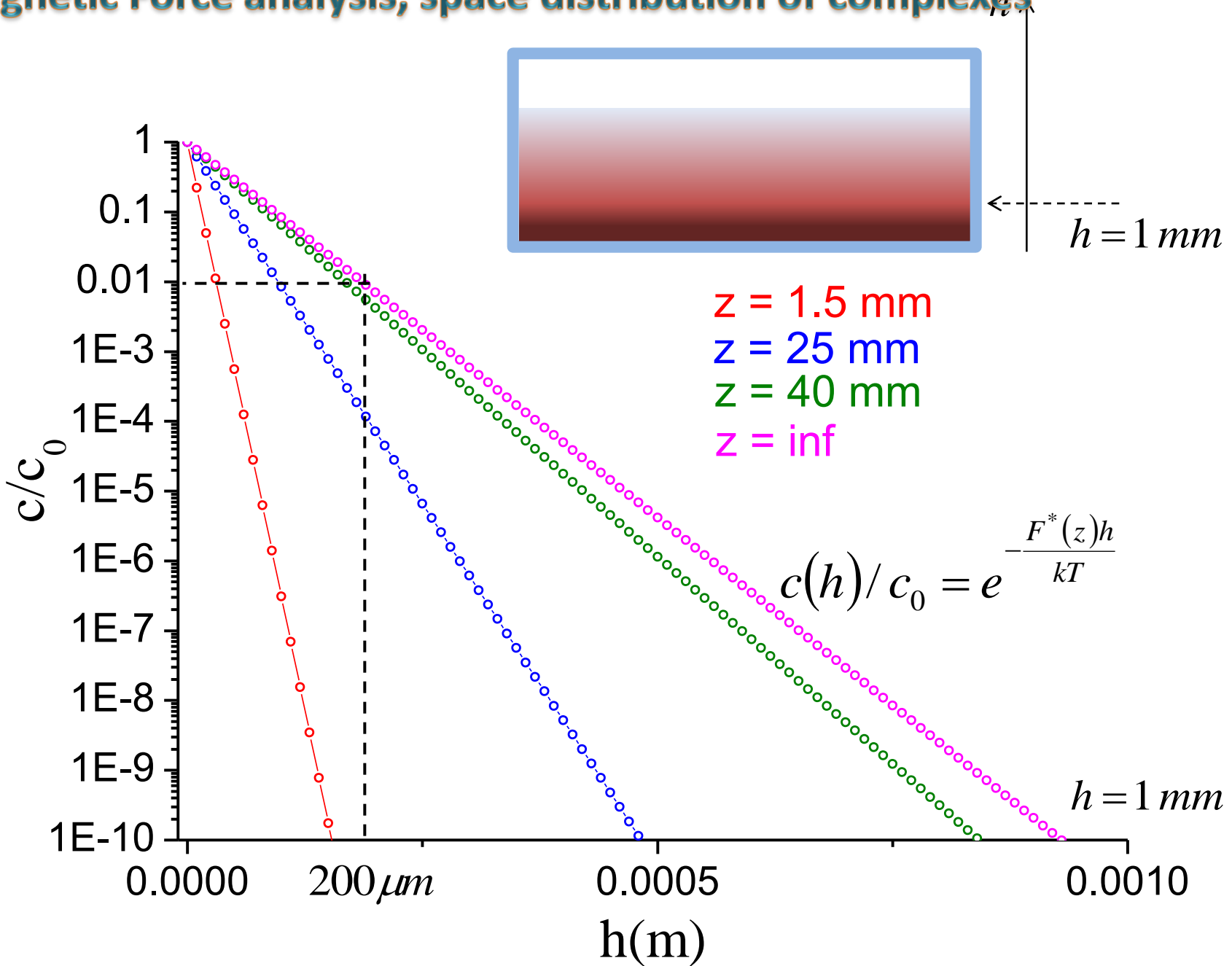
$$U = F^* z$$

*Equilibrium concentration  $c(z)$*

$$c(z)/c_0 = e^{-\frac{F^* z}{kT}}$$

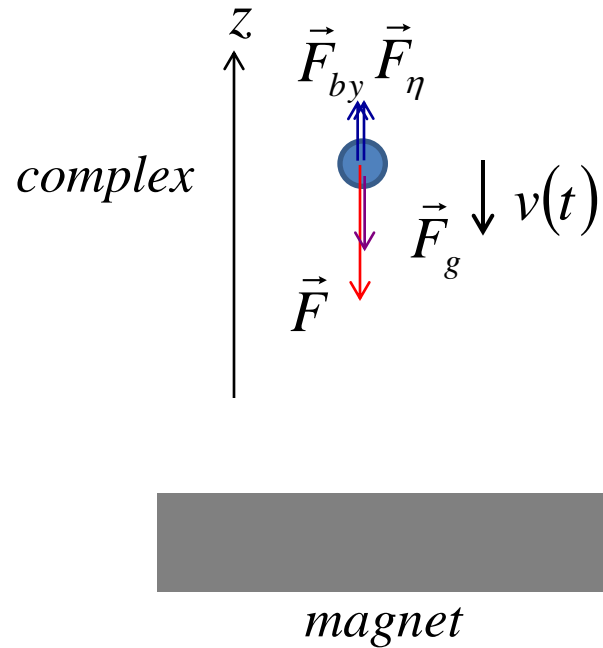


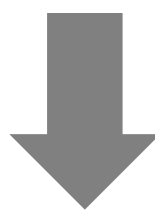
# Magnetic Force analysis, space distribution of complexes



# Magnetic Force analysis, motion of complexes

complex motion dynamics

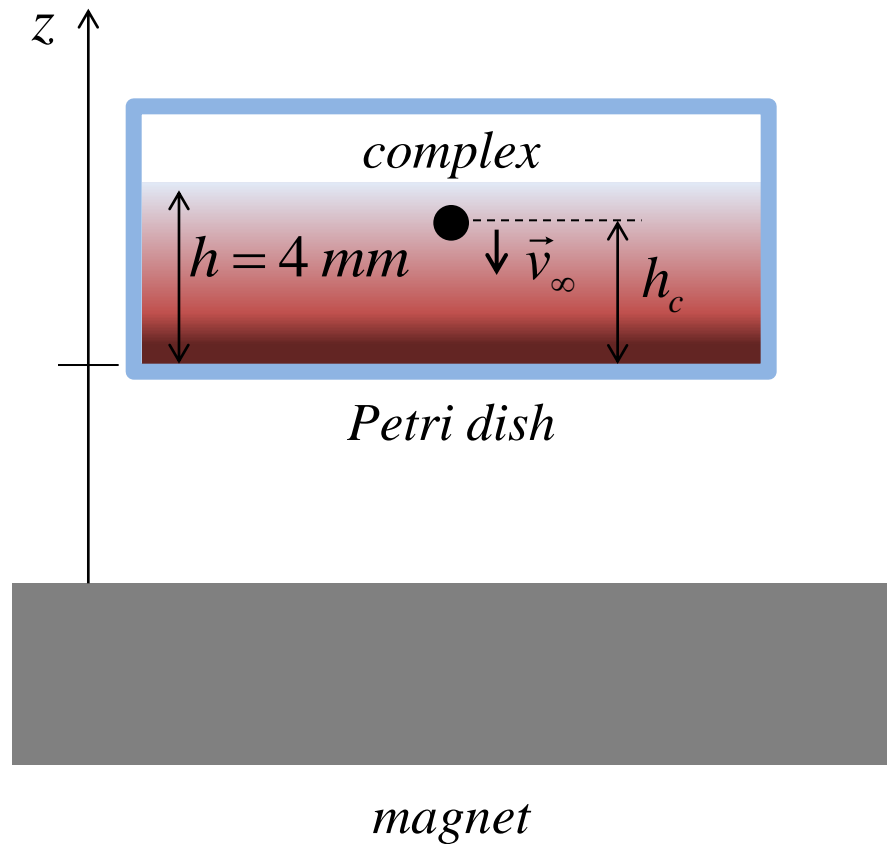



$$v(t) = \frac{F^*}{3\pi\eta D_c} \left( 1 - e^{-\frac{3\pi\eta D_c t}{m_p}} \right)$$

*complex speed*

# Magnetic Force analysis, theoretical

$f_{30}$ : fraction of complexes reaching bottom of PD in 30 min (mf time  $t_{mf}$ )



$$v_\infty = \frac{F^*}{6\pi\eta R}$$

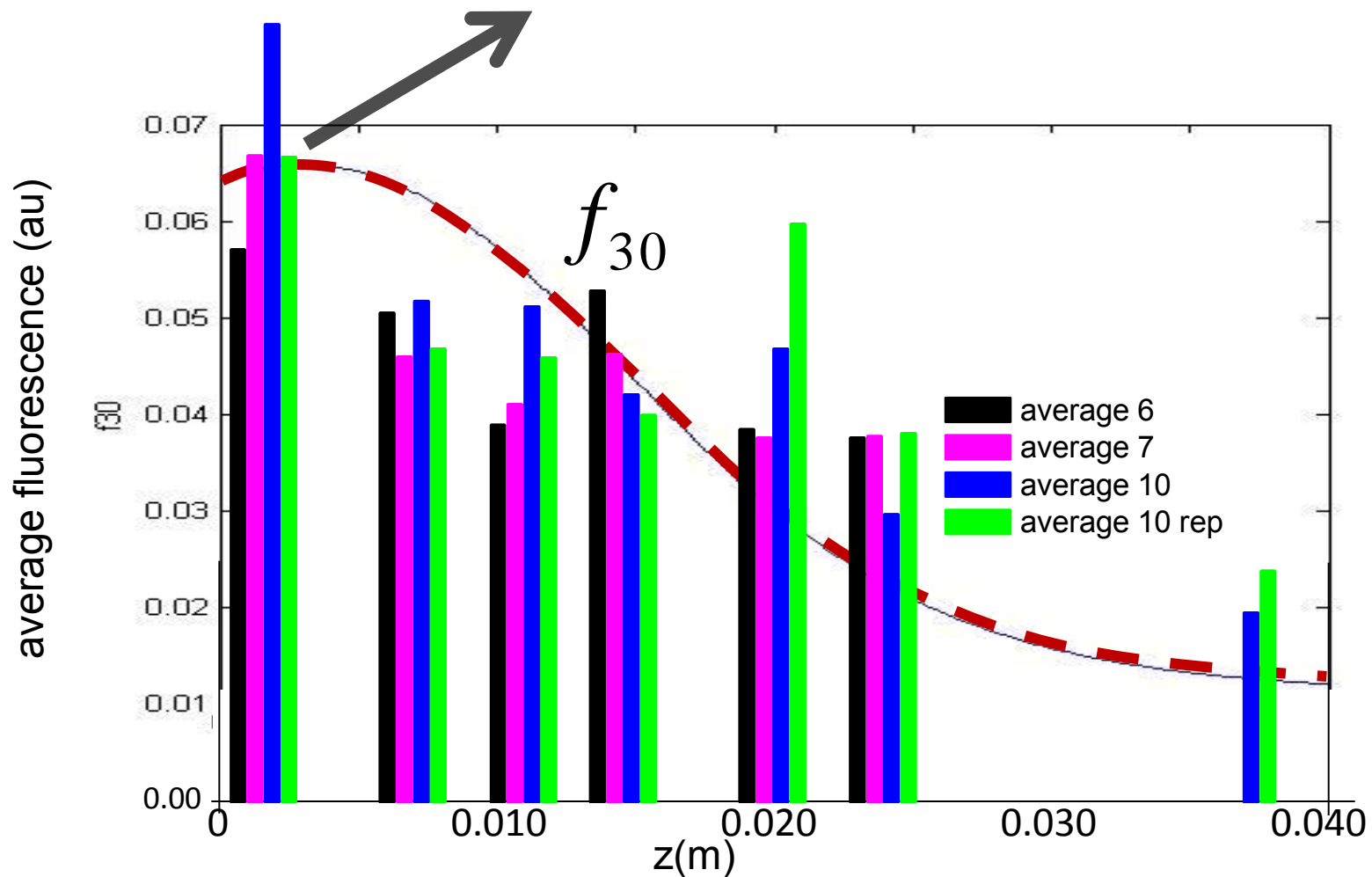
$$t_{mf} = 30 \text{ min}$$

$$h_c = v_\infty t_{mf}$$

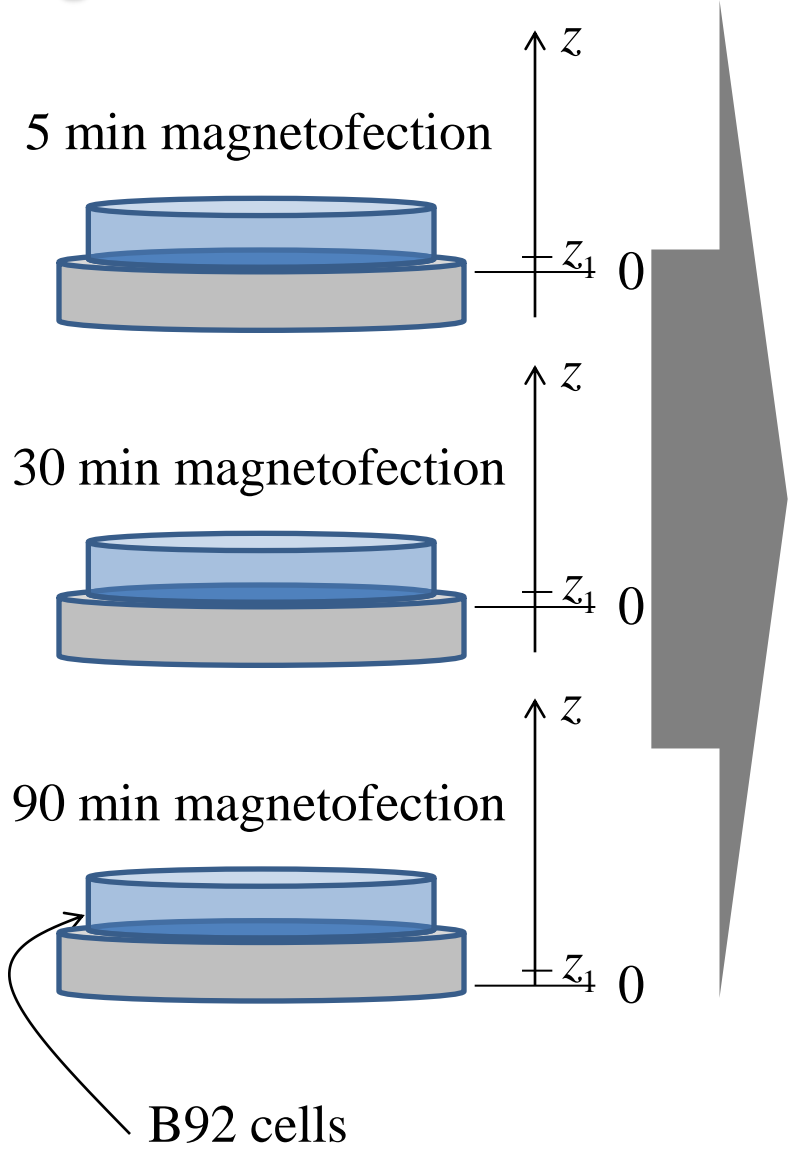
$$f_{30} = h_c / h$$

# Comparison with experiment

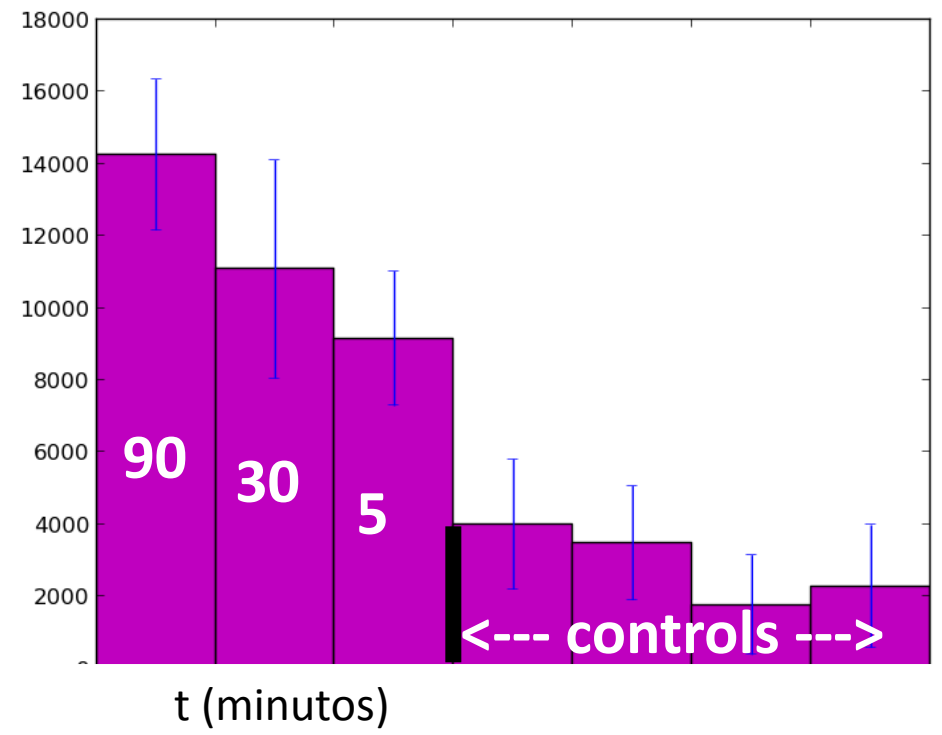
*in 30 min ~ 7 % of complexes have reached bottom of PD*



# Magnetofection vs. time



washing → 72 h with MEM + FBS → fluorescence vs  $t$





## Conclusions III

Magnetic force analysis gives a fair semiquantitative account of experiments of magnetofection vs  $z$

Field applicators can be designed to optimize magnetic force for in vivo magnetofection at moderate depths

Longer times or **Stronger** magnetic force is needed in order to improve magnetofection effects.

# Ferrogels PVA/Fe oxide

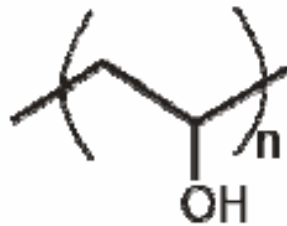
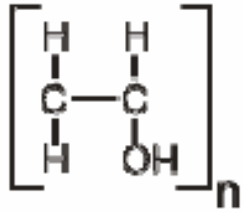
*Arciniegas L, Pasquevich GA, Mendoza Zélis P,  
González J., Álvarez V, Hoppe C, Sánchez FH*

*Ferrogels: magnetic hydrogels with high hydration ability and strong magnetic response.*

*Hydrogels are constituted by crosslinked polymeric chains. Physically crosslinked PVA hydrogels are soft biocompatible materials.*

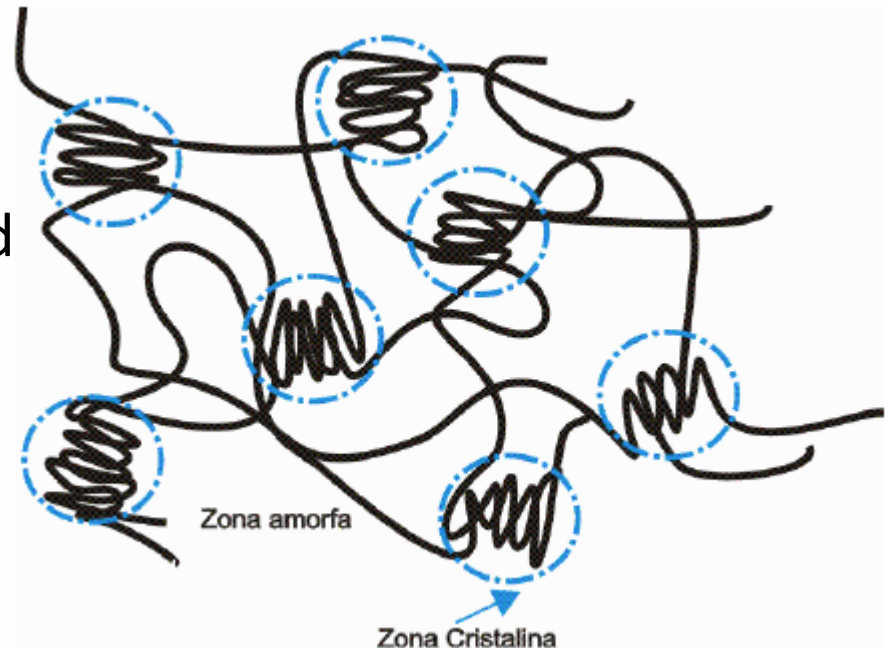
*With nanometric additions (nanocelulose, hidroxiapatite, iron oxide) produce composites with specific potential applications in biomedicina:*

# Hydrogels

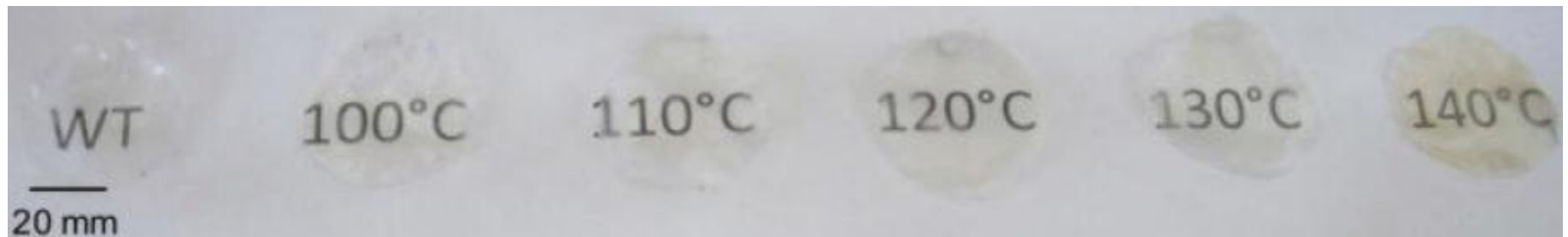
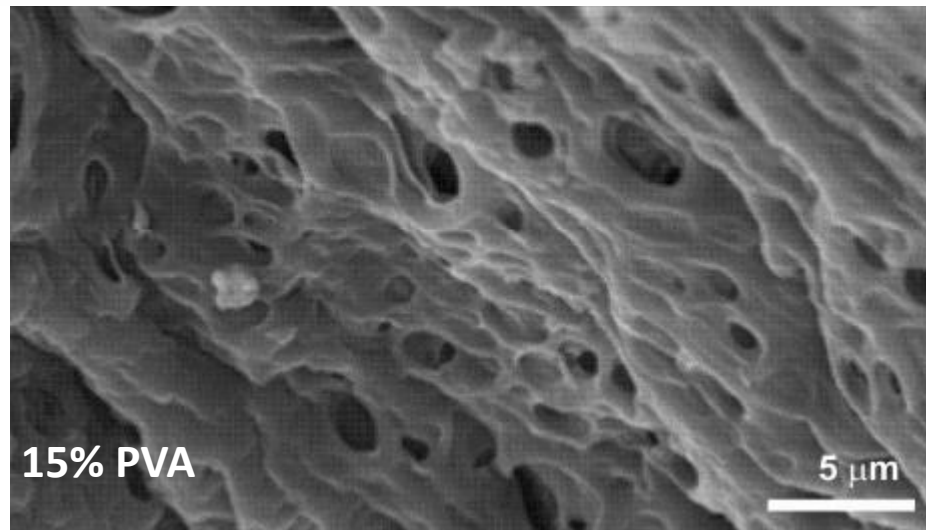
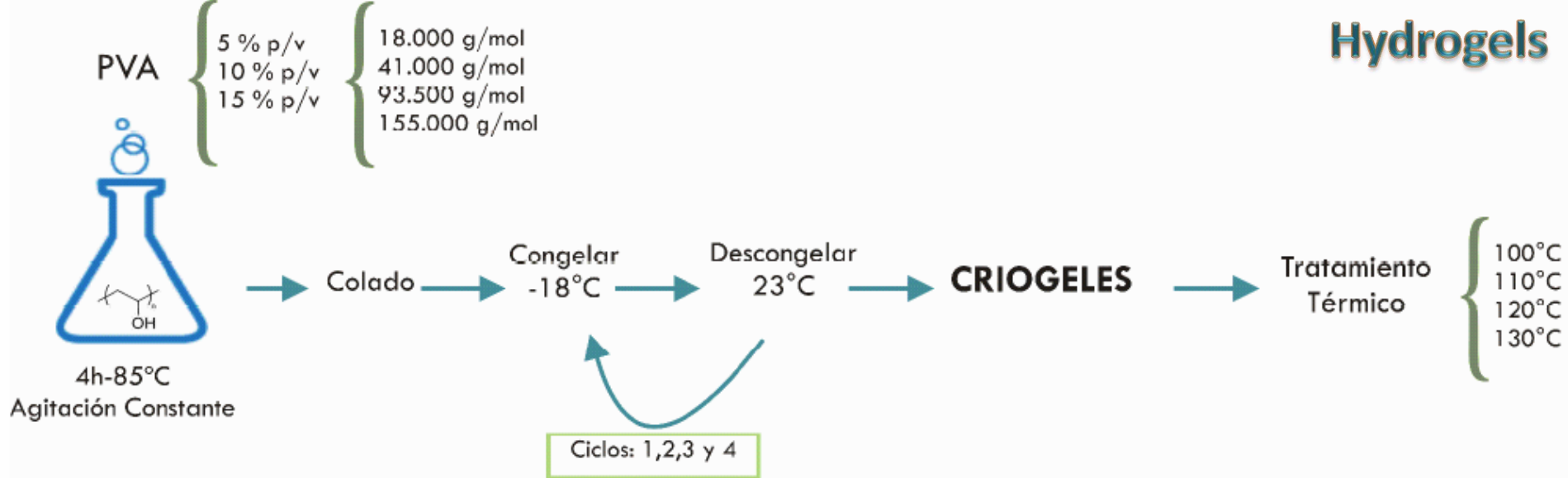


PVA

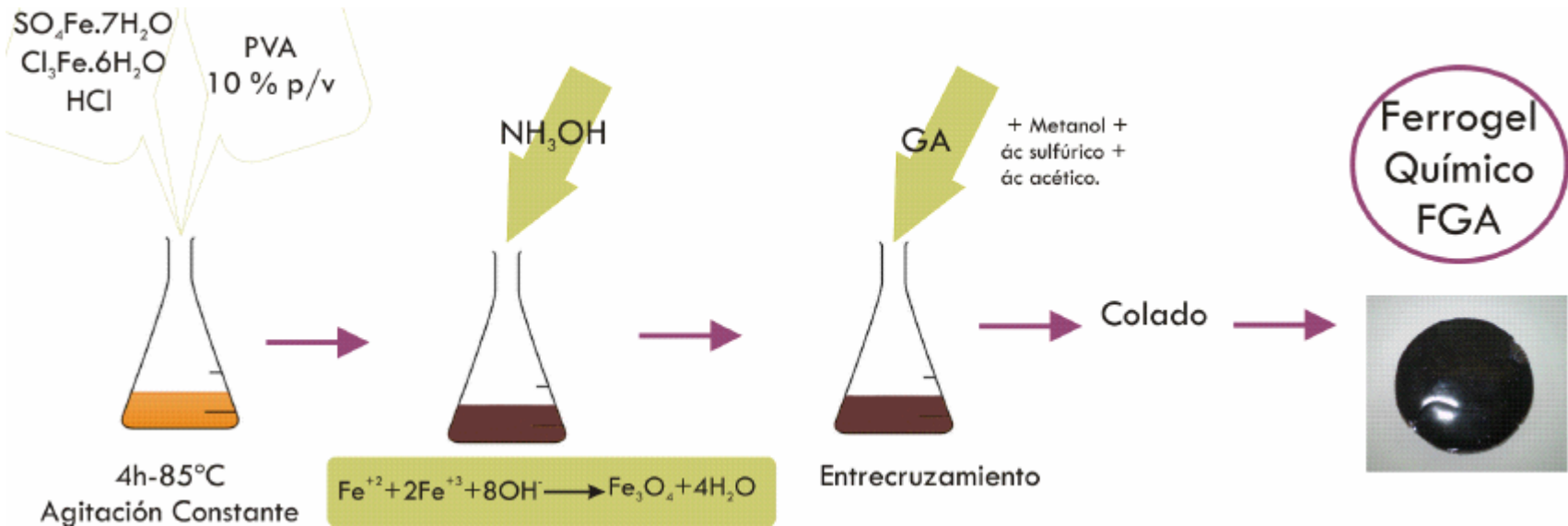
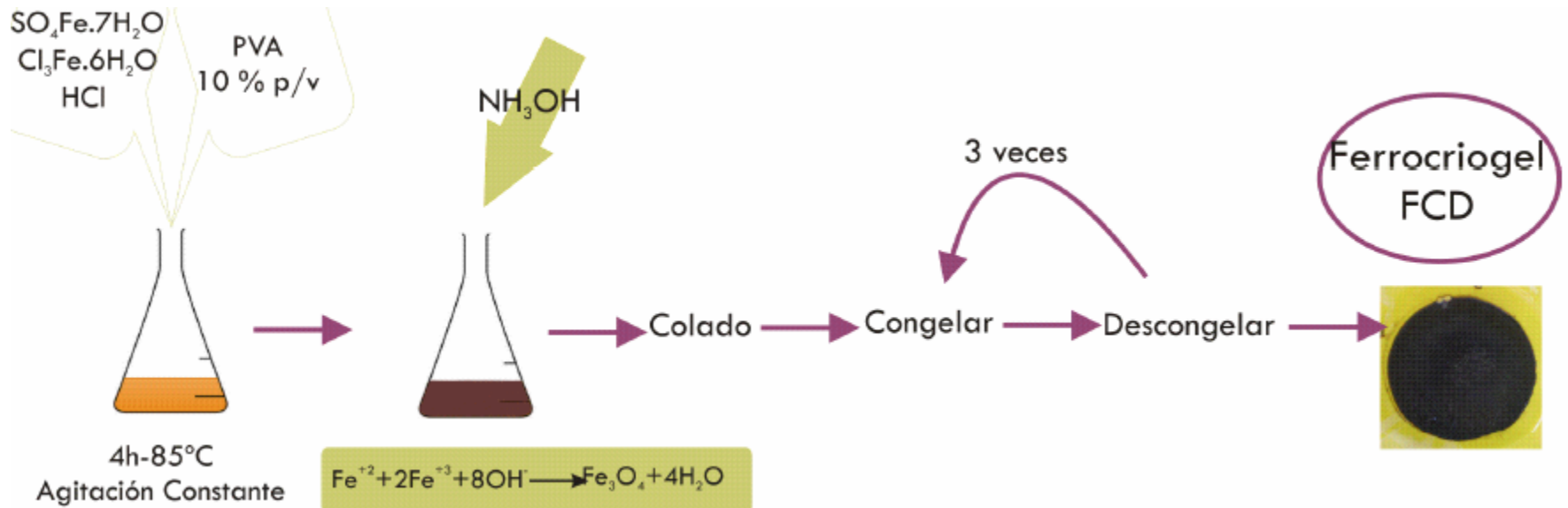
PVA hydrogel crosslinked  
by freezing – thawing  
cycling



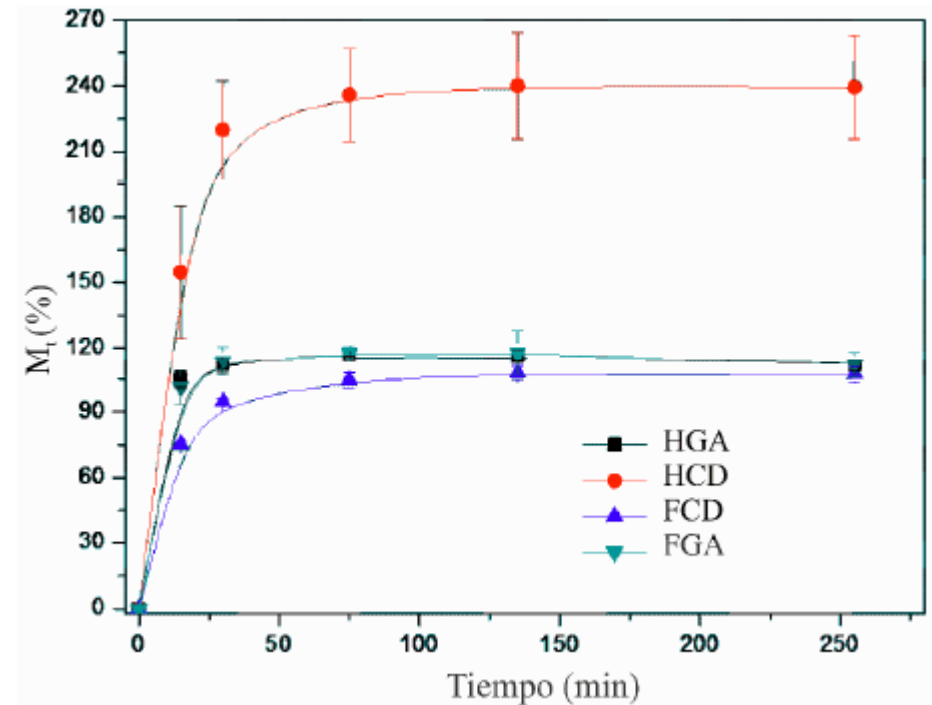
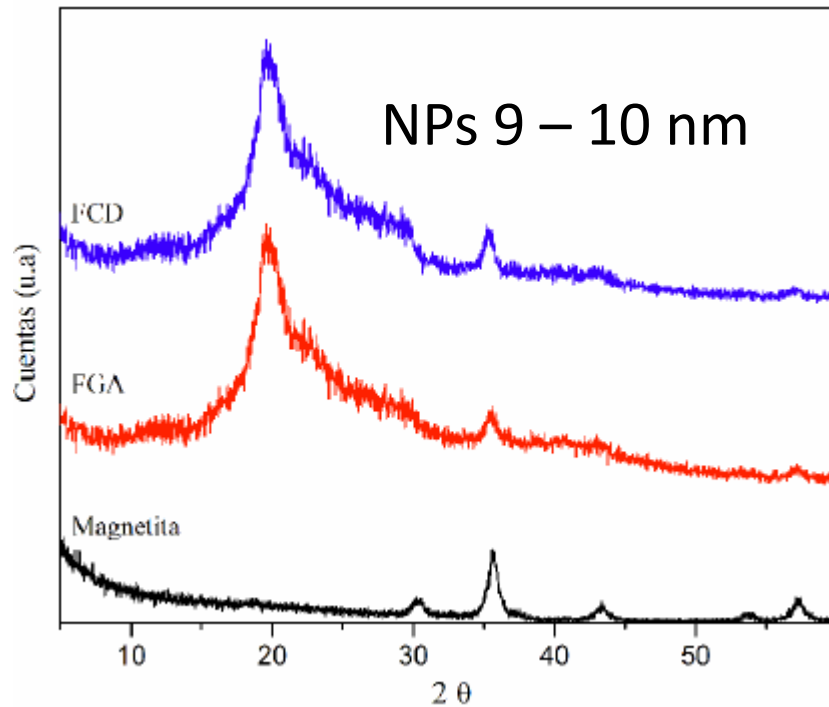
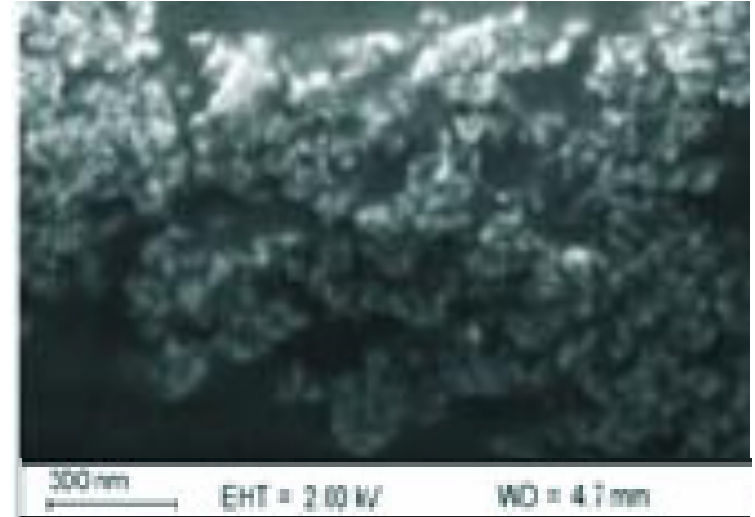
# Hydrogels



# Ferrogels (one pot synthesis)



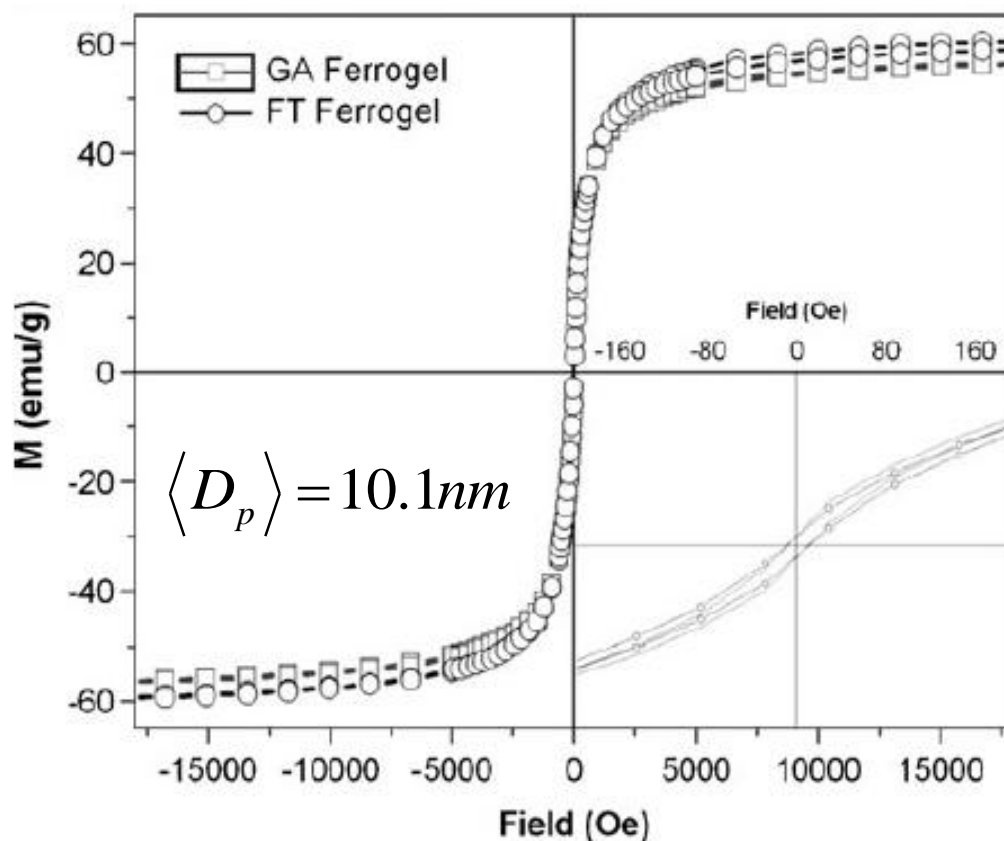
$$x_{\text{FeO}} \approx 0.06 \text{ (mass ratio)}$$



# Synthesis and characterization of PVA ferrogels obtained through a one-pot freezing–thawing procedure

Jimena S. Gonzalez • Cristina E. Hoppe •

Diego Muraca • Francisco H. Sánchez • Vera A. Alvarez Colloid Polym Sci (2011) 289:1839–1846



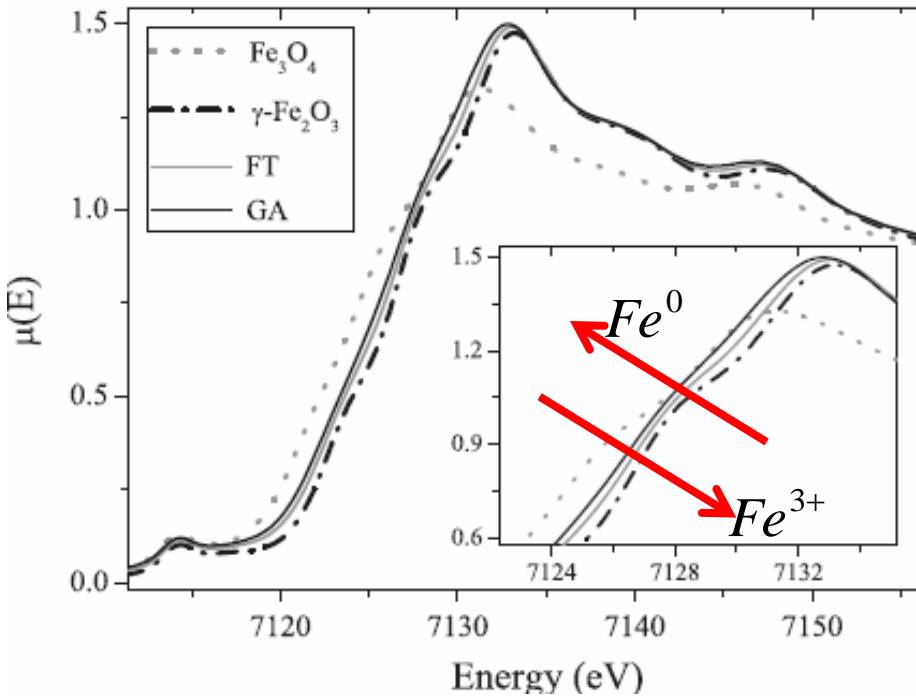
$$M = \frac{N \langle \mu \rangle \int_0^{\infty} \mu L(\alpha \mu) P(\mu) d\mu}{\int_0^{\infty} \mu P(\mu) d\mu} \quad \alpha = B/kT,$$

# Magnetic properties study of iron-oxide nanoparticles/PVA ferrogels with potential biomedical applications

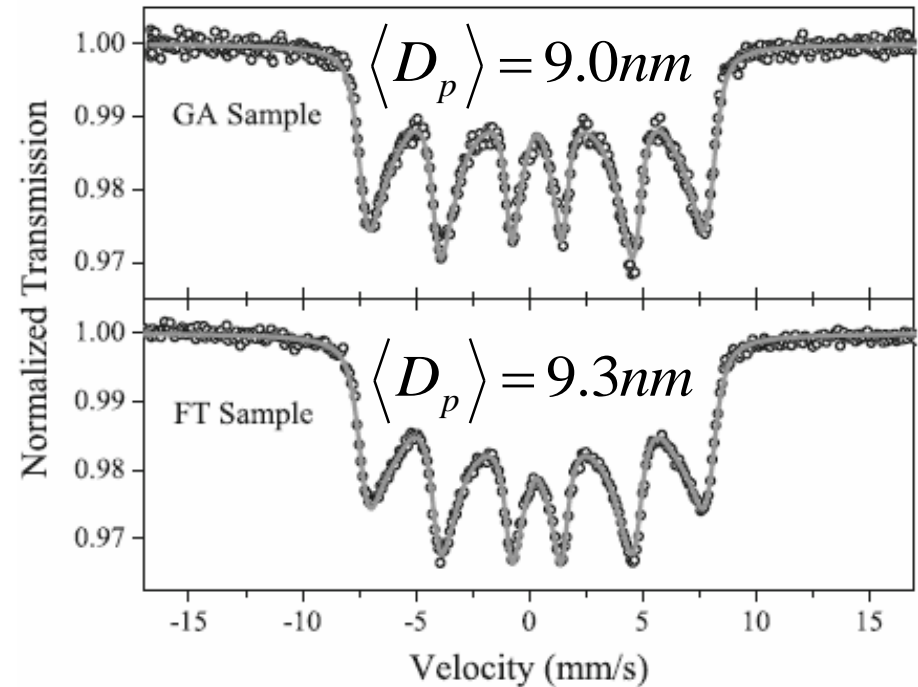
P. Mendoza Zélis · D. Muraca · J. S. Gonzalez ·  
G. A. Pasquevich · V. A. Alvarez · K. R. Pirota ·  
F. H. Sánchez

J Nanopart Res (2013) 15:1613

### Fe XANES

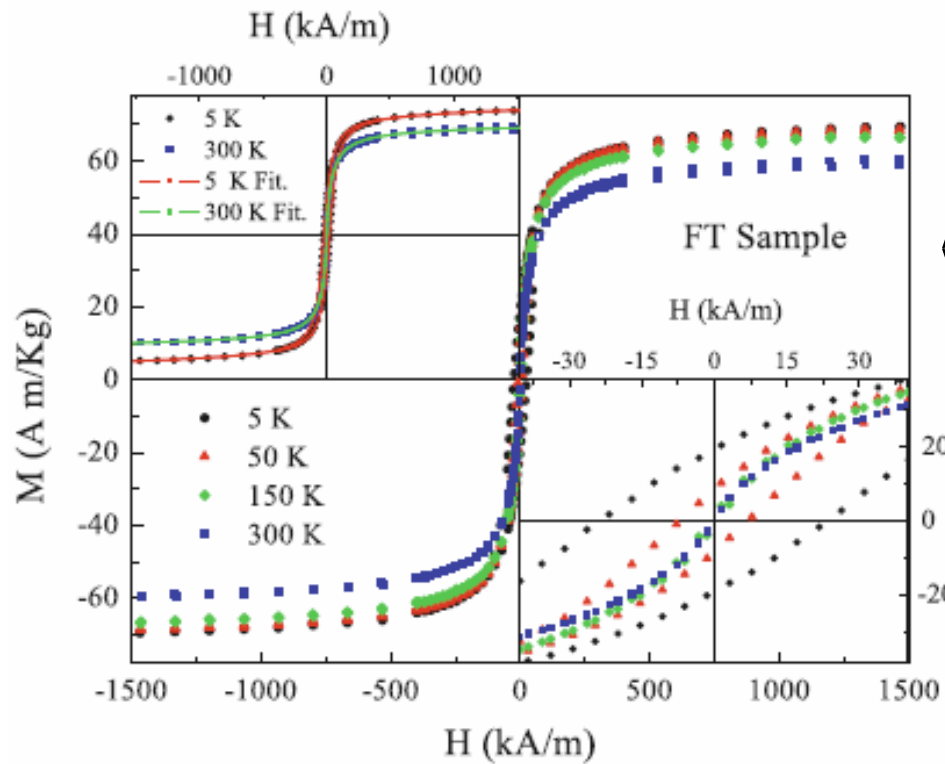


### $^{57}Fe$ Mössbauer

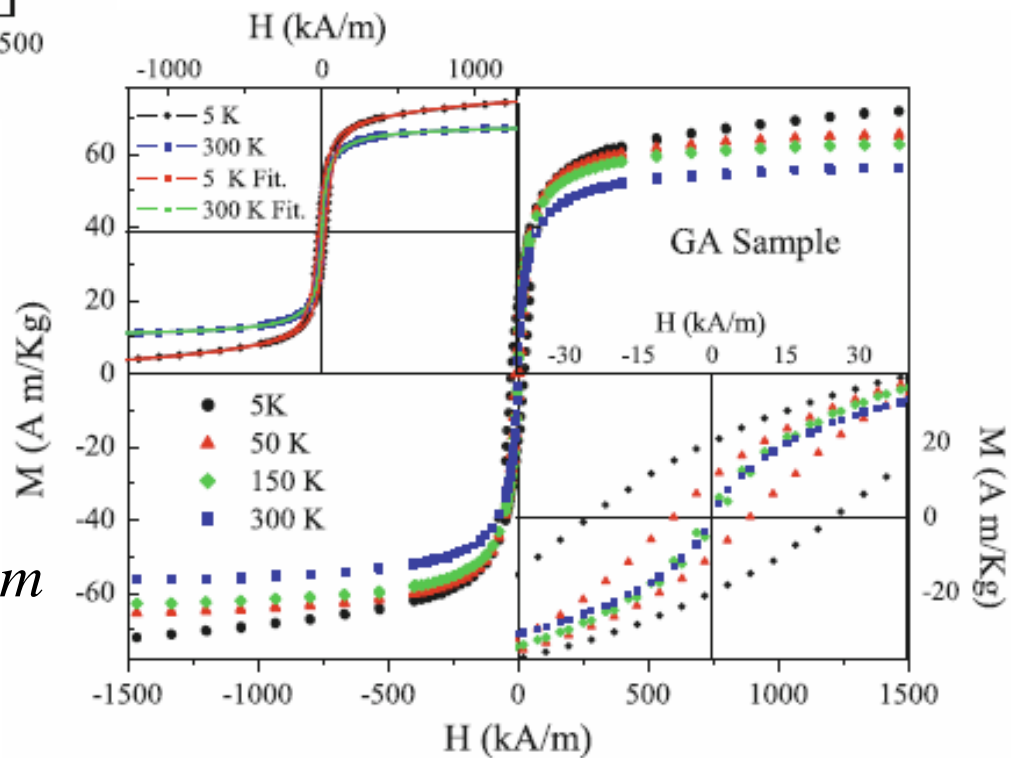


$$H_{hf}(V, T) \simeq H_{hf}(V = \infty, T) \left( 1 - \frac{k_B T}{2KV} \right)$$

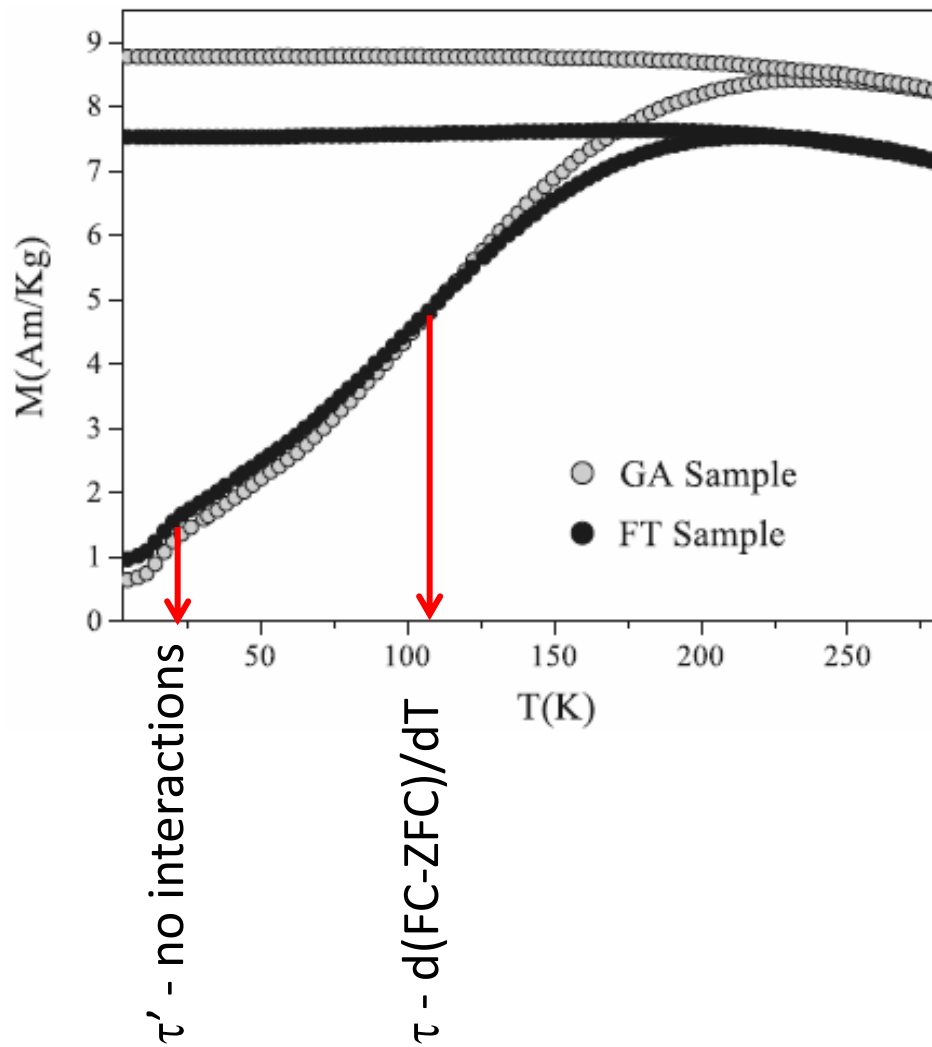
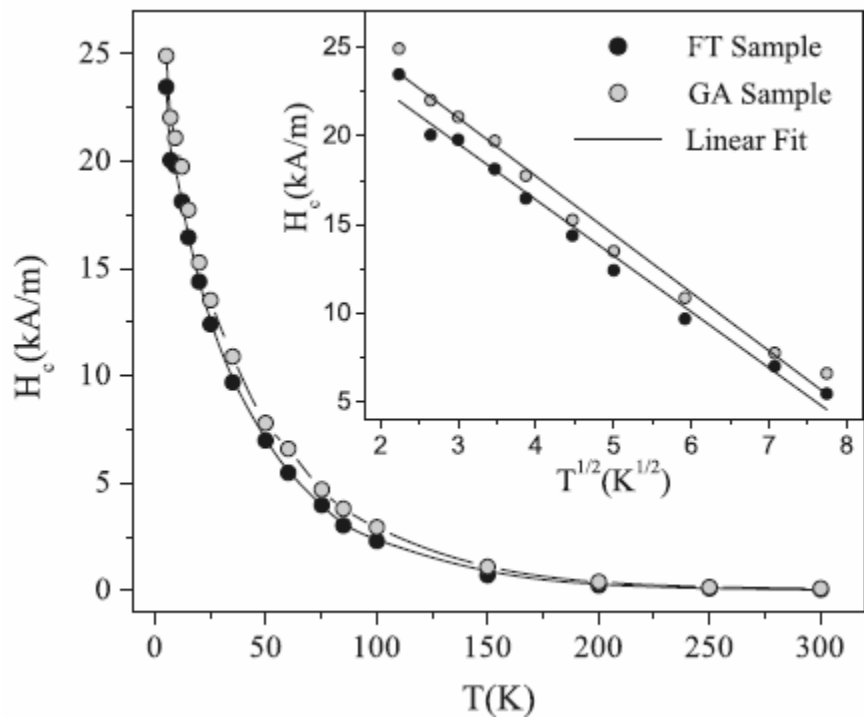




$$\langle D_p \rangle = 8.1nm$$



$$\langle D_p \rangle = 9.1nm$$



$$M(H, T) = \frac{N_a \langle \mu_a \rangle \int_0^\infty \mu_a L(b \mu_a) P(\mu_a) d\mu_a}{\int_0^\infty \mu_a P(\mu_a) d\mu_a} \quad b = \mu_0 H / kT$$

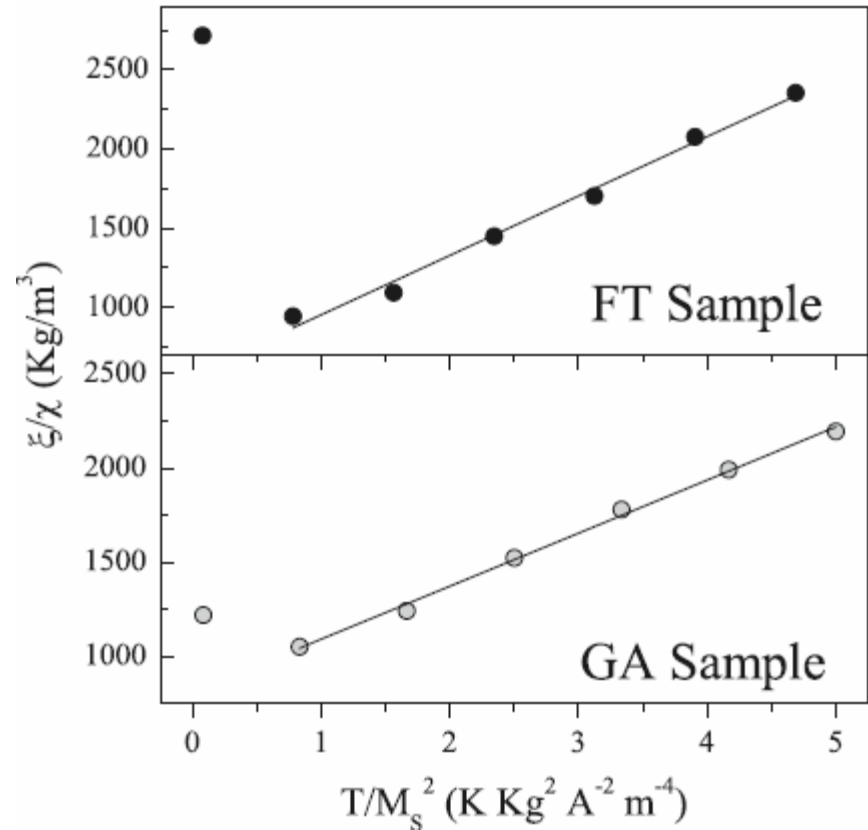
$$M(H, T) = \frac{N \langle \mu \rangle \int_0^\infty \mu L(b^* \mu) P(\mu) d\mu}{\int_0^\infty \mu P(\mu) d\mu} \quad b^* = \mu_0 H / k(T + T^*)$$

Allia et al.

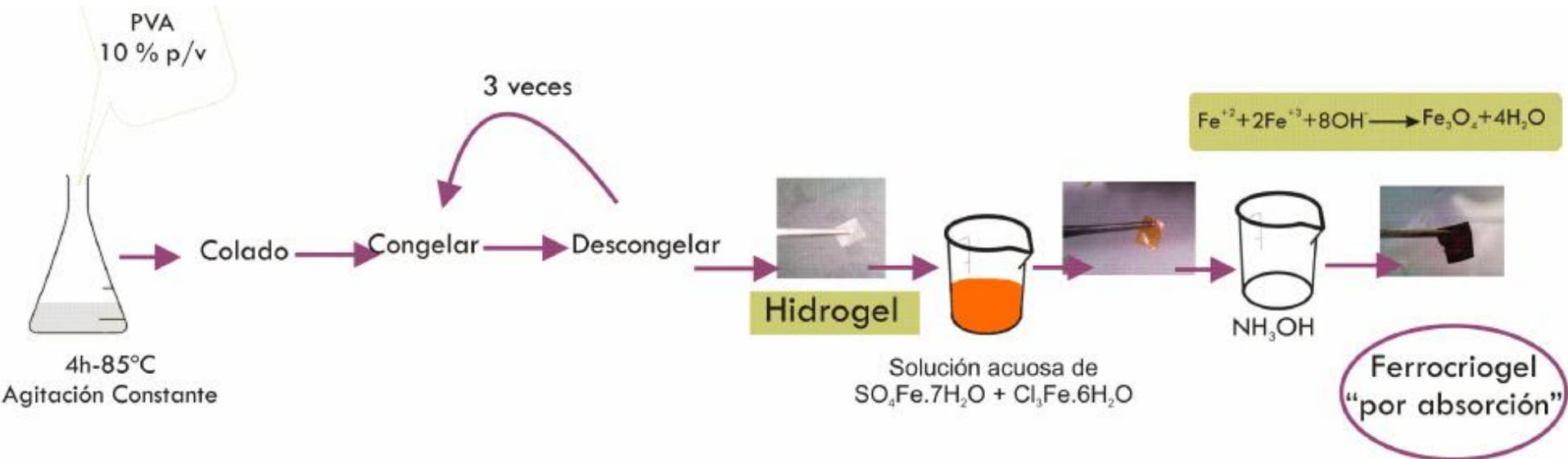
$$\frac{\xi}{\chi} = \frac{3kN}{\mu_0} \left( \frac{T}{M_s^2} \right) + 3\alpha$$

$$\xi = \frac{\langle \mu^2 \rangle}{\langle \mu \rangle^2} = \frac{\langle \mu_a^2 \rangle}{\langle \mu_a \rangle^2}$$

$$\varepsilon_d = \alpha \frac{\mu_0 \mu^2}{d^3}$$



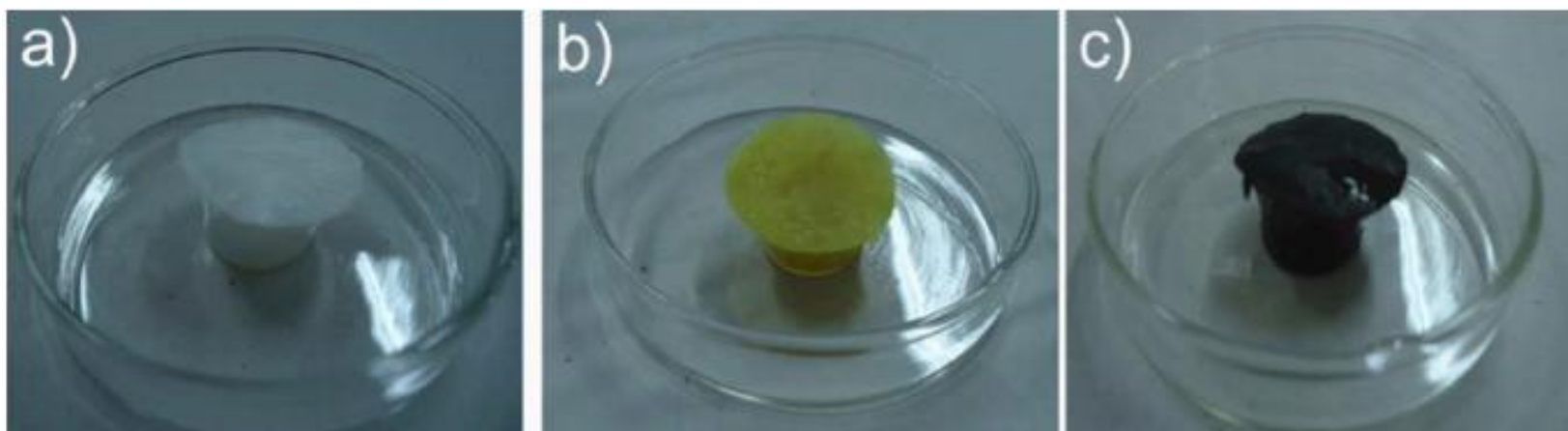
# Ferrogels (impregnation synthesis)



# A SIMPLE AND EFFICIENT PROCEDURE FOR THE SYNTHESIS OF FERROGELS BASED ON PHYSICALLY CROSSLINKED PVA

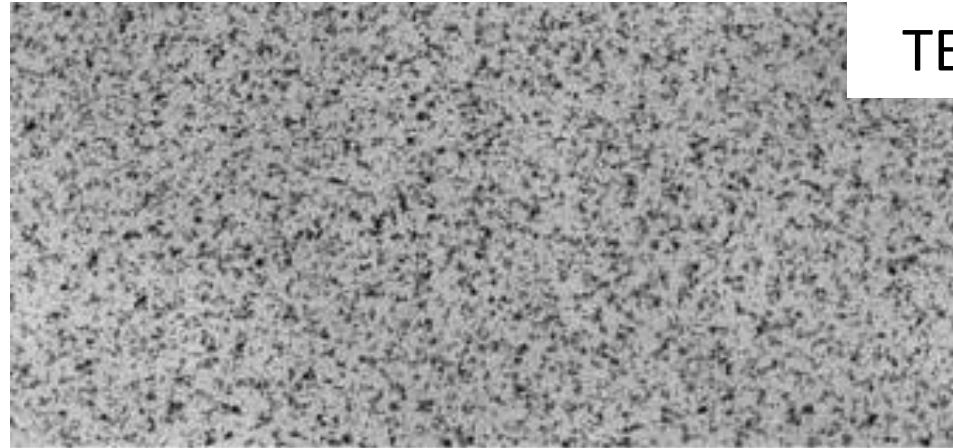
Jimena S. Gonzalez<sup>a\*</sup>, Cristina E. Hoppe<sup>a</sup>, Pedro Mendoza Zélis<sup>b</sup>, Lorena Arciniegas<sup>b</sup>,  
Gustavo A. Pasquevich<sup>b</sup>, Francisco H. Sánchez<sup>b</sup> and Vera A. Alvarez<sup>a</sup>

Industrial & Engineering Chemistry Research (under evaluation)

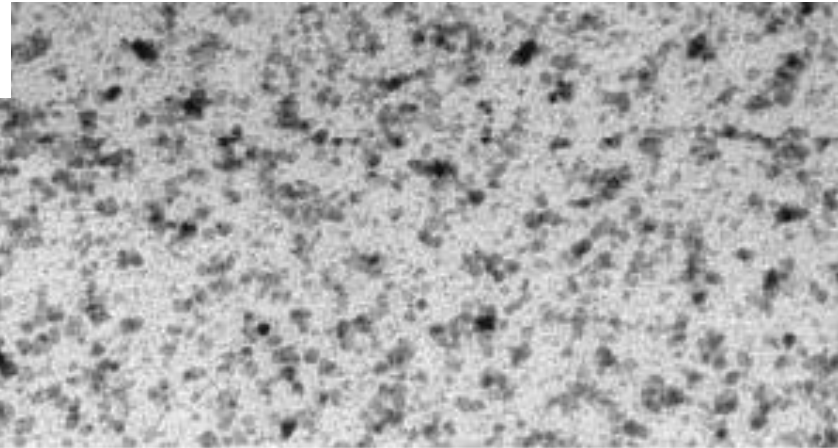


	$T_m$ °C	$X_{cr}$ %	$T_p$ °C	$w_t$ % $Fe_2O_3$
<b>Hydrogel</b>	220.5±0.3	31.80±3.2	284.3±5.3	0
<b>Ferrogel</b>	223.7±0.6	35.3±5.1	295.9±6.4	15.7±3.1

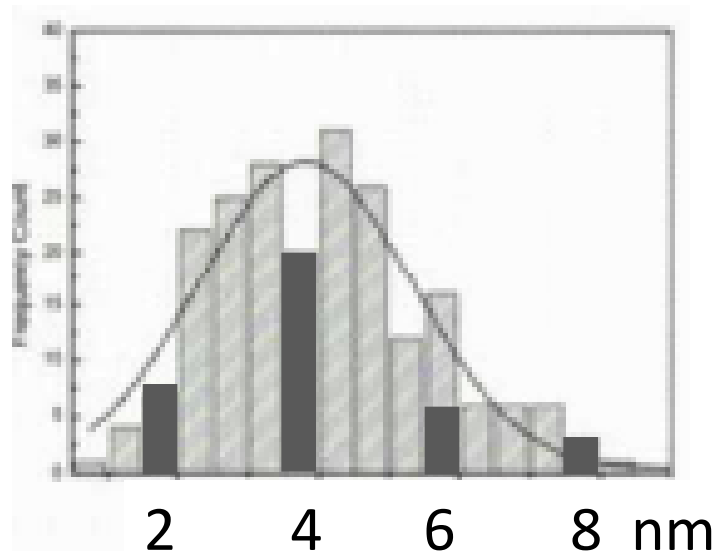
TEM

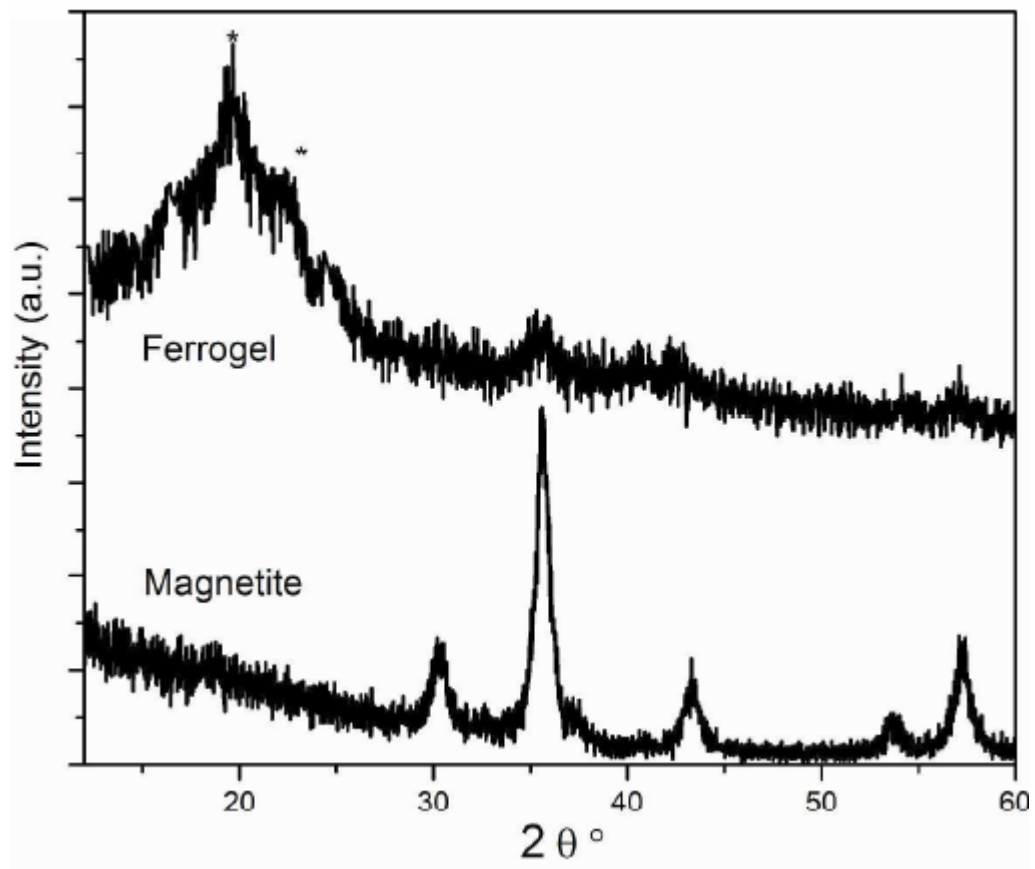


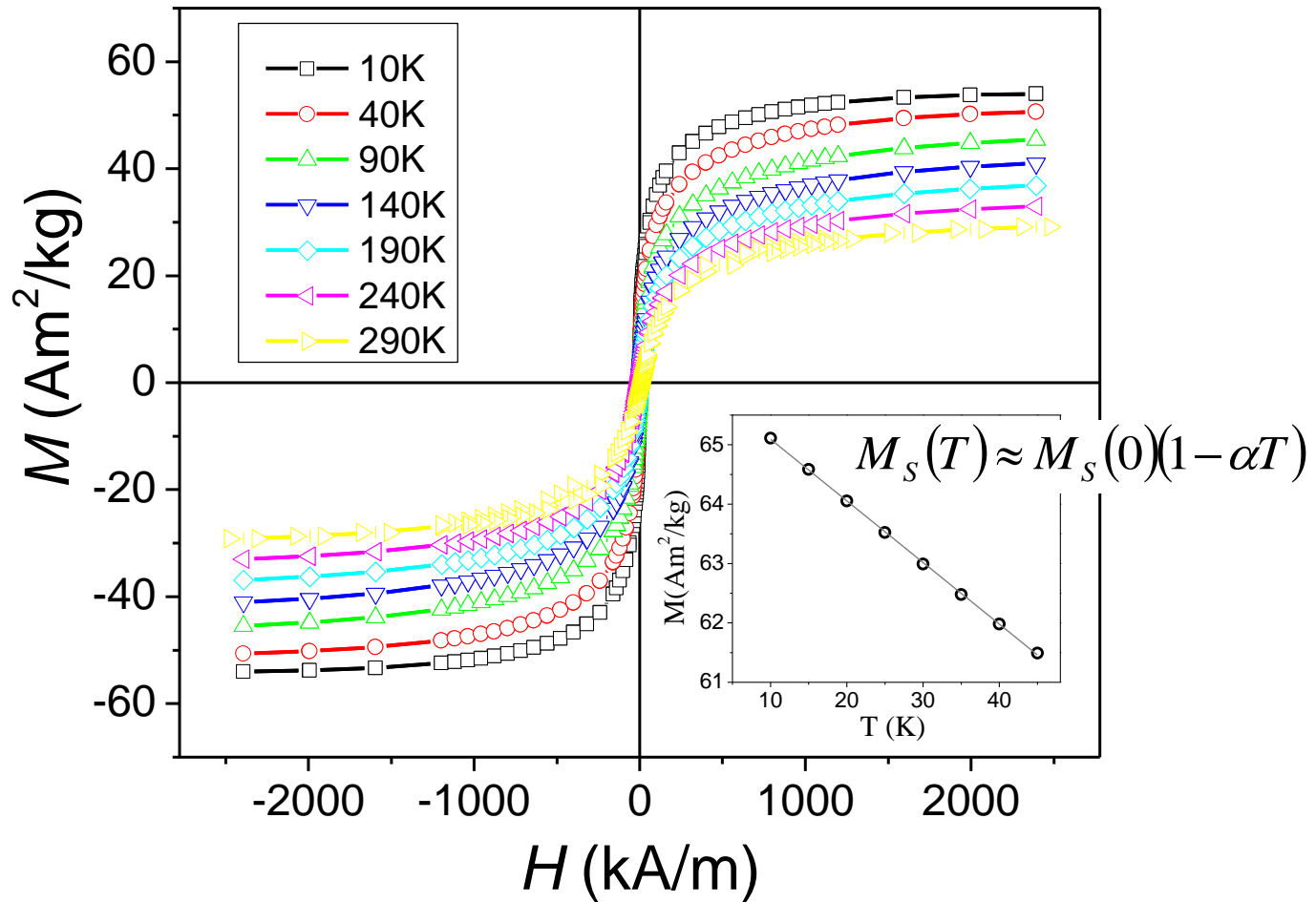
200 nm



50 nm

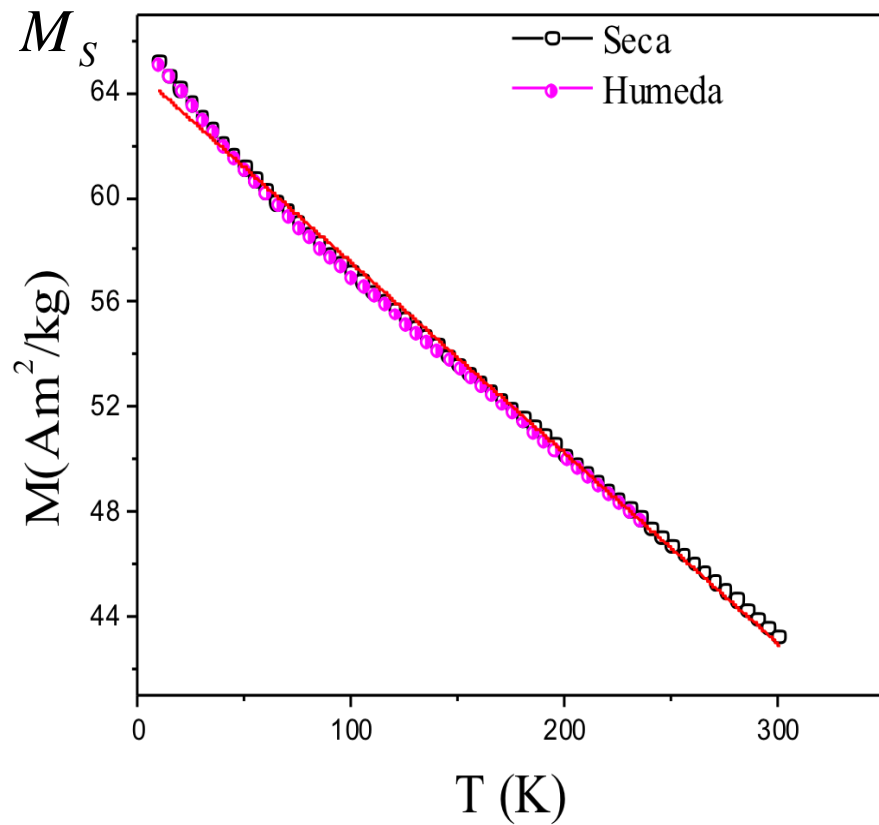






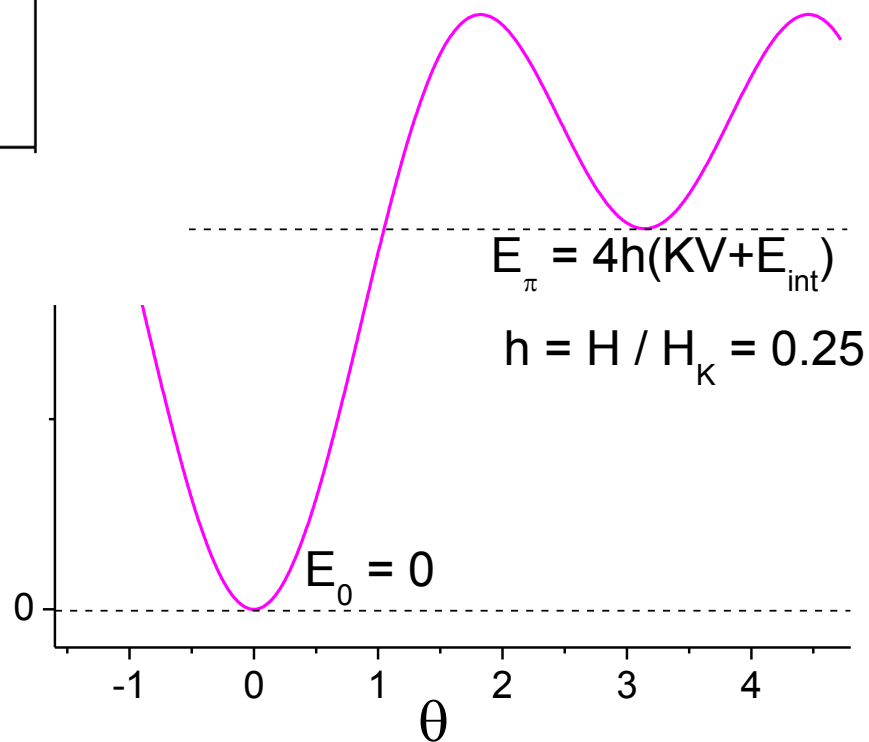
$$H_c = 0 \quad \text{for} \quad T \geq 40\text{K}$$



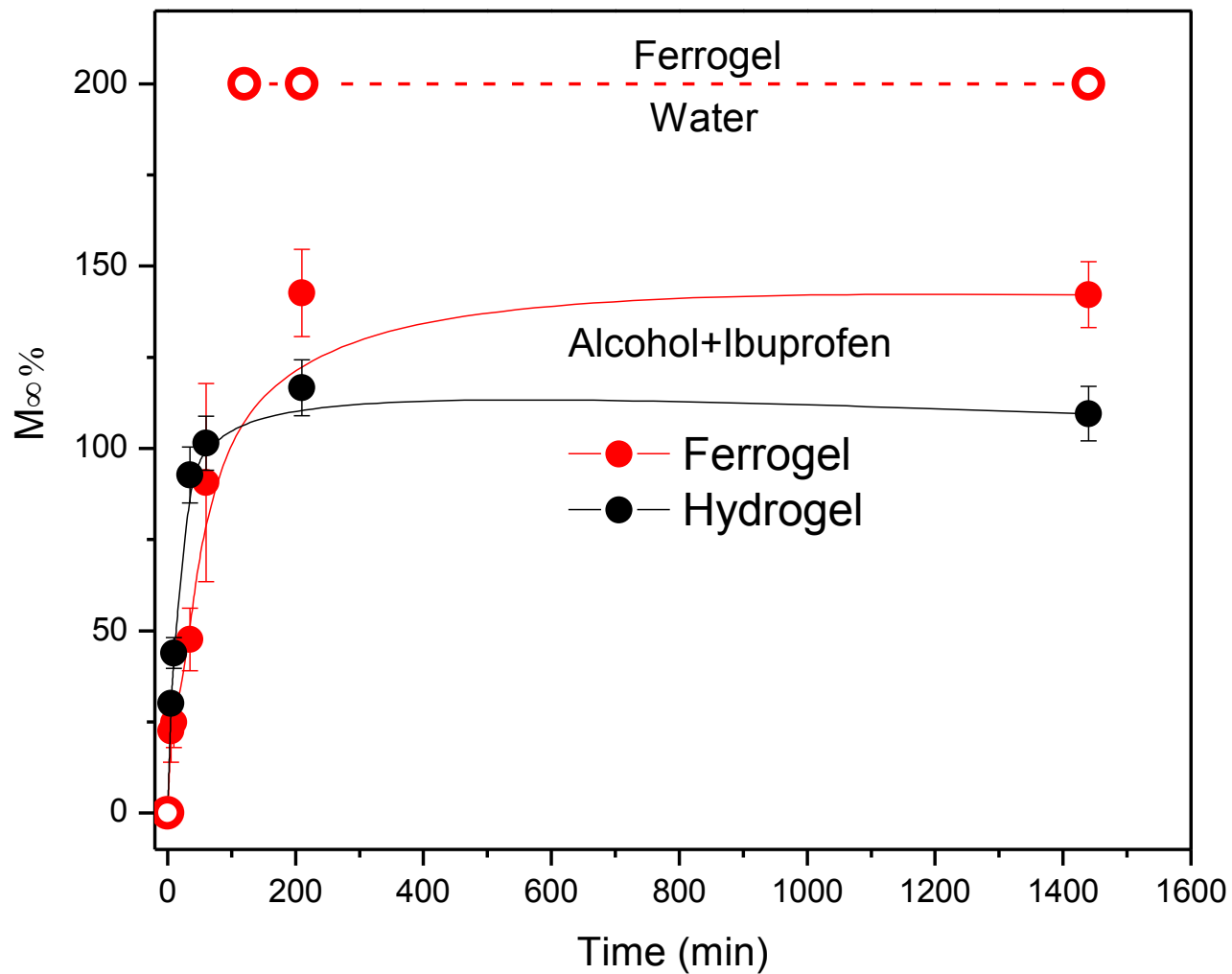


$$M_S(T) \approx M_S(0)(1 - \alpha T)$$

$$\alpha \approx k / 8(KV + E_{\text{int}})$$



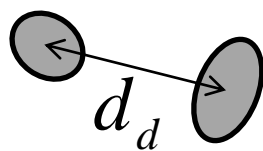
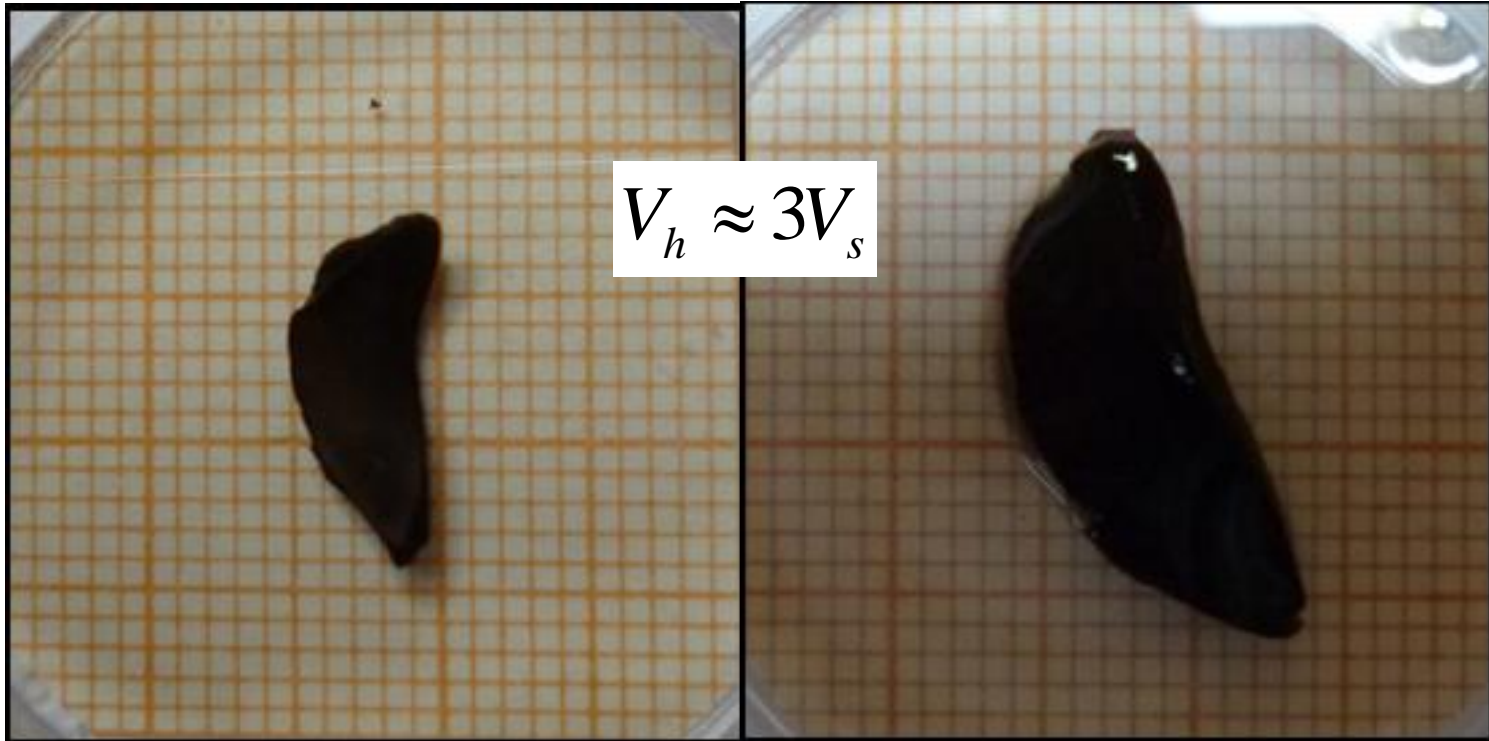
# Ferrogel swelling



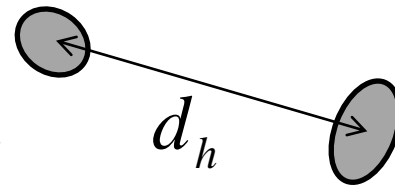
# Ferrogel swelling

dry

hydrated



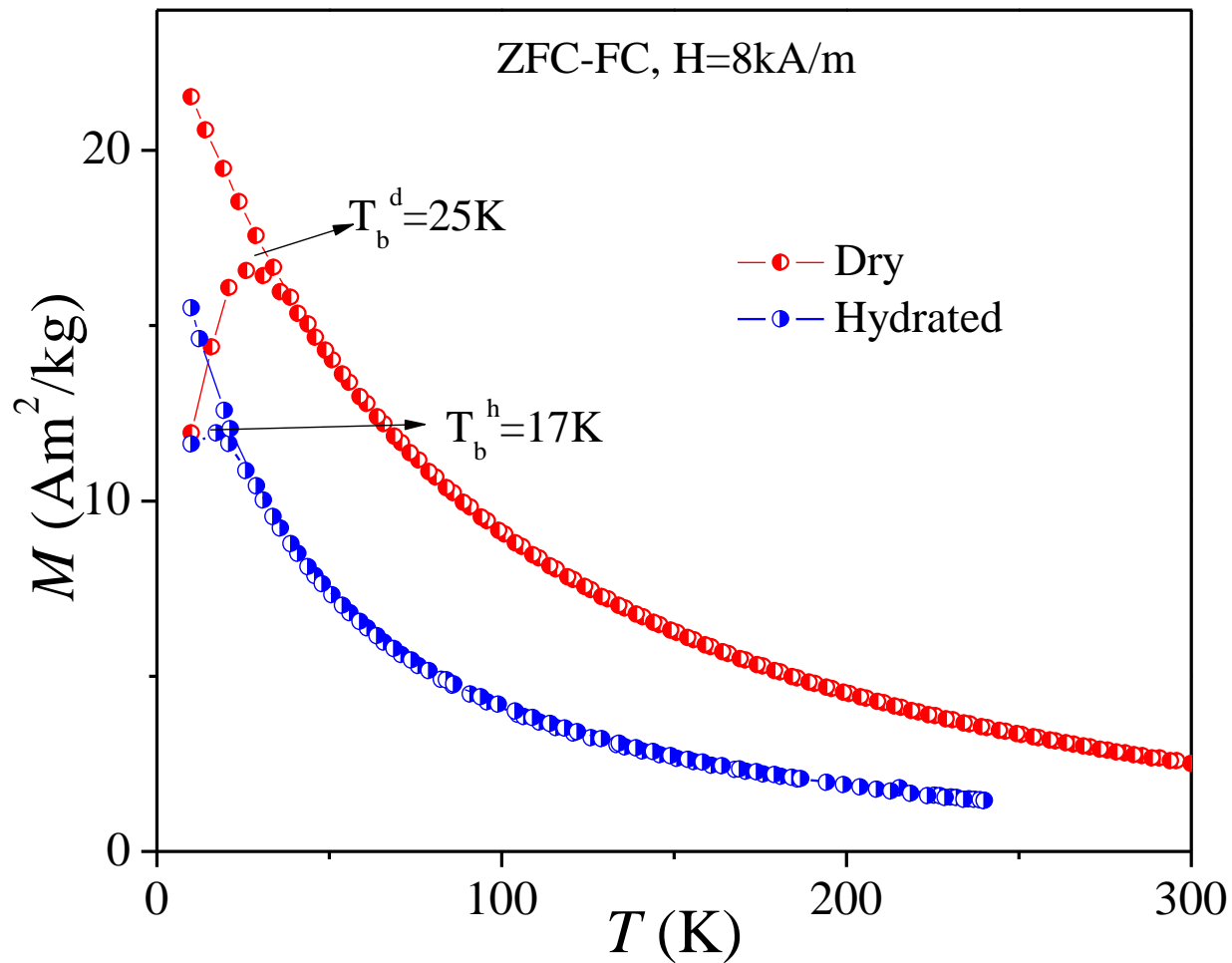
$$d_h \approx 1.44 d_d$$



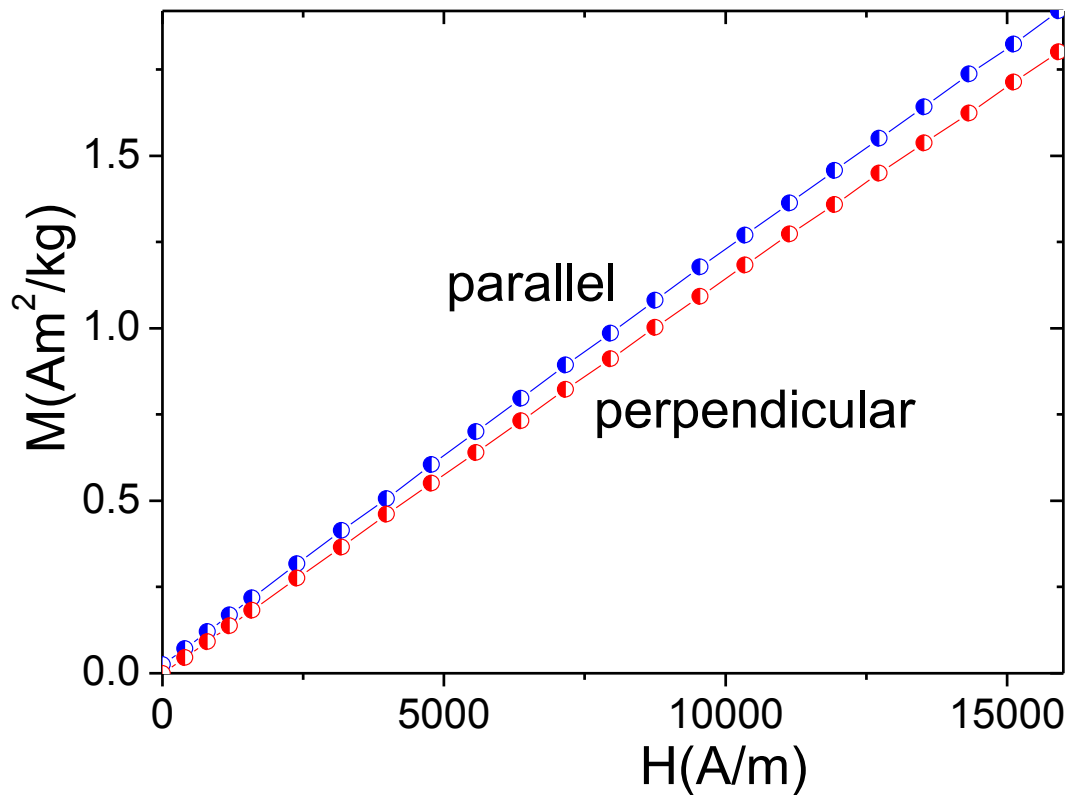
$$\tau_{seco} \approx \tau_0 e^{(KV + E_{int}^{seco})/kT}$$

$$\tau_{hid} \approx \tau_0 e^{(KV + E_{int}^{hid})/kT}$$

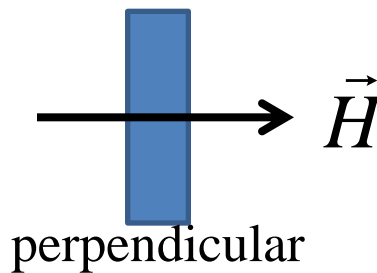
$$E_{int}^{seco} > E_{int}^{hid} \Rightarrow T_B^{seco} > T_B^{hid}$$



# Shape Effects



➔  $\gamma \approx 2.1$



# Response to a Magnetic RF Field

Optical Probe



Water



Ferrogel



Dewar



Duty Coil

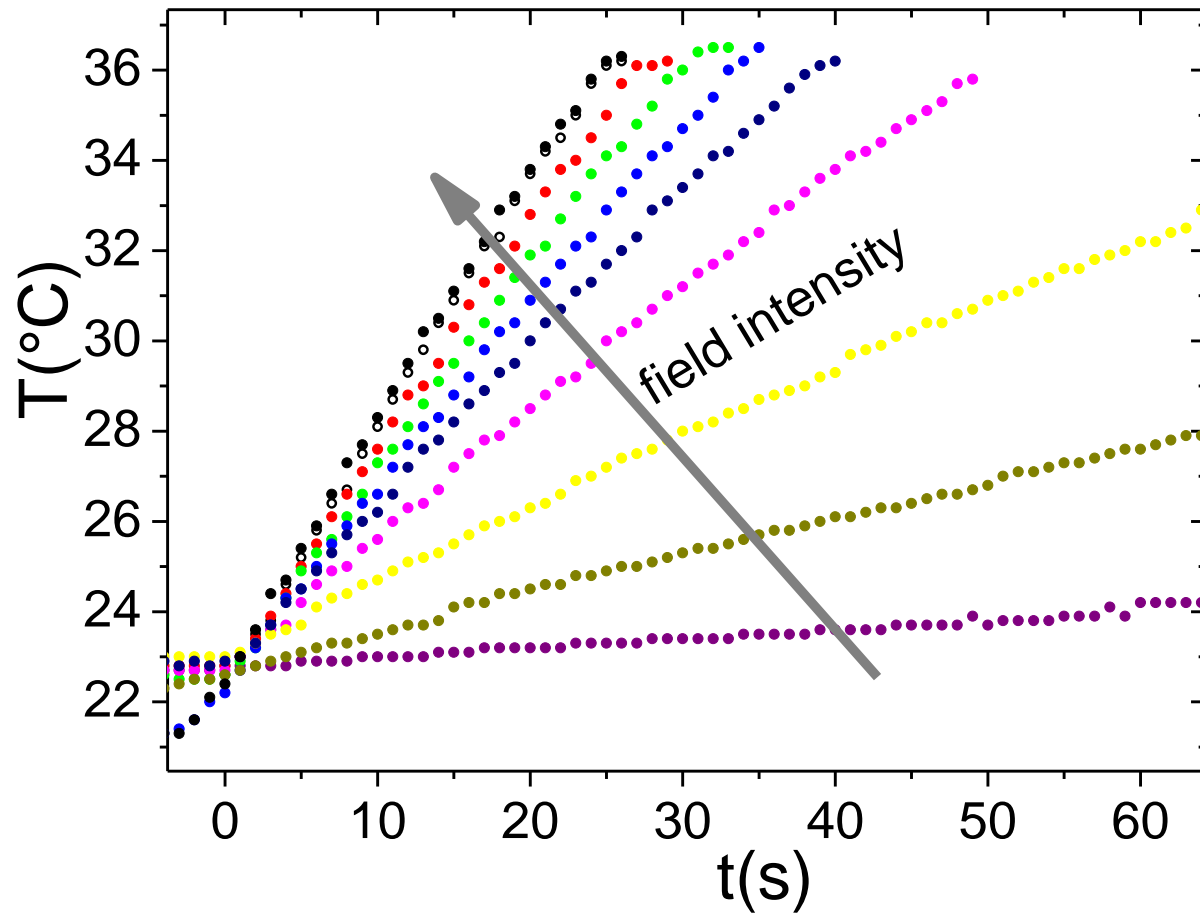


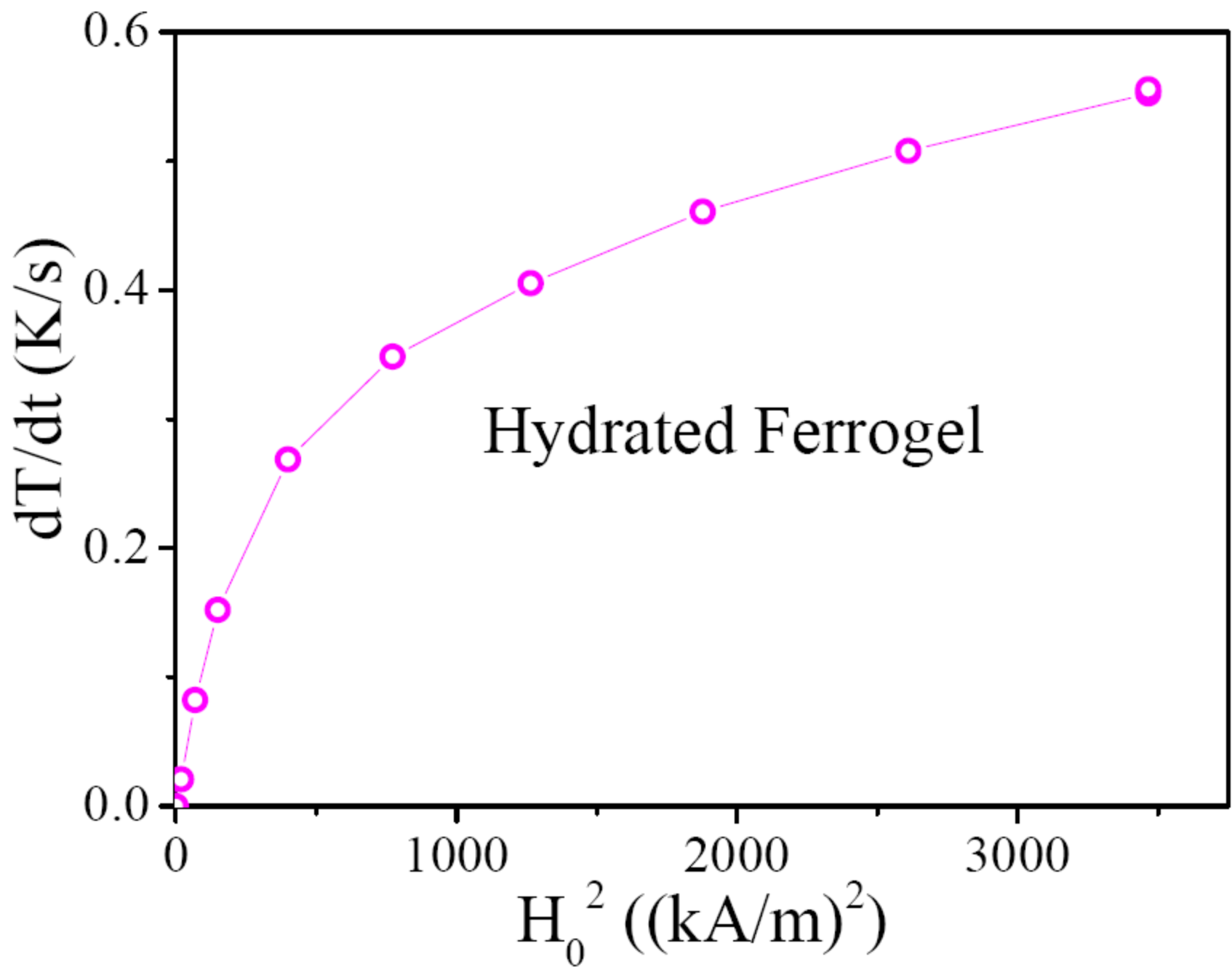
$$f = 260 \text{ kHz}$$

# Response to a Magnetic RF Field

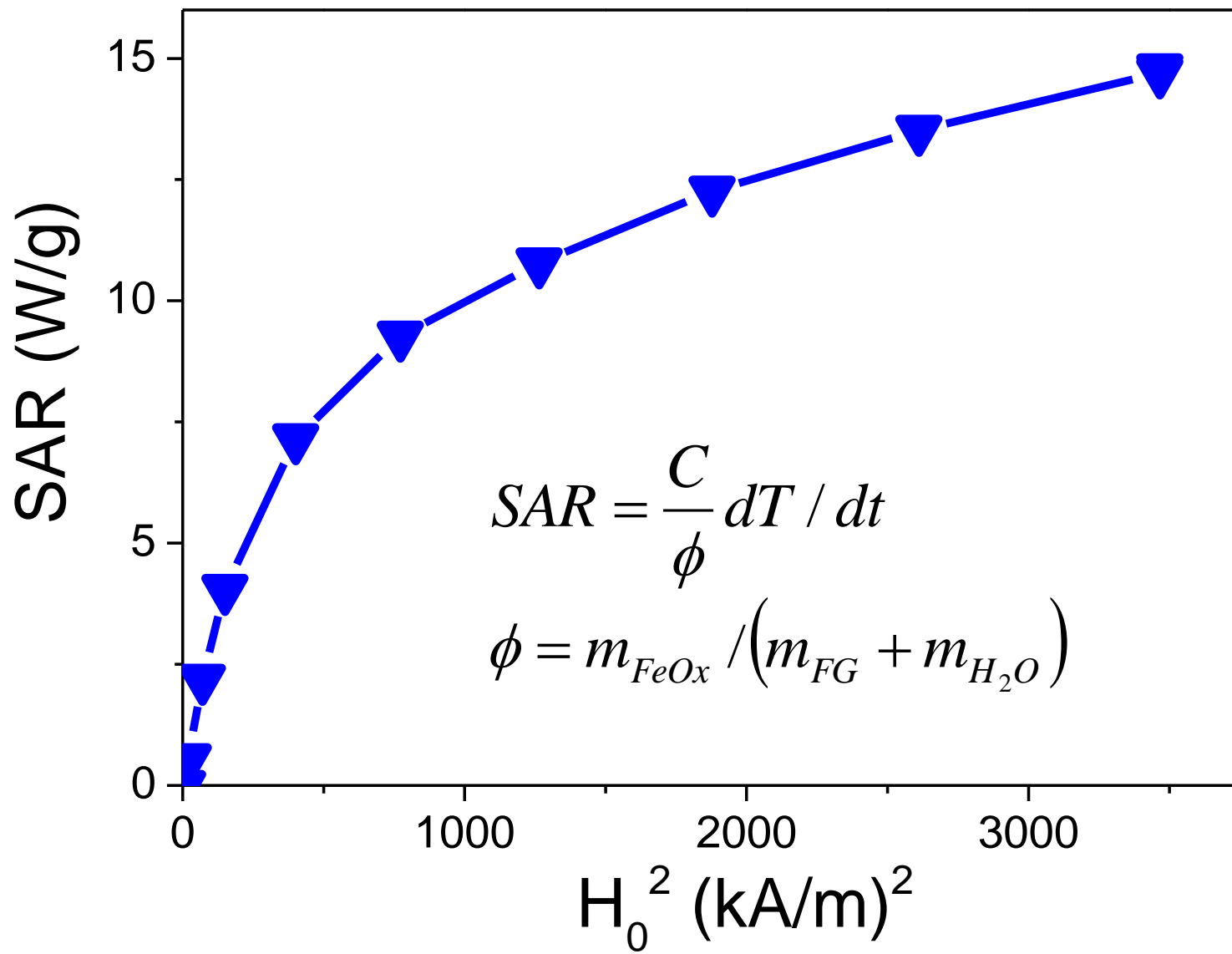
$$f = 260 \text{ kHz}$$

$$H_0 = 3.83 \text{ kA/m} - 58.2 \text{ kA/m}$$

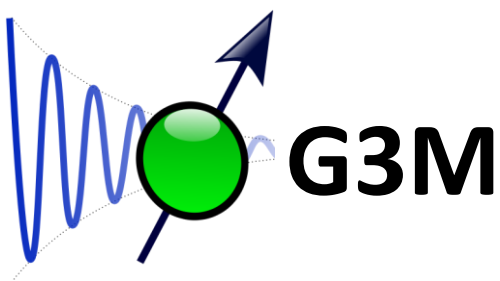








Muito obrigado!



**G3M**

*Marcela B. Fernández van Raap*

*Pedro Mendoza Zélis*

*Gustavo A. Pasquevich*

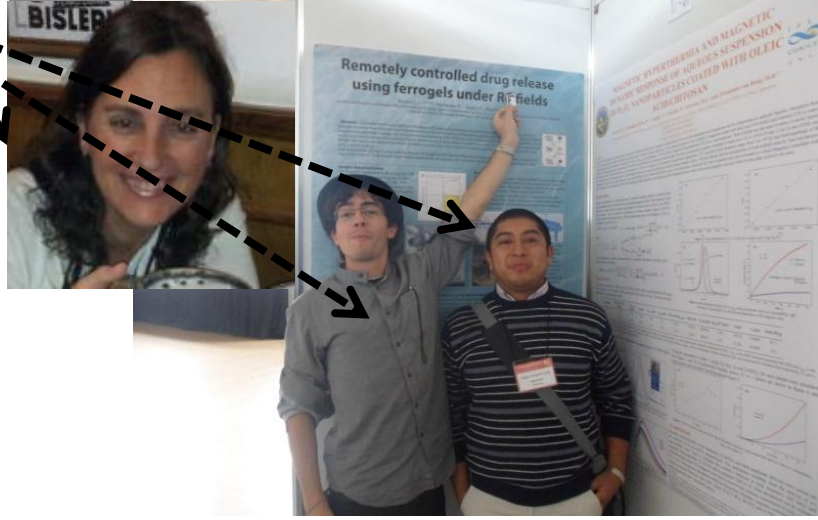
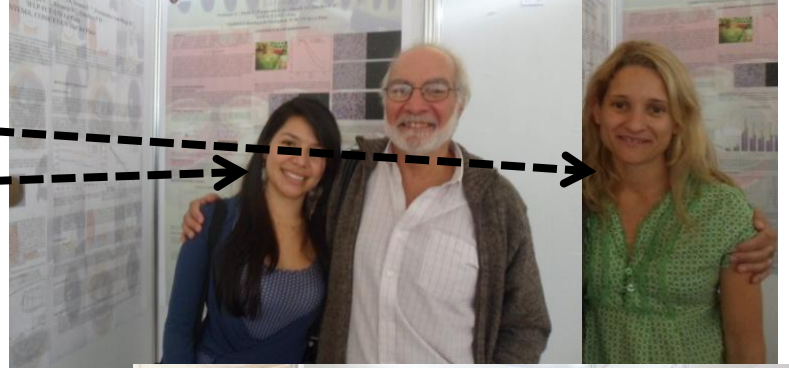
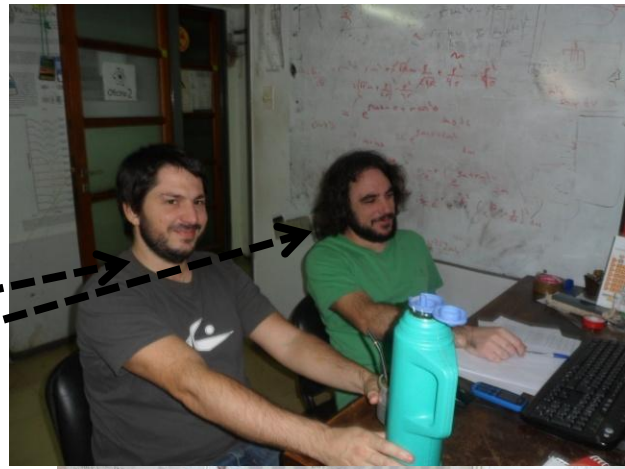
*Ignacio Bruvera*

*Elisa de Sousa*

*Diego Coral*

*Lorena Arciniegas*

Francisco H. Sánchez



e-mail: [sanchez@fisica.unlp.edu.ar](mailto:sanchez@fisica.unlp.edu.ar)

**BioMag Nanomaterials**

# Subjects & People

## In-vitro magnetofection

**Olga Mykhailyc**  
**Rodolfo Goya**  
**Joaquín Pardo**  
Gustavo A. Pasquevich  
Lorena Arciniegas

## La-Mn-perovskites

Marcela B. Fernández  
van Raap  
Pedro Mendoza Zélis  
Gustavo A. Pasquevich  
**Gabriela Leyva**  
Ignacio Bruvera  
Cecilia Laborde

## Zn-doped magnetites

Marcela B. Fernández  
van Raap  
Pedro Mendoza Zélis  
Gustavo A. Pasquevich  
**Silvia Jacobo**  
**Juan Apesteguy**  
Ignacio Bruvera  
Cecilia Laborde  
Betiana Pianciola

## Co-Ferrites

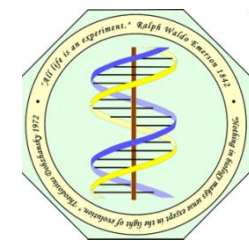
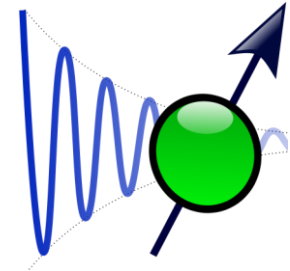
Marcela B. Fernández  
van Raap  
Pedro Mendoza Zélis  
Gustavo A. Pasquevich  
**Gerardo Goya**  
**Clara Marquina**  
**Teo Torres**  
Diego Coral

## Functionalized SDION cell internalization

Marcela B. Fernández  
van Raap  
Pedro Mendoza Zélis  
Gustavo A. Pasquevich  
**Ricardo Dewey**  
**Nolberto Martínez**  
**Verónica Lasalle**  
Elisa de Sousa  
Diego Coral  
J.L. Alessandrini

## PVA Ferrogels

**Cristina Hoppe**  
**Vera Alvarez**  
**Jimena González**  
Pedro Mendoza Zélis  
Gustavo Pasquevich  
Lorena Aciniegas



# G3M

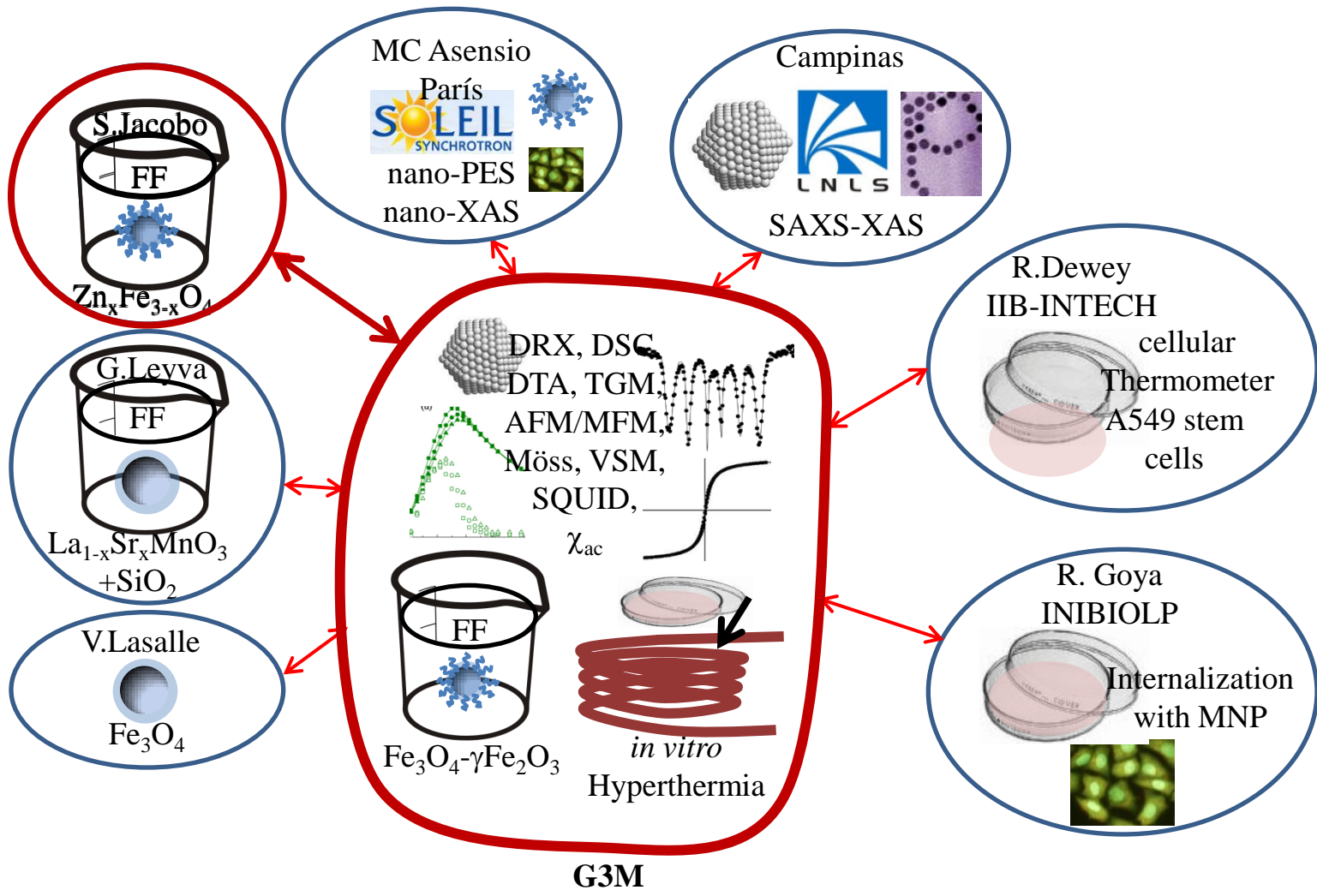
e-mail: [sanchez@fisica.unlp.edu.ar](mailto:sanchez@fisica.unlp.edu.ar)

## Recent related papers

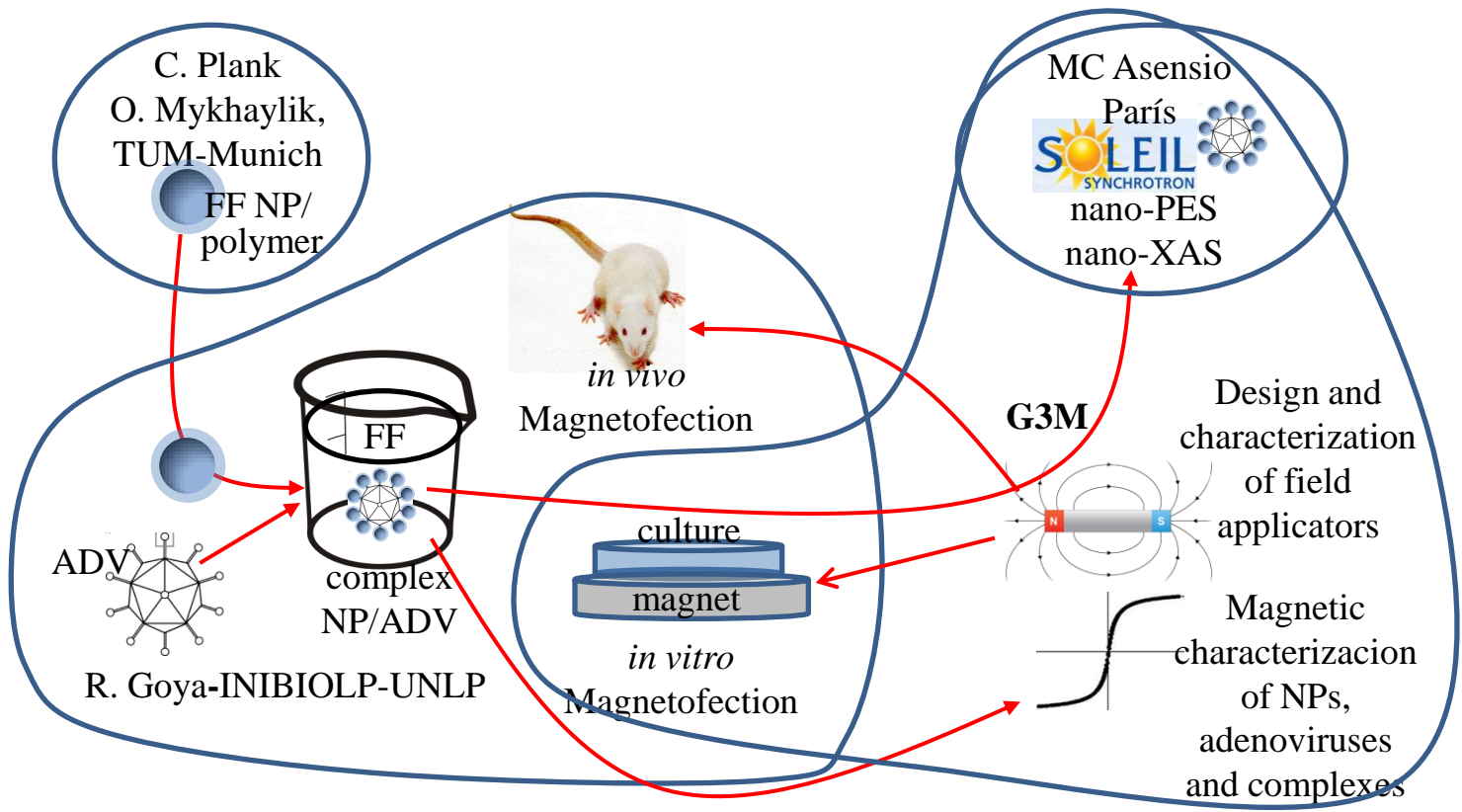
- Synthesis and Characterization of PVA Ferrogels obtained through a One-pot Freezing-Thawing Procedure, *JS Gonzalez, CE Hoppe, D Muraca, FH Sánchez and VA Alvarez*, **COLLOID & POLYMER SCIENCE**, Volume 289, Numbers 17-18, 1839-1846, (2011), DOI: 10.1007/s00396-011-2501-1
- Ascorbic Acid Encapsulation in Hydrophobic Silica Xerogel, *MV Revuelta; MB Fernández van Raap; P Mendoza Zélis; FH Sánchez; GR Castro*, **Food Technol. Biotechnol.** 49 (3) 347–351 (2011)
- Self organization in oleic acid-coated CoFe<sub>2</sub>O<sub>4</sub> colloids: a SAXS study, *MB Fernández van Raap Mendoza Zélis P, Coral, DF, TETorres, C Marquina, GF Goya, and FHSánchez*, **J Nanopart Res** (2012) 14:1072-1, 1072-5
- Magnetic field-assisted gene transfer: Studies in glial cells, **Acta Bioquímica Latinoamericana**, *Pardo J, Sosa YE, Reggiani PC, Arciniegas ML, Sánchez FH y Goya RG*; ISSN 1851-6114, 47, 399-406 (2013).
- Structural and magnetic study of zinc-doped magnetite nanoparticles and ferrofluids for hyperthermia applications, *P Mendoza Zélis, GA Pasquevich, SJ Stewart, MB Fernández van Raap, J Apesteguy, IJ Bruvera, C Laborde, B Pianciola, S Jacobo, FH Sánchez*, **J. Phys. D: Appl. Phys.** 46, (2013), 125006 doi:10.1088/0022-3727/46/12/125006.
- Stability and relaxation mechanisms of citric acid coated magnetite nanoparticles for magnetic hyperthermia, *de Sousa M, Fernandez van Raap M, Rivas P, Mendoza Zélis P, Girardin P, Pasquevich G, Alessandrini J, Muraca D, Sánchez FH*, **J. Phys. Chem. C**, (2013), 117 (10), pp 5436–5445. DOI: 10.1021/jp311556b.
- Magnetic properties study of PVA/iron oxide ferrogels with potential biomedical applications, *P Mendoza Zelis, D Muraca, JS Gonzalez, GA Pasquevich, VA Alvarez, KR Pirota, FH Sánchez*, **J Nanopart Res**, (2013) in the press.

THANKS!

# Collaborations scheme Hyperthermia



# Collaborations scheme Magnetofection

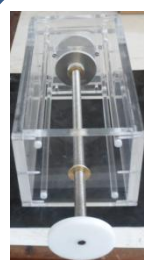
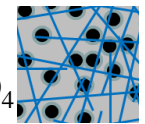
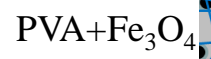


# Collaborations scheme Smart membranes

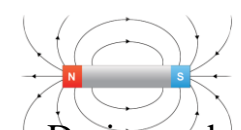
J. González, V. Alvarez,  
C. Hoppe, INTEMA



Hydrogel+NP=Ferrogel

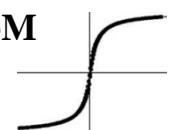


field  
Applicator



Design and  
characterization  
of field  
applicators

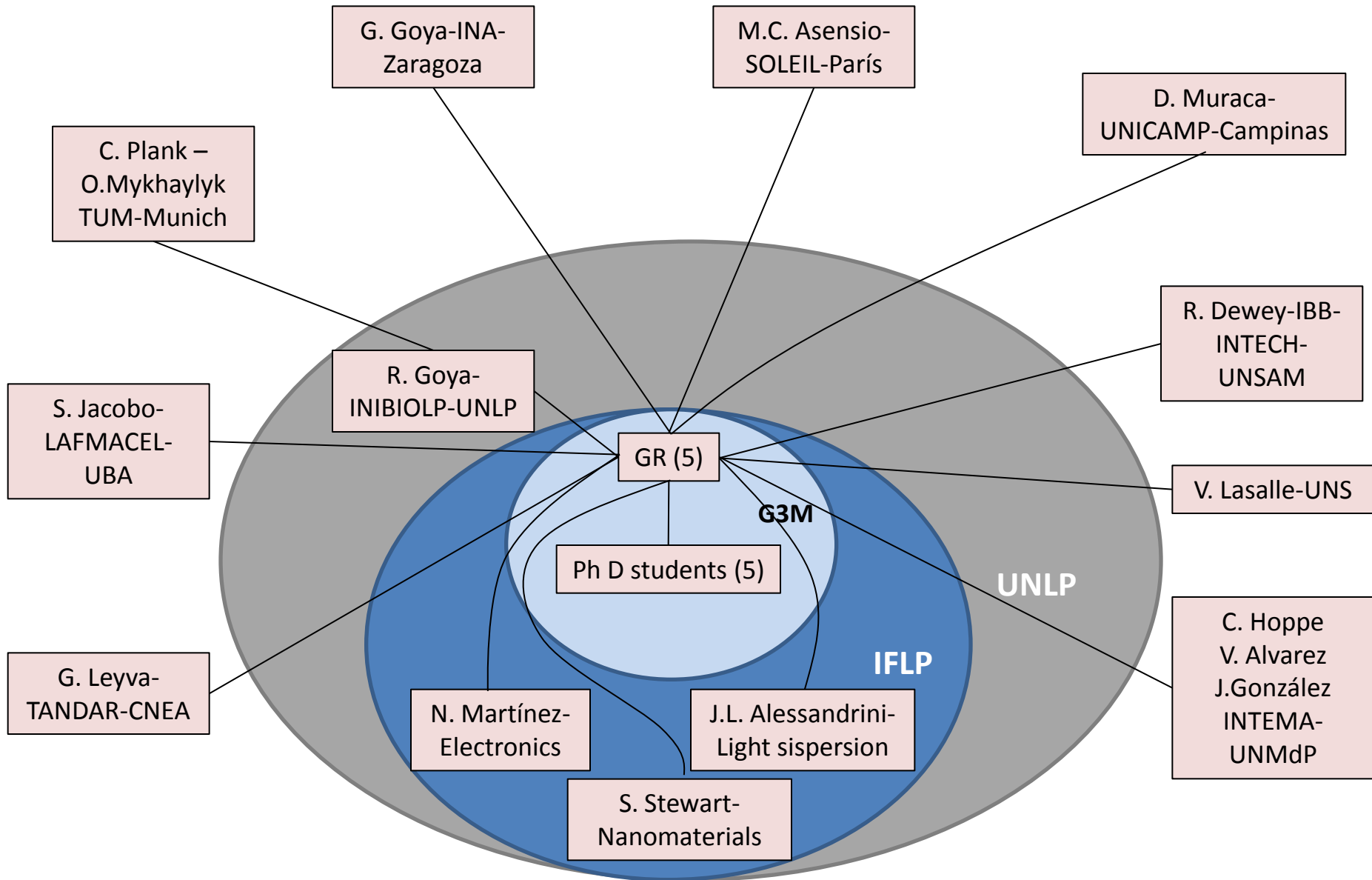
**G3M**



Magnetic  
characterization  
of dry and  
hydrated FG



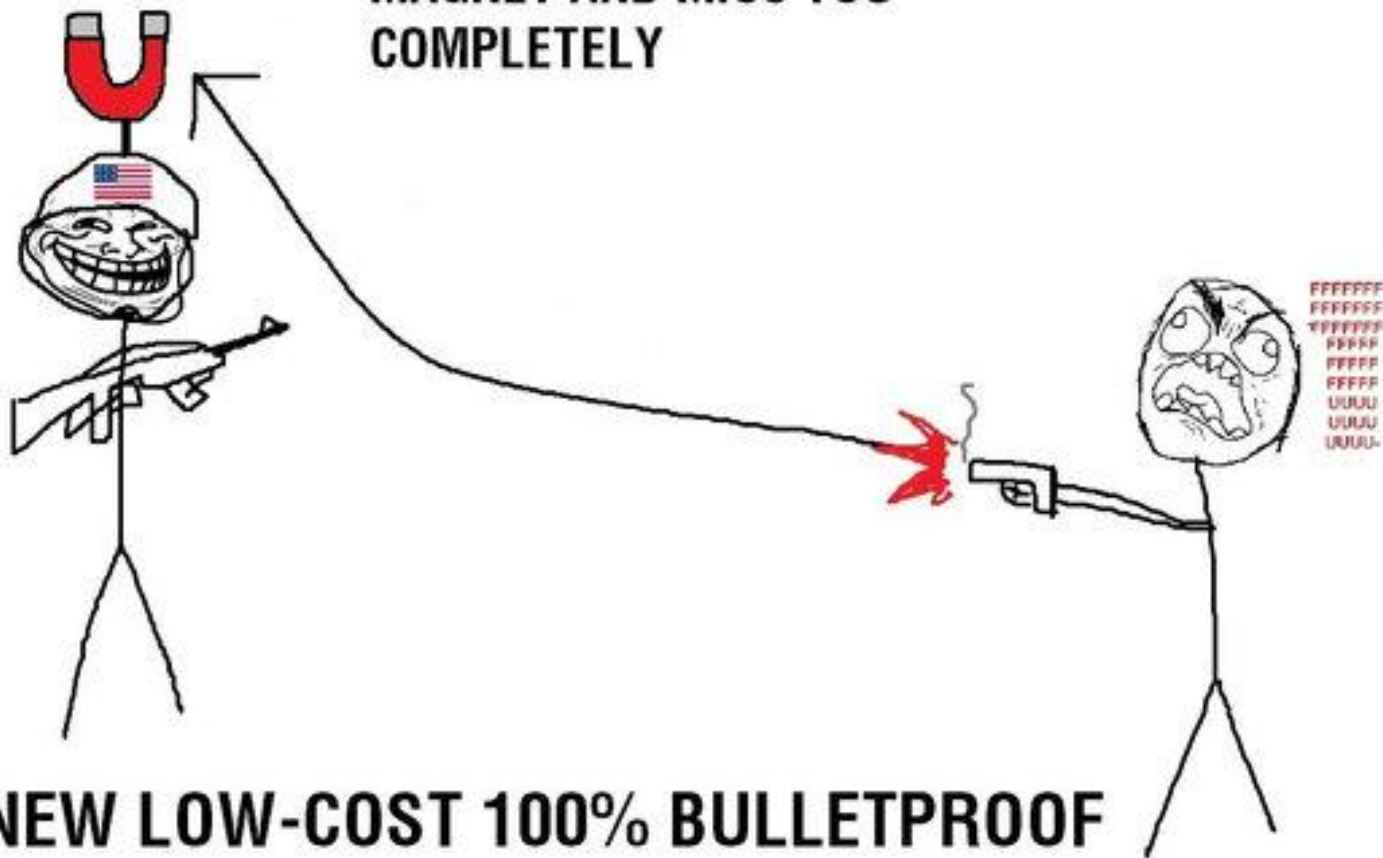
# Collaborations scheme





Mi nane is Gofman. G as in Gauss, O as in Oersted, F as in Ferrita, M as in magnetization, A as in Ampere and N as for the number of moments per volume unit!!

**BULLETS GET ATTRACTED TO THE  
MAGNET AND MISS YOU  
COMPLETELY**



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ARMOR/HELMET**

**HOW HAS THE ARMY NOT THOUGHT OF  
THIS YET?**

**DOLORES?**

**PAIN?**



**GENIOL**

QUITA EL DOLOR

REMOVES PAIN



21st Century Childhood Pranks.



*It does exist!!*